



US011001853B2

(12) **United States Patent**
Sayre et al.

(10) **Patent No.:** US 11,001,853 B2
(45) **Date of Patent:** *May 11, 2021

(54) **CARBON FIXATION SYSTEMS IN PLANTS AND ALGAE**(71) Applicant: **NMC, INC.**, Los Alamos, NM (US)(72) Inventors: **Richard Thomas Sayre**, Los Alamos, NM (US); **Somya S. Subramanian**, Los Alamos, NM (US); **Natalia Friedland**, Palo Alto, CA (US)(73) Assignee: **NMC, INC.**, Los Alamos, NM (US)

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

This patent is subject to a terminal disclaimer.

(21) Appl. No.: **16/917,028**(22) Filed: **Jun. 30, 2020**(65) **Prior Publication Data**

US 2020/0347397 A1 Nov. 5, 2020

Related U.S. Application Data

(63) Continuation of application No. 16/358,331, filed on Mar. 19, 2019, now Pat. No. 10,696,977, which is a continuation of application No. 15/411,854, filed on Jan. 20, 2017, now Pat. No. 10,233,458, which is a continuation of application No. PCT/US2015/041617, filed on Jul. 22, 2015.

(60) Provisional application No. 62/027,354, filed on Jul. 22, 2014.

(51) **Int. Cl.**

C12N 15/82	(2006.01)
C12P 1/00	(2006.01)
C12N 1/12	(2006.01)
C07K 14/415	(2006.01)

(52) **U.S. Cl.**

CPC	C12N 15/8269 (2013.01); C07K 14/415 (2013.01); C12N 1/12 (2013.01); C12N 15/8261 (2013.01); C12P 1/00 (2013.01); Y02A 40/146 (2018.01)
-----------	--

(58) **Field of Classification Search**

CPC	C12N 15/8269
See application file for complete search history.	

(56) **References Cited**

U.S. PATENT DOCUMENTS

4,554,101 A	11/1985	Hopp
5,164,316 A	11/1992	McPherson et al.
5,196,525 A	3/1993	McPherson et al.
5,322,938 A	6/1994	McPherson et al.
5,352,605 A	10/1994	Fraley et al.
5,359,142 A	10/1994	McPherson et al.
5,424,200 A	6/1995	McPherson et al.
5,510,474 A	4/1996	Quail et al.
5,589,583 A	12/1996	Klee et al.
5,593,874 A	1/1997	Brown et al.
5,599,686 A	2/1997	Defeo-Jones et al.

5,641,876 A	6/1997	McElroy et al.
5,659,122 A	8/1997	Austin
6,784,340 B1	8/2004	Aoyama et al.
6,989,265 B2	1/2006	Blattner et al.
7,053,205 B1	5/2006	Verdaguer et al.
7,303,906 B2	12/2007	Blattner et al.
8,039,243 B2	10/2011	Blattner
8,043,842 B2	10/2011	Blattner et al.
8,119,365 B2	2/2012	Blattner et al.
8,178,339 B2	5/2012	Campbell et al.
10,233,458 B2 *	3/2019	Sayre
10,696,977 B2	6/2020	Sayre et al.
10,696,979 B2 *	6/2020	Christensen
2011/0256605 A1	10/2011	Liphardt et al.
2012/0219994 A1	8/2012	Blattner et al.
2013/0007916 A1	1/2013	Spalding
2019/0203222 A1	7/2019	Sayre et al.

FOREIGN PATENT DOCUMENTS

EP	0507698	10/1992
EP	0633317	1/1995
EP	1483367	5/2010
WO	84/02913	8/1984

(Continued)

OTHER PUBLICATIONS

Ainley, W. Michael, et al., "Regulatable endogenous production of cytokinins up to 'toxic' levels in transgenic plants and plant tissues", Plant Molecular Biology, vol. 22, 1993, 13-23.

Allen, Doug K., et al., "Carbon and Nitrogen Provisions Alter the Metabolic Flux in Developing Soybean Embryos", Plant Physiology, vol. 161, 2013, 1458-1475.

Allen, Doug K., et al., "Comparing Photosynthetic and Photovoltaic Efficiencies and Recognizing the Potential for Improvement", Phytochemistry, vol. 68, 2007, 2197-2210.

(Continued)

Primary Examiner — Russell Kallis(74) *Attorney, Agent, or Firm* — Peacock Law P.C.; Janeen Vilven(57) **ABSTRACT**

Provided are heterologous nucleic acid constructs, vectors and methods for elevating cyclic electron transfer activity, improving carbon concentration, and enhancing carbon fixation in C3 and C4 plants, and algae, and producing biomass or other products from C3 or C4 plants, and algae, selected from among, for example, starches, oils, fatty acids, lipids, cellulose or other carbohydrates, alcohols, sugars, nutraceuticals, pharmaceuticals, fragrance and flavoring compounds, and organic acids, as well as transgenic plants produced thereby. These methods and transgenic plants and algae encompass the expression, or overexpression, of various combinations of genes that improve carbon concentrating systems in plants and algae, such as bicarbonate transport proteins, carbonic anhydrase, light driven proton pump, cyclic electron flow regulators, etc.

20 Claims, 13 Drawing Sheets

Specification includes a Sequence Listing.

(56)

References Cited

FOREIGN PATENT DOCUMENTS

WO	87/007644	12/1987
WO	95/006742	3/1995
WO	96/06932	3/1996
WO	97/48819	12/1997
WO	2004/053135	6/2004
WO	07/098042	8/2007
WO	2012/101118	8/2012
WO	2012/125737	9/2012
WO	2017/218959	12/2017

OTHER PUBLICATIONS

- Allen, Doug K., et al., "Isotope labelling of Rubisco subunits provides in vivo information on subcellular biosynthesis and exchange of amino acids between compartments", *Plant, Cell and Environment*, Vo. 35, 2012, 1232-1244.
- Allen, Doug K., et al., "Metabolic flux analysis in plants: coping with complexity", *Plant, Cell and Environment*, vol. 32, 2009, 1241-1257.
- Allen, Doug K., et al., "Quantification of Peptide m/z Distributions from 13C-Labeled Cultures with High-Resolution Mass Spectrometry", *Anal. Chem.*, vol. 86, 2014, 1894-1901.
- Allen, Doug K., et al., "The role of light in soybean seed filling metabolism", *The Plant Journal*, vol. 58, 2009, 220-234.
- Alicic, Jean , "Cyclic electron flow around photosystem I in unicellular green algae", *Photosynth Res*, vol. 106, 2010, 47-56.
- Altschul, Stephen F., et al., "Basic Local Alignment Search Tool", *J. Mol. Biol.*, vol. 215, 1990, 403-410.
- Altschul, Stephen F., et al., "Gapped Blast and PSI-BLAST: a new generation of protein database search programs", *Nucleic Acids Research*, vol. 25, No. 17, 1997, 3389-3402.
- Amunts, Alexey , et al., "The structure of a plant photosystem I supercomplex at 3.4A resolution", *Nature*, vol. 447, 2007, 58-63.
- Arrivault, Stephanie , "Use of reverse-phase liquid chromatography, linked to tandem mass spectrometry, to profile the Calvin cycle and other metabolic intermediates in *Arabidopsis* rosettes at different carbon dioxide concentrations", *The Plant Journal*, vol. 59, 2009, 824-839.
- Avsian-Kretchmer, Orna , et al., "The Salt-Stress Signal Transduction Pathway That Activates the gpx1 Promoter Is Mediated by Intracellular H2O2, Different from the Pathway Induced by Extracellular H2O2", *Plant Physiology*, vol. 135, 2004, 1685-1696.
- Baumann, Kim , et al., "The DNA Binding Site of the Dof Protein NtBBF1 Is Essential for Tissue-Specific and Auxin-Regulated Expression of the roIB Oncogene in Plants", *The Plant Cell*, vol. 11, 1999, 323-333.
- Beja, Oded , et al., "Bacterial Rhodopsin: Evidence for a New Type of Phototrophy in the Sea", *Science*, vol. 289, 2000, 1902-1906.
- Benfey, Philip N., et al., "The CaMV 35S enhancer contains at least two domains which can confer different developmental and tissuespecific expression patterns", *The EMBO Journal*, vol. 8, No. 8, 1989, 2195-2202.
- Bihmidine, Saadia , et al., "Regulation of assimilate import into sink organs: update on molecular drivers of sink strength", *Frontiers in Plant Science*, vol. 4, Issue 177, 2013, 1-15.
- Blanco, Nicolas E., et al., "Expression of the Minor Isoform Pea Ferredoxin in Tabacco Alters Photosynthetic Electron Partitioning and Enhances Cyclic Eletron Flow", *Plant Physiology*, vol. 161, 2013, 866-879.
- Blankenship, Robert E., "Comparing Photosynthetic and Photovoltaic Efficiencies and Recognizing the Potential for Improvement", *Science*, vol. 332, 2011, 805-809.
- Blázquez, Miguel A., et al., "Gibberellins Promote Flowering of *Arabidopsis* by Activating the Leafy Promoter", *The Plant Cell*, vol. 10, 1998, 791-800.
- Blume, Beatrix , et al., "Expression of ACC oxidase promoter-GUS fusions in tomato and Nicotiana plumbaginifolia regulated by developmental and environmental stimuli", *The Plant Journal*, vol. 12, No. 4, 1997, 731-746.
- Breyton, Cecile , et al., "Redox Modulation of Cyclic Electron Flow around Photosystem I in C3 Plants", *Biochemistry*, vol. 45, 2006, 13465-13475.
- Buchel, Annemarie , et al., "Mutation of GT-1 binding sites in the Pr-1A promoter influences the level of inducible gene expression in vivo", *Plant Molecular Biology*, vol. 40, No. 3, 1999, 387-396.
- Busch, Karin B., et al., "Dynamics of bioenergetic microcompartments", *Biol. Chem.*, vol. 394, No. 2, 2013, 163-188.
- Busk, Peter Kamp, et al., "Regulatory elements in vivo in the promoter of the abscisic acid responsive gene rab17 from maize", *The Plant Journal*, vol. 11, No. 6, 1997, 1285-1295.
- Callis, Judy , et al., "Introns increase gene expression in cultured maize cells", *Genes & Development*, vol. 1, 1987, 1183-1200.
- Cao, Yi , et al., "Relationship of Proton Release at the Extracellular Surface to Deprotonation of the Schiff Base in the Bacteriorhodopsin Photocycle", *Biophysical Journal*, vol. 68, 1995, 1518-1530.
- Cardol, Pierre , et al., "Regulation of electron transport in microalgae", *Biochimica et Biophysica Acta*, vol. 1807, 2011, 912-918.
- Cardon, Guillermo H., et al., "Functional analysis of the *Arabidopsis thaliana* SBP-box gene SPL3: a novel gene involved in the floral transition", *The Plant Journal*, vol. 12, No. 2, 1997, 367-377.
- Carrillo, Humberto , et al., "The Multiple Sequence Alignment Problem in Biology", *SIAM Journal on Applied Mathematics*, vol. 48, No. 5, 1988, 1073-1082.
- Carrington, James C., et al., "Cap-Independent Enhancement of Translation by a Plant Potyvirus 5' Nontranslated Region", *Journal of Virology*, vol. 64, No 4, 1990, 1590-1597.
- Chaubet-Gigot, Nicole , et al., "Tissue-dependent enhancement of transgene expression by introns of replacement histone H3 genes of *Arabidopsis*", *Plant Molecular Biology*, vol. 45, 2001, 17-30.
- Chen, Wenqiong , et al., "The promoter of a H2O2-inducible, *Arabidopsis* glutathione S-transferase gene contains closely linked OBF- and OBP1-binding sites", *The Plant Journal*, vol. 10, No. 6, 1996, 955-966.
- Choi, Jungik , et al., "Tandem massspectrometry:Anovelapproach ormetabolicfluxanalysis", *Metabolic Engineering*, vol. 13, 2011, 225-233.
- Clancy, Maureen , et al., "Splicing of the Maize Sh1 First Intron Is Essential for Enhancement of Gene Expression, and a T-Rich Motif Increases Expression without Affecting Splicing", *Plant Physiol.* vol. 130, 2002, 918-929.
- Claverie, Jean-Michel , "Information Enhancement Methods for Large Scale Sequence Analysis", *Computers Che.*, vol. 17, No. 2, 1993, 191-201.
- Dalcorso, Giovanni , et al., "A Complex Containing PGRL1 and PGR5 Is Involved in the Switch Between Linear and Cyclic Electron Flow in *Arabidopsis*", *Cell* 132, 2008, 273-285.
- Dalcorso, Giovanni , et al., "A Complex Containing PGRL1 and PGR5 Is Involved in the Switch between Linear and Cyclic Electron Flow in *Arabidopsis*", *Cell*, vol. 132, 2008, 273-285.
- Datla, Raju S.S., et al., "Improved high-level constitutive foreign gene expression in plants using an AMV RNA4 untranslated leader sequence", *Plant Science*, vol. 94, 1993, 139-149.
- De Veylder, Lieven , et al., "Herbicide Safener-Inducible Gene Expression in *Arabidopsis thaliana*", *Plant Cell Physiol.*, vol. 38, No. 5, 1997, 568-577.
- Dioumaev, Andrei K., et al., "Proton Transfers in the Photochemical Reaction Cycle of Proteorhodopsin", *Biochemistry*, vol. 41, 5348-5358, 2002.
- Duanmu, Deqiang , et al., "Knockdown of limiting-CO2-induced gene HL3 decreases HCO3 transport and photosynthetic Ci affinity in *Chlamydomonas reinhardtii*", *PNAS*, vol. 106, No. 14, 2009, 5990-5995.
- Duckwall, Casey Scott, et al., "Mapping cancer cell metabolism with 13C flux analysis: Recent progress and future challenges", *Journal of Carcinogenesis*, vol. 12, No. 13, 2013, 1-7.
- Egnatchik, R. A., et al., "Palmitate-induced activation of mitochondrial metabolism promotes oxidative stress and apoptosis in H4IIEC3 rat hepatocytes", *Metabolism*, vol. 62, No. 2, 2014, 283-295.
- Elleby, Bjorn , et al., "Characterization of carbonic anhydrase from *Neisseria gonorrhoeae*", *Eur. J. Biochem*, vol. 268, 2001, 1613-1619.

(56)

References Cited**OTHER PUBLICATIONS**

- Fabre, Nicolas , et al., "Characterization and expression analysis of genes encoding a and b carbonic anhydrases in *Arabidopsis*", Plant, Cell and Environment, vol. 30, 2007, 617-629.
- Farquhar, G. D., et al., "Carbon Isotope Discrimination and Photosynthesis", Annu. Rev. Plant Physiol. Plant Mol. Biol., vol. 40, 1989, 503-537.
- Friedrich, Thomas , et al., "Proteorhodopsin is a Light-driven Proton Pump with Variable Vectoriality", J. Mol. Biol., vol. 321, 2002, 821-838.
- Froehlich John E., et al., "The role of the transmembrane domain in determining the targeting of membrane proteins to either the inner envelope or thylakoid membrane", The Plant Journal, vol. 68, 2011, 844-856.
- Furbank, Robert T., et al., "C4 rice: a challenge for plant phenomics", Functional Plant Biology, vol. 36, No. 11, 2009, 845-856.
- Wunsche, Jens N., et al., "Physiological and biochemical leaf and tree responses to crop load in apple", Tree Physiology, vol. 25, 2005, 1253-1263.
- Yamaguchi-Shinozaki, Kazuko , et al., "A Novel cis-Acting Element in an *Arabidopsis* Gene Is Involved in Responsiveness to Drought, Lowtemperature, or High-Salt Stress", The Plant Cell, vol. 6, 1994, 251-264.
- Young, Jamey D., et al., "An Elementary Metabolite Unit (EMU) Based Method of Isotopically Nonstationary Flux Analysis", Biotechnology and Bioengineering, vol. 99, No. 3, 2008, 686-699.
- Young, Jamey D., "INCA: a computational platform for isotopically non-stationary metabolic flux analysis", Bioinformatics, vol. 30, No. 9, 2014, 1333-1335.
- Young, Jamey D., et al., "Isotopomer Measurement Techniques in Metabolic Flux Analysis II: Mass Spectrometry", Methods in Molecular Biology, vol. 1083, Chapter 7, 2014, 85-108.
- Young, Jamey D., "Mapping photoautotrophic metabolism with isotopically nonstationary ^{13}C flux analysis", Metabolic Engineering, vol. 13, 2011, 656-665.
- Young, Jamey D., "Metabolic flux rewiring in mammalian cell cultures", Current Opinion in Biotechnology, vol. 24, 2013, 1108-1115.
- Zhu, Xin-Guang , et al., "C4 Rice—an Ideal Arena for Systems Biology Research", Journal of Integrative Plant Biology, vol. 52, Issue 8, 2010, 762-770.
- Zuo, Jianru , et al., "An estrogen receptor-based transactivator XVE mediates highly inducible gene expression in transgenic plants", The Plant Journal, vol. 24, No. 2, 2000, 265-273.
- Okutani, Satoshi , "Three Maize Leaf Ferredoxin:NADPH Oxidoreductases Vary in Subchloroplast Location, Expression, and Interaction with Ferredoxin", Plant Physiology, vol. 139, 2005, 1451-1459.
- Outchkourov, N. S., et al., "The promoter-terminator of chrysanthemum rbcS1 directs very high expression levels in plants", Planta, vol. 216, 2003, 1003-1012.
- Parry, Martin A.J., et al., "Rubisco activity and regulation as targets for crop improvement", Journal of Experimental Botany, vol. 64, No. 3, 2013, 717-730.
- Paul, Matthew J., et al., "Sink regulation of photosynthesis", Journal of Experimental Botany, vol. 52, No. 360, 2001, 1383-1400.
- Peltier, Gilles , "Auxiliary electron transport pathways in chloroplasts of microalgae", Photosynth Res, vol. 106, 2010, 19-31.
- Peng, Lianwei , et al., "Supercomplex Formation with Photosystem I Is Required for the Stabilization of the Chloroplast NADPH Dehydrogenase-Like Complex in *Arabidopsis*", Plant Physiology, vol. 155, 2011, 1629-1639.
- Perrine, Zee , et al., "Optimization of photosynthetic light energy utilization by microalgae", Algal Research, vol. 1, 2012, 134-142.
- Peterhansel, Christoph , et al., "Photorespiratory bypasses: how can they work?", Journal of Experimental Botany, vol. 64, No. 3, 2013, 709-715.
- Price, G. Dean, et al., "The cyanobacterial CCM as a source of genes for improving photosynthetic CO₂ fixation in crop species", Journal of Experimental Botany, vol. 64, No. 3, 2013, 753-768.
- Reeck, Gerald R., et al., "'Homology' in Proteins and Nucleic Acids: A Terminology Muddle and a Way out of It", Cell, vol. 50, 1987, 667.
- Reiser, Leonore , et al., "The BELL7 Gene Encodes a Homeodomain Protein Involved in Pattern Formation in the *Arabidopsis* Ovule Primordium", Cell, vol. 83, 1995, 735-742.
- Ringli, Christoph , et al., "Specific interaction of the tomato bZIP transcription factor VSF-1 with a non-palindromic DNA sequence that controls vascular gene expression", Plant Molecular Biology, vol. 37, 1998, 977-988.
- Romisch-Margl, Werner , et al., " $^{13}\text{CO}_2$ as a universal metabolic tracer in isotopologue perturbation experiments", Phytochemistry, vol. 68, 2007, 2273-2289.
- Roslan, Hairul A., et al., "Characterization of the ethanol-inducible alc geneexpression system in *Arabidopsis thaliana*", The Plant Journal, vol. 28, No. 2, 2001, 225-235.
- Sage, Tammy L., et al., "The Functional Anatomy of Rice Leaves: Implications for Refixation of Photorespiratory CO₂ and Efforts to Engineer C4 Photosynthesis into Rice", Plant Cell Physiol. vol. 50, No. 4, 2009, 756-772.
- Sage, Rowan F., "Variation in the Kcat of Rubisco in C3 and C4 plants and some implications for photosynthetic performance at high and low temperature", Journal of Experimental Botany, vol. 53, No. 369, 2002, 609-620.
- Sakai, Tatsuya , et al., "Analysis of the Promoter of the Auxin-Inducible Gene, parC, of Tobacco", Plant Cell Physiol., vol. 37, No. 7, 1996, 906-913.
- Sakamoto, Masahiro , et al., "Structure and Characterization of a Gene for Light-Harvesting Chi a/b Binding Protein from Rice", Plant Cell Physiol., vol. 32, No. 3, 1991, 385-393.
- Samac, Deborah A., et al., "A comparison of constitutive promoters for expression of transgenes in alfalfa (*Medicago sativa*)", Transgenic Research, vol. 13, 2004, 349-361.
- Sanger, Margaret , et al., "Characteristics of a strong promoter from figwort mosaic virus: comparison with the analogous 35S promoter from cauliflower mosaic virus and the regulated mannopine synthase promoter", Plant Molecular Biology, vol. 14, 1990, 433-443.
- Schaffner, Anton R., et al., "Maize rbcS Promoter Activity Depends on Sequence Elements Not Found in Dicot rbcS Promoters", The Plant Cell, vol. 3, 1991, 997-1012.
- Sekiyama, Yasuyo , et al., "Towards dynamic metabolic network measurements by multi-dimensional NMR-based fluxomics", Phytochemistry, vol. 68, 2007, 2320-2329.
- Shastri, Avantika A., et al., "A transient isotopic labeling methodology for ^{13}C metabolic flux analysis of photoautotrophic microorganisms", Phytochemistry, vol. 68, 2007, 2302-2312.
- Sheen, Jen , "Ca²⁺-dependent protein kinases and stress signal transduction in plants", Science, vol. 274, No. 5294, 1996, 1900-1902.
- Shi, Rebecca , et al., "Engineering *Oryza sativa* to Express the Proteorhodopsin Photosystem", [http://openwetware.org/wiki/20.109\(F12\):Mod3_OrangeTR_Pre-proposal](http://openwetware.org/wiki/20.109(F12):Mod3_OrangeTR_Pre-proposal), 2012, 1-4.
- Shi, Lifang , et al., "Gibberellin and abscisic acid regulate GAST1 expression at the level of transcription", Plant Molecular Biology, vol. 38, 1998, 1053-1060.
- Shikanai, Toshiharu , "Central role of cyclic electron transport around photosystem I in the regulation of photosynthesis", Current Opinion in Biotechnology, vol. 26, 2014, 25-30.
- Si, Li-Zhen , et al., "Isolation of a 1 195 bp 5-Flanking Region of Rice Cytosolic Fructose-1, 6-bisphosphatase and Analysis of Its Expression in Transgenic Rice", Acta Botanica Sinica, vol. 3, 2003, 359-364.
- Siebertz, Barbara , et al., "cis-Analysis of the Wound-Inducible Promoter wun7 in Transgenic Tobacco Plants and Histochemical Localization of Its Expression", The Plant Cell, vol. 1, 1989, 961-968.
- Sinclair, Thomas R., "Historical Changes in Harvest Index and Crop Nitrogen Accumulation", Crop Science, vol. 38, No. 3, 1998, 638-643.
- Slewinski, Thomas L., et al., "Current perspectives on the regulation of whole-plant carbohydrate partitioning", Plant Science, vol. 178, 2010, 341-349.

(56)

References Cited**OTHER PUBLICATIONS**

- Sonnewald, U. , "Manipulation of sink-source relations in transgenic plants", *Plant, Cell and Environment*, vol. 17, 1994, 649-658.
- Sonnewald, Uwe , et al., "Molecular Approaches to Sink-Source Interactions", *Plant Physiol.*, vol. 99, 1992, 1267-1270.
- Srouf, Orr , et al., "Fluxomers: a new approach for ^{13}C metabolic flux analysis", *BMC Systems Biology*, vol. 5, No. 129, 2011, 1-15.
- Stange, Claudia , et al., "Phosphorylation of nuclear proteins directs binding to salicylic acid-responsive elements", *The Plant Journal*, vol. 11, No. 6, 1997, 1315-1324.
- Streit, Wolfgang R., et al., "A Biotin-Regulated Locus, *bioS*, in a Possible Survival Operon of *Rhizobium meliloti*", *MPMI* vol. 10, No. 7, 1997, 933-937.
- Subramanian, Sowmya , "Comparative energetics and kinetics of autotrophic lipid and starch metabolism in chlorophytic microalgae: implications for biomass and biofuel production", *Biotechnology for Biofuels*, vol. 6, No. 150, 2013, 1-12.
- Suorsa, Marjaana , et al., "PGR5-PGRL1-Dependent Cyclic Electron Transport Modulations Linear Electron Transport Rate in *Arabidopsis thaliana*", *Molecular Plant*, vol. 9, 2016, 271-288.
- Sweetlove, L. J., et al., "Source metabolism dominates the control of source to sink carbon flux in tuberizing potato plants throughout the diurnal cycle and under a range of environmental conditions", *Plant, Cell and Environment*, vol. 23, 2000, 523-529.
- Szecowka, Marek , et al., "Metabolic Fluxes in an Illuminated *Arabidopsis* Rosette", *The Plant Cell*, vol. 25, 2013, 694-714.
- Takahashi, Hiroko , et al., "Cyclic electron flow is redox-controlled but independent of state transition", *Nature Communication*, vol. 4, No. 1954, 2013, 1-8.
- Van Der Kop, Dianne A.M., et al., "Selection of *Arabidopsis* mutants overexpressing genes driven by the promoter of an auxin-inducible glutathione S-transferase gene", *Plant Molecular Biology*, vol. 39, 1999, 970-990.
- Victorio, Reynaldo G., et al., "Growth, Partitioning, and Harvest Index of Tuber-Bearing *Solanum* Genotypes Grown in Two Contrasting Peruvian Environments", *Plant Physiol.*, vol. 82, 1986, 103-108.
- Vos, J. , "The nitrogen response of potato (*Solanum tuberosum* L.) in the field: nitrogen uptake and yield, harvest index and nitrogen concentration.", *Potato Research*, vol. 40, 1997, 237-248.
- Walter, Jessica M., et al., "Light-powering *Escherichia coli* with proteorhodopsin", *PNAS*, vol. 104, No. 7, 2007, 2408-2412.
- Walter, Jessica M., et al., "Potential of light-harvesting proton pumps for bioenergy applications", *Current Opinion in Biotechnology*, vol. 21, 2010, 265-270.
- Wang, Yingjun , et al., "Carbon dioxide concentrating mechanism in *Chlamydomonas reinhardtii*: inorganic carbon transport and CO₂ recapture", *Photosynth Res*, vol. 109, 2011, 115-122.
- Weber, Andreas PM, et al., "Plastid transport and metabolism of C3 and C4 plants—comparative analysis and possible biotechnological exploitation", *Current Opinion in Plant Biology*, vol. 13, 2010, 257-265.
- Willmott, Ruth L., et al., "DNase1 footprints suggest the involvement of at least three types of transcription factors in the regulation of alpha-Amy2/A by gibberellin", *Plant Molecular Biology*, vol. 38, 1998, 817-825.
- Wootton, John C., et al., "Statistics of Local Complexity in Amino Acid Sequences and Sequence Databases", *Computers Chem.* vol. 17, No. 2, 1993, 149-163.
- Gatz, C. , et al., "Chemical Control of Gene Expression", *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, vol. 48, 1997, 89-108.
- Goldschmidt, Eliezer E., et al., "Regulation of Photosynthesis by End-Product Accumulation in Leaves of Plants Storing Starch, Sucrose, and Hexose Sugars", *Plant Physiol.*, vol. 99, 1992, 1443-1448.
- Govindjee, Rajni , et al., "Arginine-82 Regulates the PKa of the Group Responsible for the Light-Driven Proton Release in Bacteriorhodopsin", *Biophysical Journal*, vol. 71, 1996, 1011-1023.
- Govindjee, Rajni , et al., "Mutation of a Surface Residue, Lysine-129, Reverses the Order of Proton Release and Uptake in Bacteriorhodopsin; Guanidine Hydrochloride Restores It", *Biophysical Journal*, vol. 72, 1997, 886-898.
- Govindjee, Rajni , et al., "The Quantum Efficiency of Proton Pumping by the Purple Membrane of *Halobacterium Halobium*", *Biophys. J.*, vol. 30, 1980, 231-242.
- Guevara-Garcia, Arturo , et al., "A 42 bp fragment of the pmas10 promoter containing an ocs-like element confers a developmental, wound- and chemically inducible expression pattern", *Plant Molecular Biology*, vol. 38, 1998, 743-753.
- Hanke, Guy Thomas, et al., "Multiple iso-proteins of FNR in *Arabidopsis* : evidence for different contributions to chloroplast function and nitrogen assimilation", *Plant, Cell and Environment*, vol. 28, 2005, 1146-1157.
- Harpster, Mark H., et al., "Relative strengths of the 35S califlower mosaic virus, 1', 2', and nopaline synthase promoters in transformed tobacco sugarbeet and oilseed rape callus tissue", *Mol Gen Genet*, vol. 212, 1988, 182-190.
- Hausler, Rainer E., "Overexpression of C4-cycle enzymes in transgenic C3 plants: a biotechnological approach to improve C3-photosynthesis", *Journal of Experimental Botany*, vol. 53, No. 369, 2002, 591-607.
- Hay, R. K. M., et al., "Variation in the harvest index of tropical maize: evaluation of recent evidence from Mexico and Malawi", *Ann. appl. Biol.*, vol. 138, 2001, 103-109.
- Henikoff, Steven , et al., "Amino acid substitution matrices from protein blocks", *Proc. Natl. Acad. Sci. USA*, vol. 89, 1992, 10915-10919.
- Henkes, Stefan , et al., "A Small Decrease of Plastid Transketolase Activity in Antisense Tobacco Transformants Has Dramatic Effects on Photosynthesis and Phenylpropanoid Metabolism", *The Plant Cell*, vol. 13, 2001, 535-551.
- Hertle, Alexander P., et al., "PGRL1 Is the Elusive Ferredoxin-Plastoquinone Reductase in Photosynthetic Cyclic Electron Flow", *Molecular Cell*, vol. 49, 2013, 511-523.
- Holtorf, Sonke , et al., "Comparison of different constitutive and inducible promoters for the overexpression of transgenes in *Arabidopsis thaliana*", *Plant Molecular Biology*, vol. 29, 1995, 637-646.
- Huege, Jan , et al., "GC-EI-TOF-MS analysis of in vivo carbon-partitioning into soluble metabolite pools of higher plants by monitoring isotope dilution after $^{13}\text{CO}_2$ labelling", *Phytochemistry*, vol. 68, 2007, 2258-2272.
- Ihemere, Uzoma , "Genetic modification of cassava for enhanced starch production", *Plant Biotechnology Journal*, vol. 4, 2006, 453-465.
- Jazmin, Lara J., "Isotopically Nonstationary ^{13}C Metabolic Flux Analysis", *Systems Metabolic Engineering: Methods and Protocols*, Methods in Molecular Biology, vol. 985, Chapter 18, 2013, 367-390.
- Jazmin, Lara J., "Isotopically Nonstationary MFA (INST-MFA) of Autotrophic Metabolism", *Methods in Molecular Biology*, vol. 1090, Chapter 12, 2014, 181-210.
- Johnson, Giles N., "Physiology of PSI cyclic electron transport in higher plants", *Biochimica et Biophysica Acta*, vol. 1807, 2011, 384-389.
- Joliot, Pierre , et al., "Regulation of cyclic and linear electron flow in higher plants", *PNAS*, vol. 108, No. 32, 2011, 13317-13322.
- Jonik, Claudia , et al., "Simultaneous boosting of source and sink capacities doubles tuber starch yield of potato plants", *Plant Biotechnology Journal*, vol. 10, 2012, 1088-1098.
- Karlin, Samuel , et al., "Applications and statistics for multiple high-scoring segments in molecular sequences", *Proc. Natl. Acad. Sci. USA*, vol. 90, 1993, 5873-5877.
- Kay, Robert , et al., "Duplication of CaMV 35S promoter sequences creates a strong enhancer for plant genes", *Science*, vol. 236, 1987, 1299-1302.
- Kelemen, Zsolt , et al., "Transformation vector based on promoter and intron sequences of a replacement histone H3 gene. A tool for high, constitutive gene expression in plants", *Transgenic Research*, vol. 11, 2002, 69-72.
- Kim, Joon YH, et al., "Improved production of biohydrogen in light-powered *Escherichia coli* by co-expression of proteorhodopsin and heterologous hydrogenase", *Microbial Cell Factories*, vol. 11, No. 2, 2012, 1-7.

(56)

References Cited**OTHER PUBLICATIONS**

- Kramer, David M., et al., "The Importance of Energy Balance in Improving Photosynthetic Productivity", *Plant Physiology*, vol. 155, 2011, 70-78.
- Kramer, David M., et al., "The Importance of Energy Balance in Improving Photosynthetic Productivity 1[W]", *Plant Physiology*, vol. 155, 2011, 70-78.
- Kuhlemeier, Cris , et al., "The Pea rbcS-3A Promoter Mediates Light Responsiveness but not Organ Specificity", *The Plant Cell*, vol. 1, 1989, 471-478.
- Kyte, Jack , "A simple method for displaying the hydrophobic character of a protein", *J. Mol. Biol.*, vol. 157, No. 1, 1982, 105-132.
- Lakatos, Melinda , et al., "The Photochemical Reaction Cycle of Proterorhodopsin at Low pH", *Biophysical Journal*, vol. 84, 2003, 3252-3256.
- Leamy, Alexandra , et al., "Modulating lipid fate controls lipotoxicity in palmitate-treated hepatic cells", *The FASEB Journal*, vol. 27, No. 1, 2013, 1.
- Leamy, Alexandra K., "Molecular mechanisms and the role of saturated fatty acids in the progression of non-alcoholic fatty liver disease", *Progress in Lipid Research*, vol. 52, 2013, 165-174.
- Lindqvist, Annika , et al., "Biochemical Properties of Purified Recombinant Human Beta-Carotene 15, 15'-Monooxygenase", *The Journal of Biological Chemistry*, vol. 277, No. 26, 2002, 23942-23948.
- Liu, Zhan-Bin , et al., "A G-Box-Binding Protein from Soybean Binds to the E1 Auxin-Response Element in the Soybean CH3 Promoter and Contains a Proline-Rich Repression Domain", *Plant Physiol.*, vol. 115, 1997, 397-407.
- Lu, Chaofu , et al., "Generation of transgenic plants of a potential oilseed crop *Camelina sativa* by Agrobacterium-mediated transformation", *Plant Cell Rep*, vol. 27, 2008, 273-278.
- Ma, Fangfang , et al., "Isotopically nonstationary 13C flux analysis of changes in *Arabidopsis thaliana* leaf metabolism due to high light acclimation", *PNAS*, vol. 111, No. 7, 2014, 16967-16972.
- Mandel, Therese , et al., "Definition of constitutive gene expression in plants: the translation initiation factor 4A gene as a model", *Plant Molecular Biology*, vol. 29, 1995, 995-1004.
- Mandy, Dominic E., et al., "Metabolic flux analysis using 13C peptide label measurements", *The Plant Journal*, vol. 77, 2014, 476-486.
- Manners, John M., et al., "The promoter of the plant defensin gene PDF1.2 from *Arabidopsis* is systemically activated by fungal pathogens and responds to methyl jasmonate but not to salicylic acid", *Plant Molecular Biology*, vol. 38, 1998, 1071-1080.
- Martinez, A. , "Proterorhodopsin photosystem gene expression enables photophosphorylation in a heterologous host", *PNAS*, vol. 104, No. 13, 2007, 5590-5595.
- Mascarenhas, Desmond , et al., "Intron-mediated enhancement of heterologous gene expression in maize", *Plant Molecular Biology*, vol. 15, 1990, 913-920.
- Masclaux-Daubresse, Celine , et al., "Exploring nitrogen remobilization for seed filling using natural variation in *Arabidopsis thaliana*", *Journal of Experimental Botany*, vol. 62, No. 6, 2011, 2131-2142.
- Masgrau, Carles , et al., "Inducible overexpression of oat arginine decarboxylase in transgenic tobacco plants", *The Plant Journal*, vol. 11, No. 3, 1997, 465-473.
- Mcatee, Allison G., et al., "Role of Chinese hamster ovary central carbon metabolism in controlling the quality of secreted biotherapeutic proteins", *Pharm. Bioprocess.*, vol. 2, No. 1, 2014, 63-74.
- Minagawa, Jun , "State transitions—The molecular remodeling of photosynthetic supercomplexes that controls energy flow in the chloroplast", *Biochimica et Biophysica Acta*, vol. 1807, 2011, 897-905.
- Miyagawa, Yoshiko , et al., "Overexpression of a cyanobacterial fructose-1,6-/sedoheptulose-1,7-bisphosphatase in tobacco enhances photosynthesis and growth", *Nature Biotechnology*, vol. 19, 2001, 965-969.
- Moroney, James V., et al., "Photorespiration and carbon concentrating mechanisms: two adaptations to high O₂, low CO₂ conditions", *Photosynth Res*, vol. 117, 2013, 121-131.
- Nakamura, Naoy , et al., "Promotion of cyclic electron transport around photosystem I during the evolution of NADP-malic enzyme-type C4 photosynthesis in the genus *Flaveria*", *New Phytologist*, vol. 199, 2013, 832-842.
- Odell, Joan T., et al., "Identification of DNA sequences required for activity of the cauliflower mosaic virus 35S promoter", *Nature*, vol. 313, No. 6005, 1985, 810-812.
- Odell, Joan T., "Seed-Specific Gene Activation Mediated by the Cre/lox Site-Specific Recombination System", *Plant Physiol.*, vol. 106, 1994, 447-458.

* cited by examiner

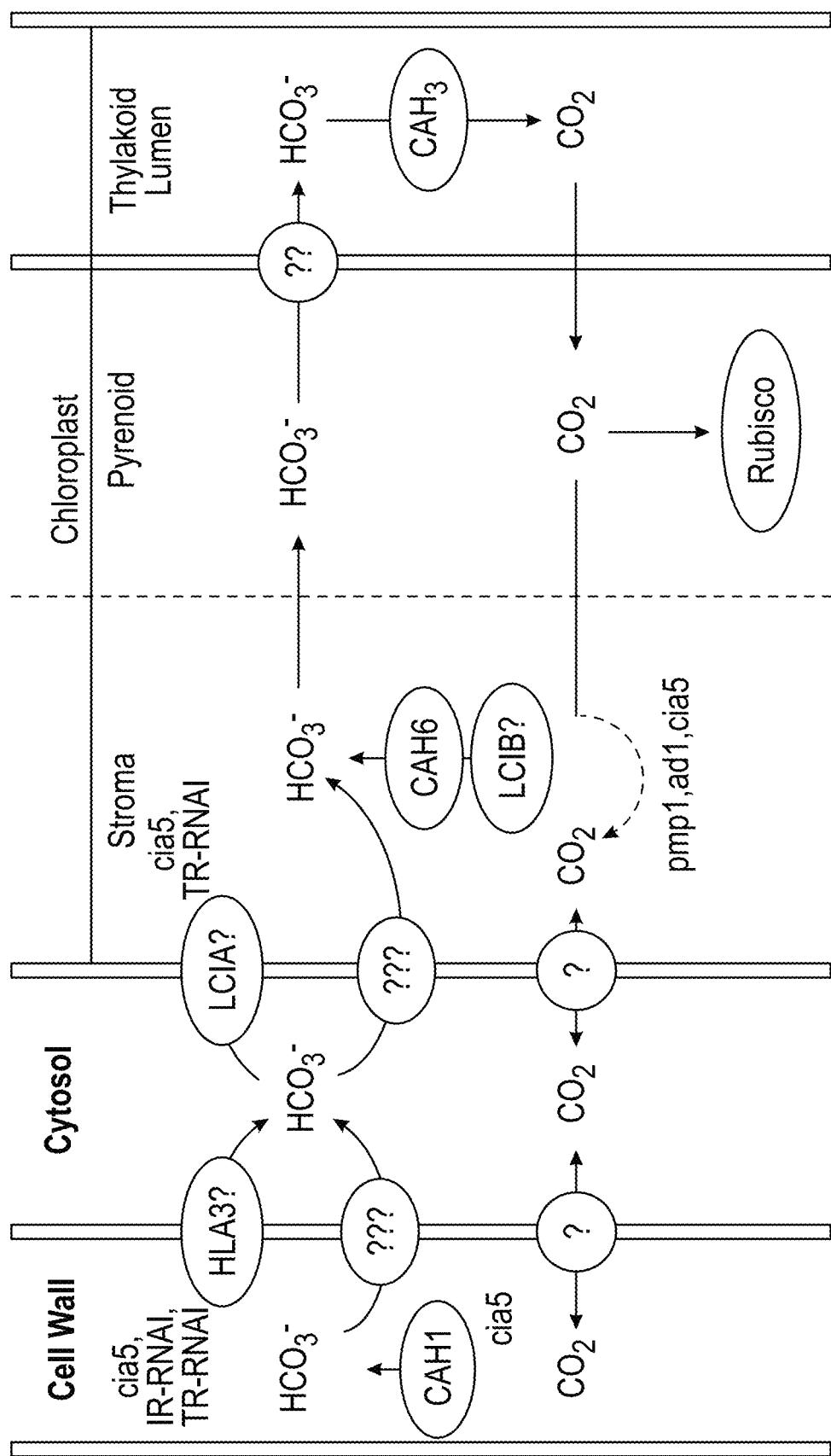
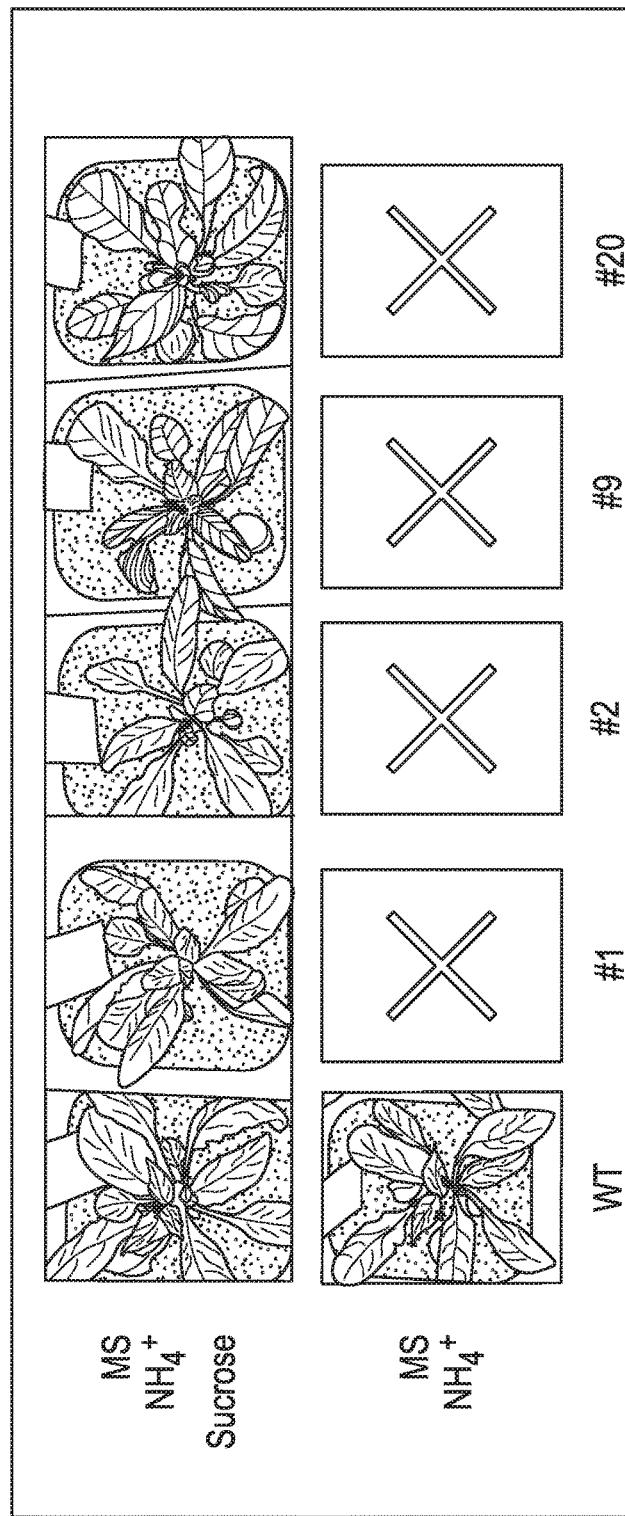
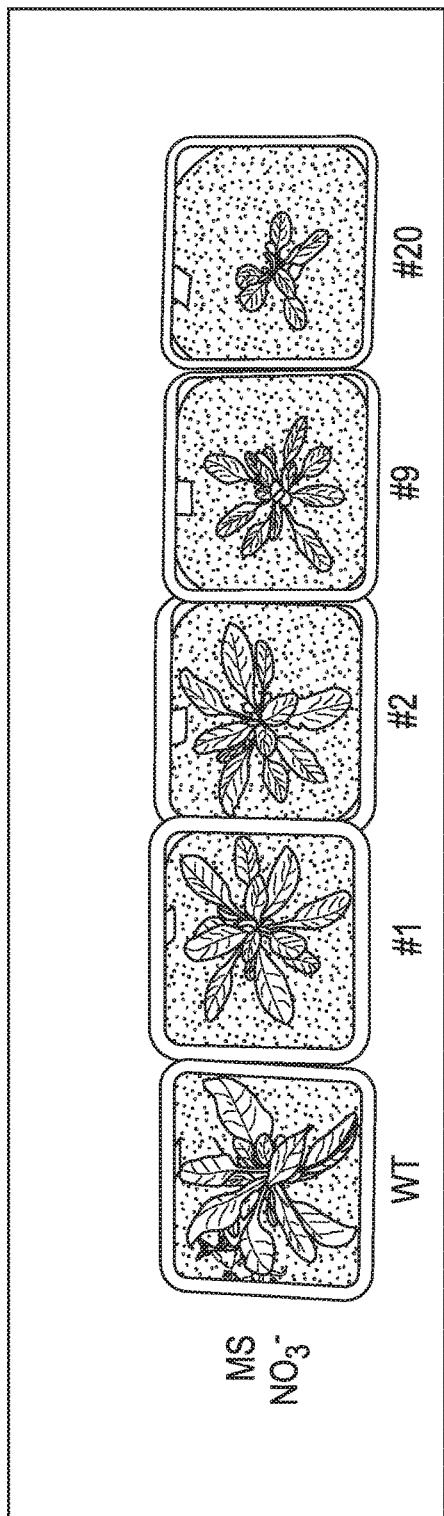


FIG. 1



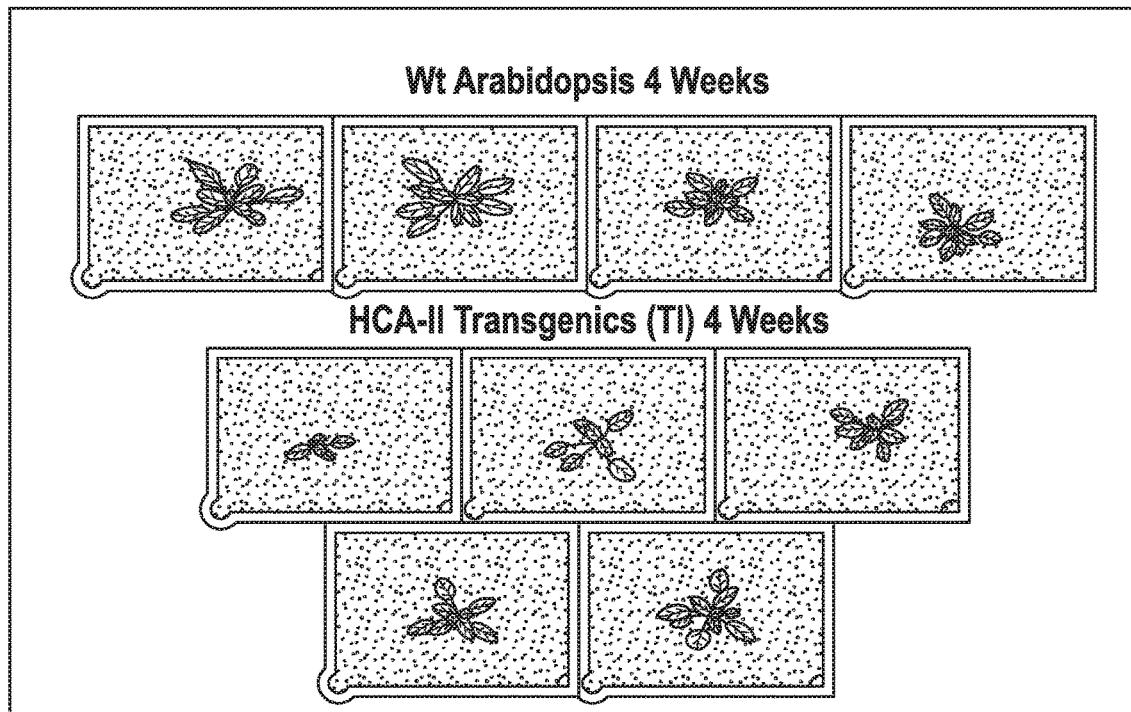


FIG. 3A

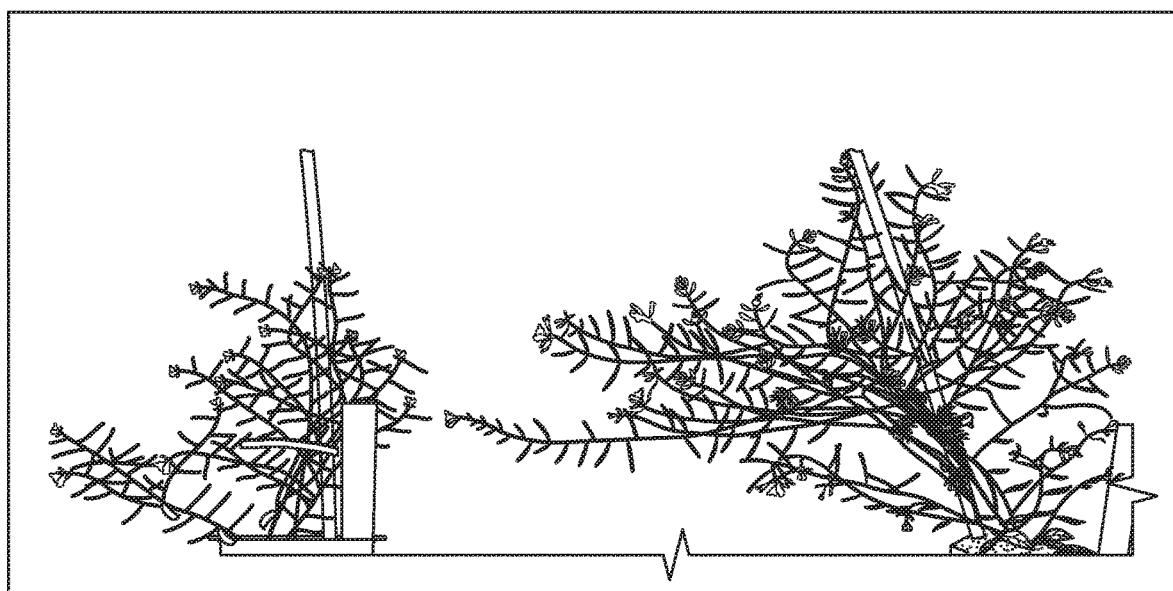
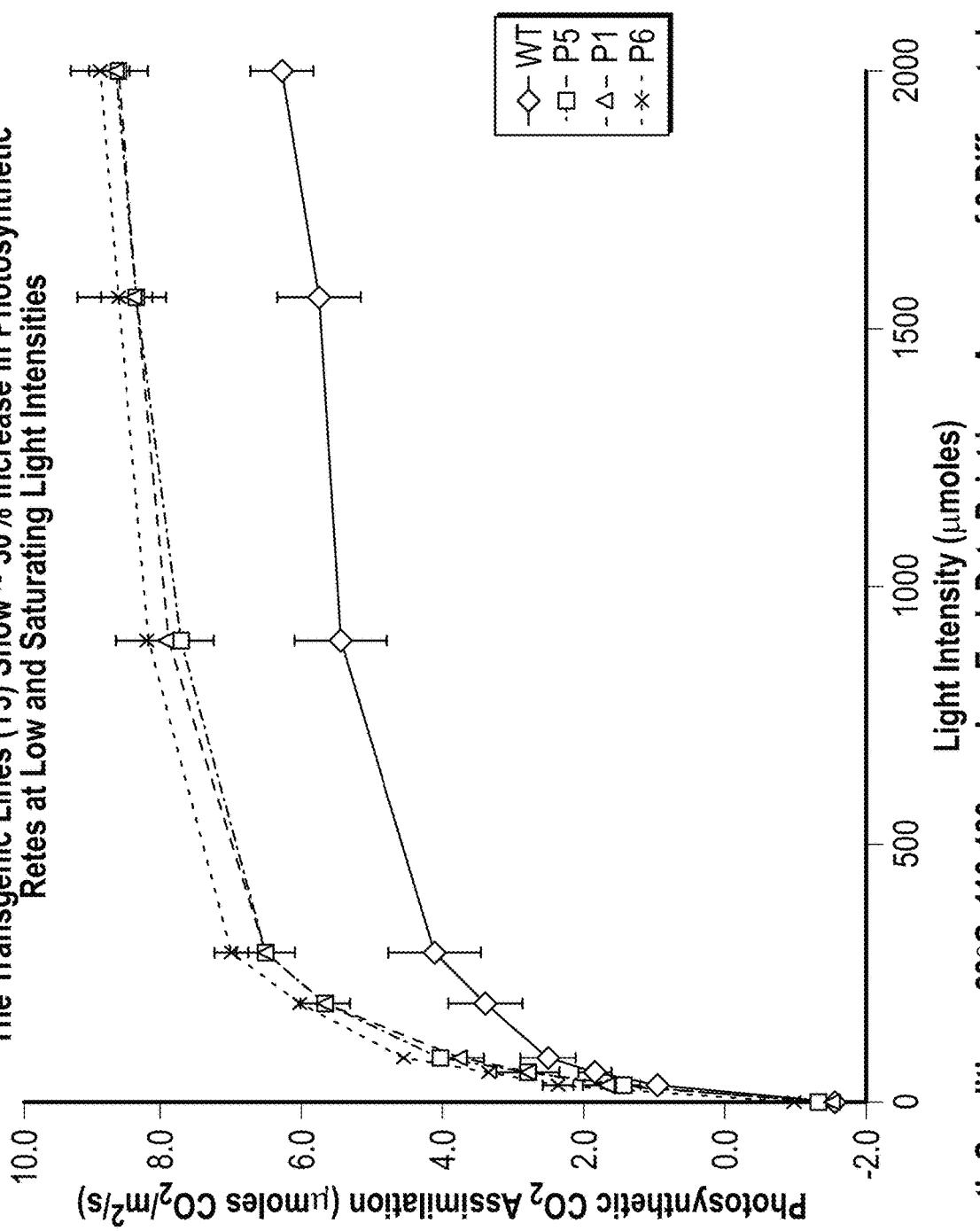


FIG. 3B

The Transgenic Lines (T3) Show ~30% Increase in Photosynthetic Rates at Low and Saturating Light Intensities



Growth Conditions: 23°C, 110-130 μmoles ; Each Data Point is an Average of 3 Different plants/Line FIG. 4

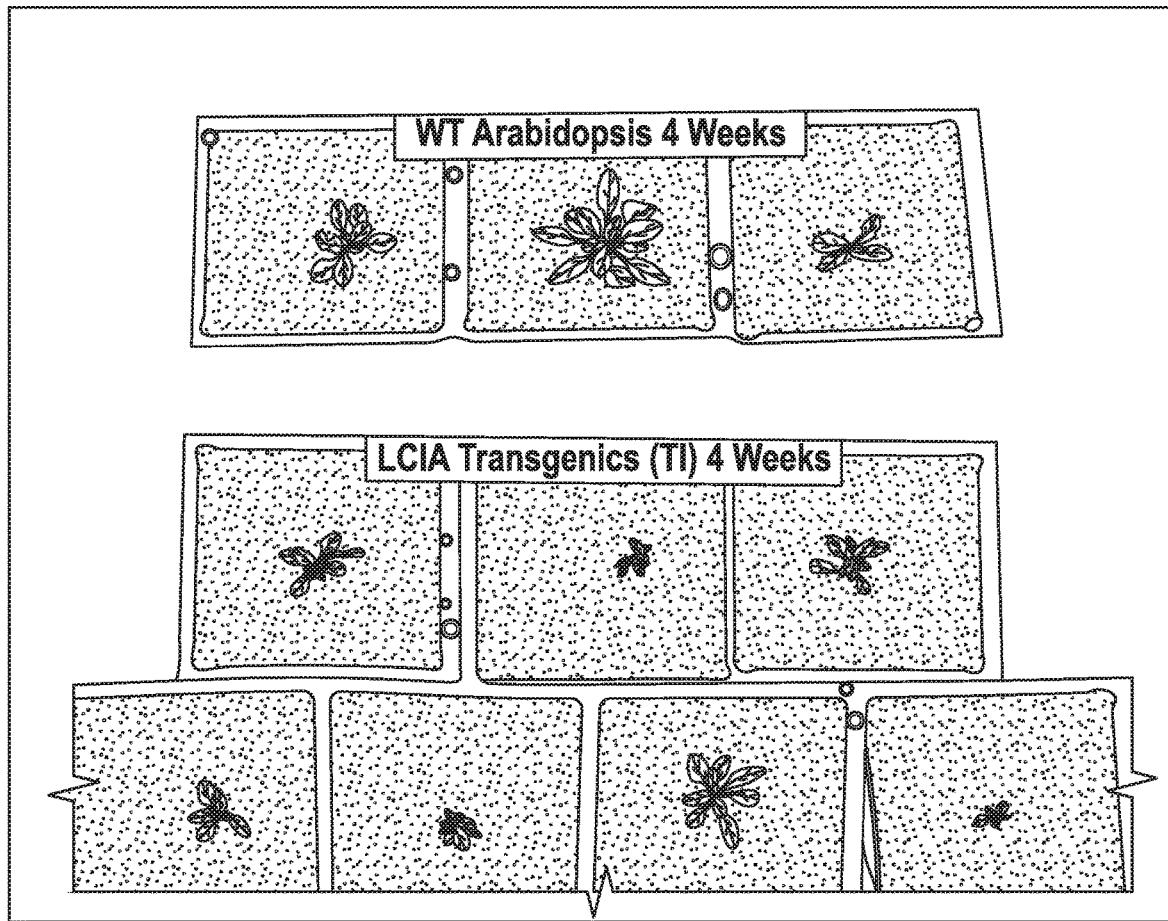


FIG. 5A

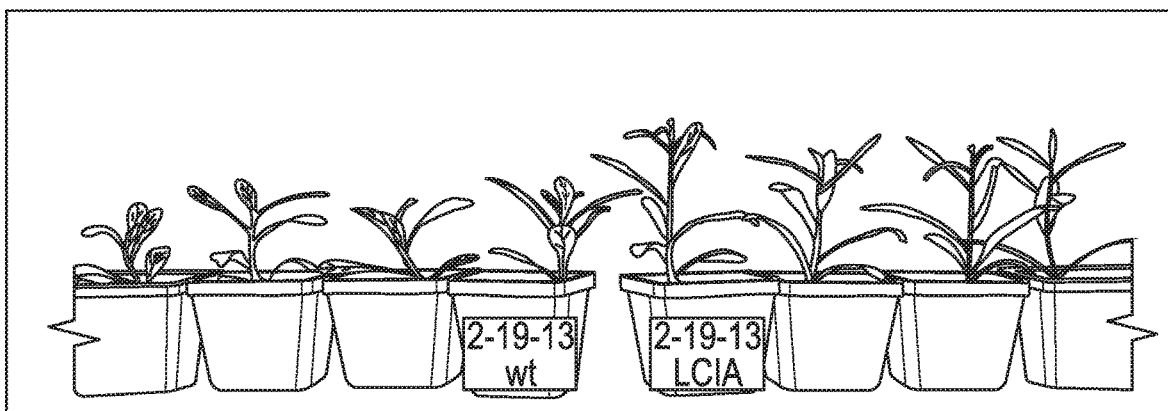


FIG. 5B

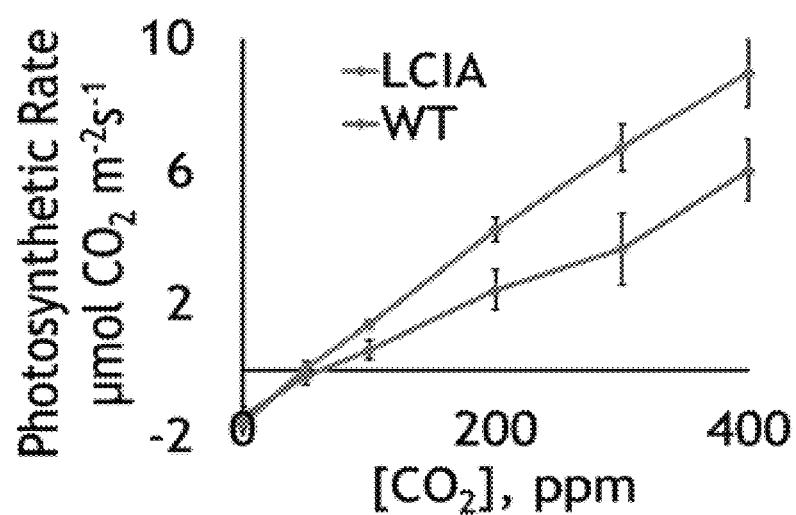
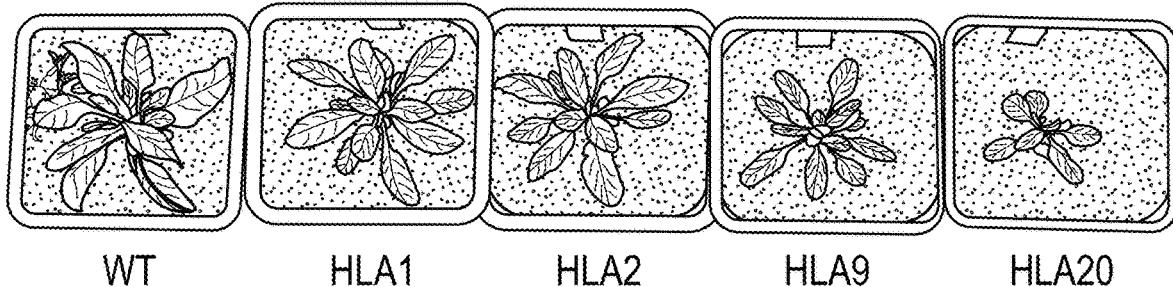


FIG. 5C



	ATP	28.2±4.2	27.5±3.8	27.0±1.6	21.5±1.6	24.4±2.0
ADP	11.0±2.4	8.4±0.8	8.2±1.1	6.8±1.1	7.7±0.8	
AMP	3.3±0.6	2.8±0.1	5.2±0.5	10.1±3.4	7.9±4.5	
P ₁	4.55±0.31	4.59±0.38	4.87±0.24	6.06±0.92	8.11±0.46	
EC	0.80±0.00	0.82±0.02	0.77±0.01	0.66±0.06	0.71±0.09	
NADPH	21.4±2.0	22.2±3.0	19.4±3.8	14.9±3.3	18.6±4.3	
NADH	5.71±0.85	7.27±0.63	4.46±0.72	2.71±0.43	2.35±0.63	
NADP	1.50±0.21	1.06±0.05	1.17±0.13	1.48±0.37	1.68±0.10	
NAD	22.1±0.04	22.7±6.6	27.4±0.7	23.1±1.7	21.6±3.0	
RP ₁	0.53±0.03	0.57±0.05	0.45±0.05	0.41±0.05	0.46±0.07	
RP ₂	0.21±0.03	0.27±0.07	0.14±0.02	0.11±0.02	0.10±0.03	

$$EC = \frac{ATP + 0.5ADP}{ATP + ADP + AMP} \quad RP_1 = \frac{NADPH + NADH}{NADP(H) + NAD(H)} \quad RP_2 = \frac{NADH}{NADH + NAD}$$

FIG. 6

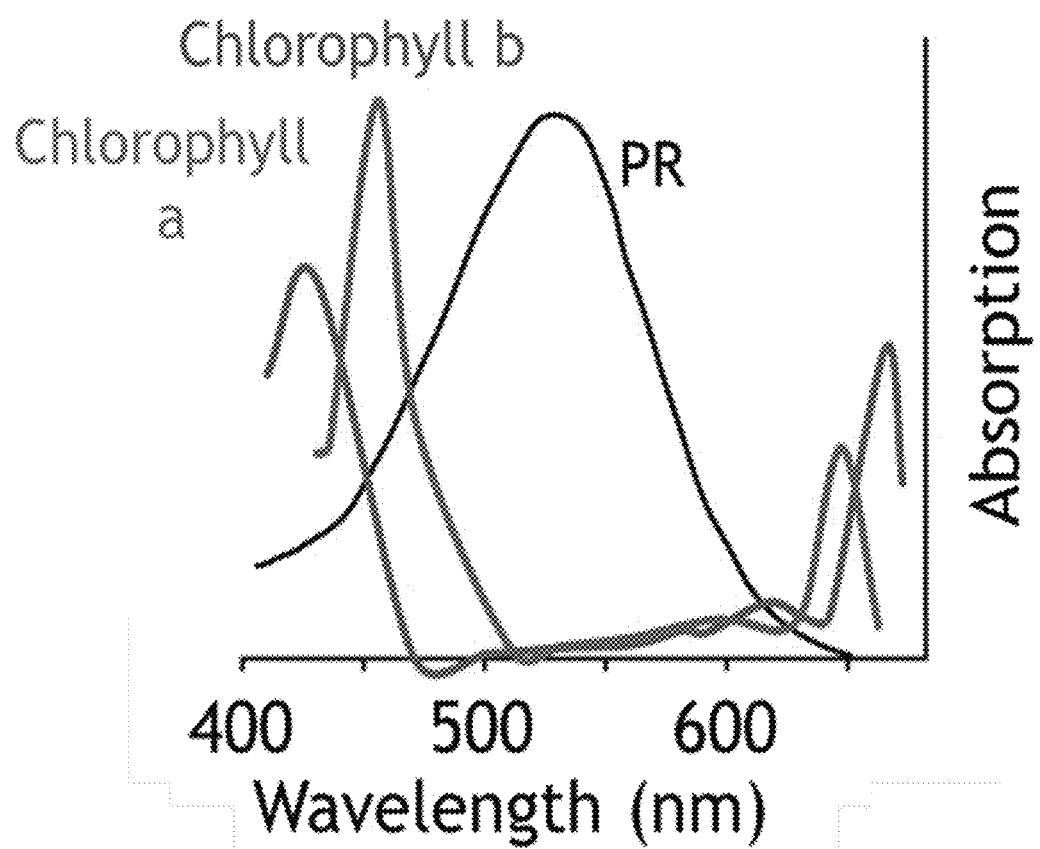


FIG. 7

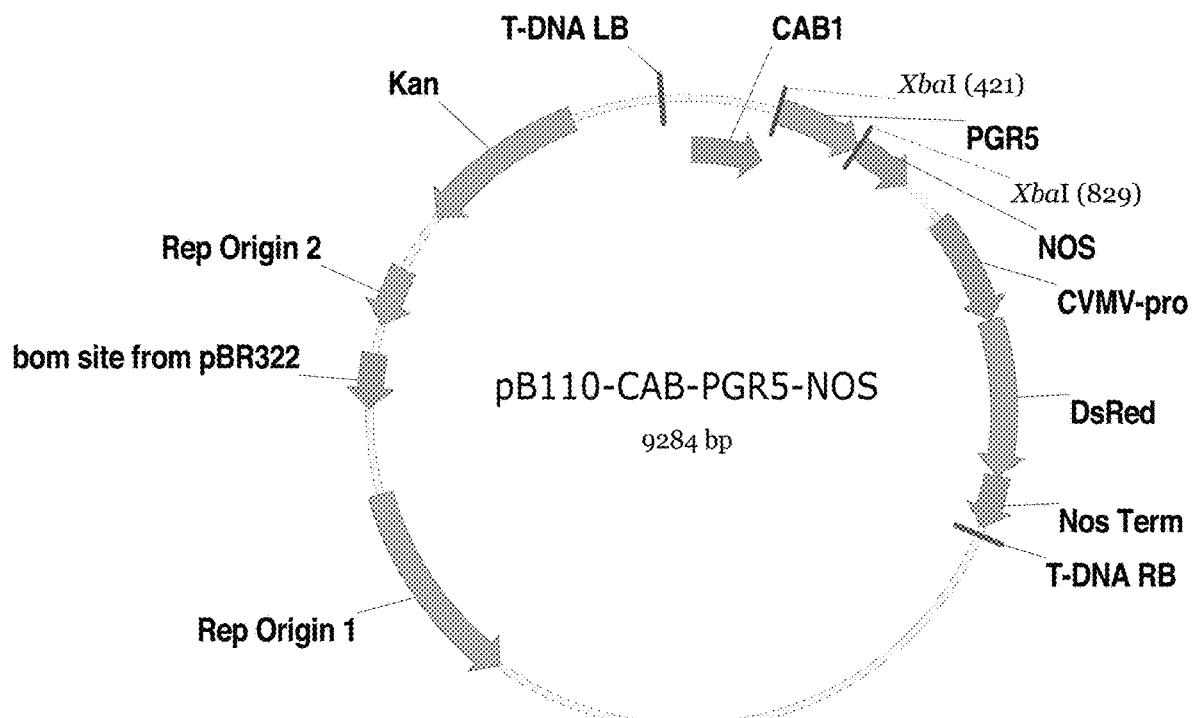


FIG. 8

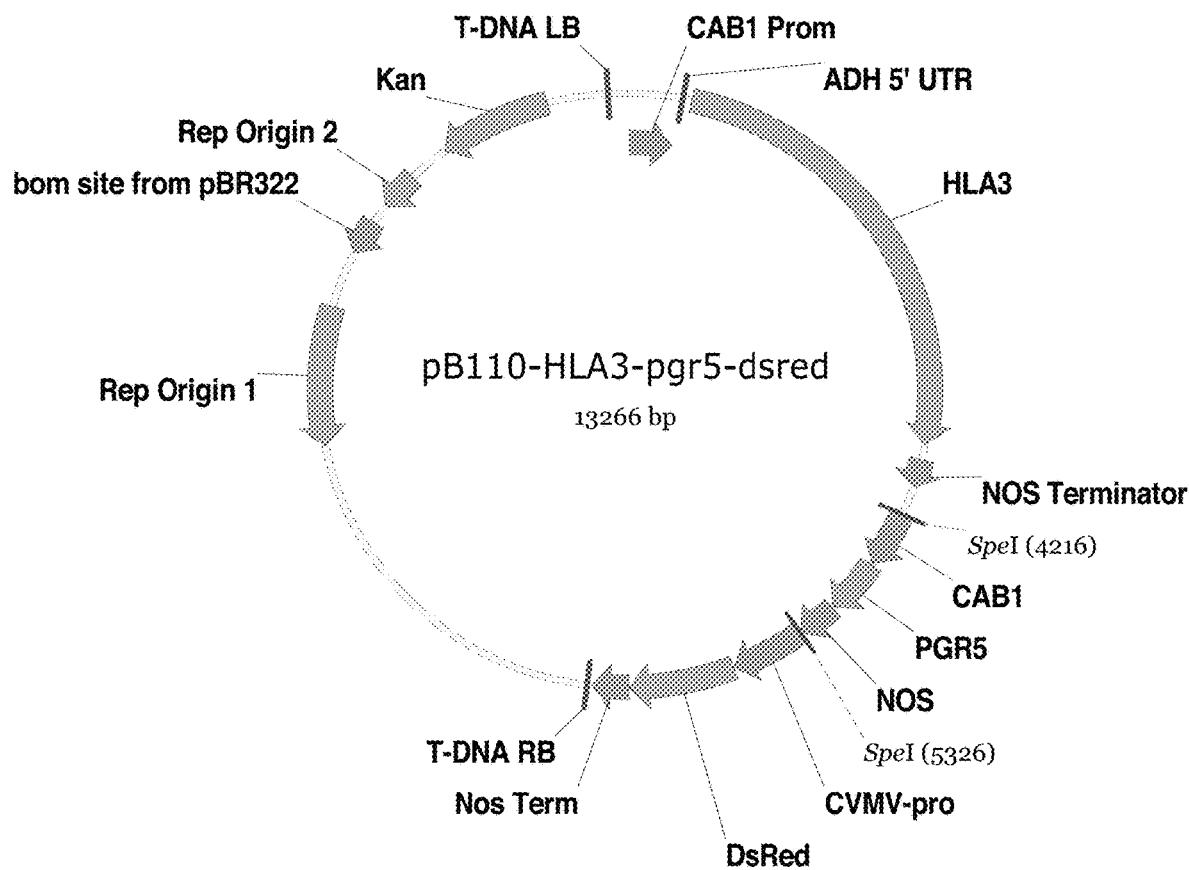


FIG. 9

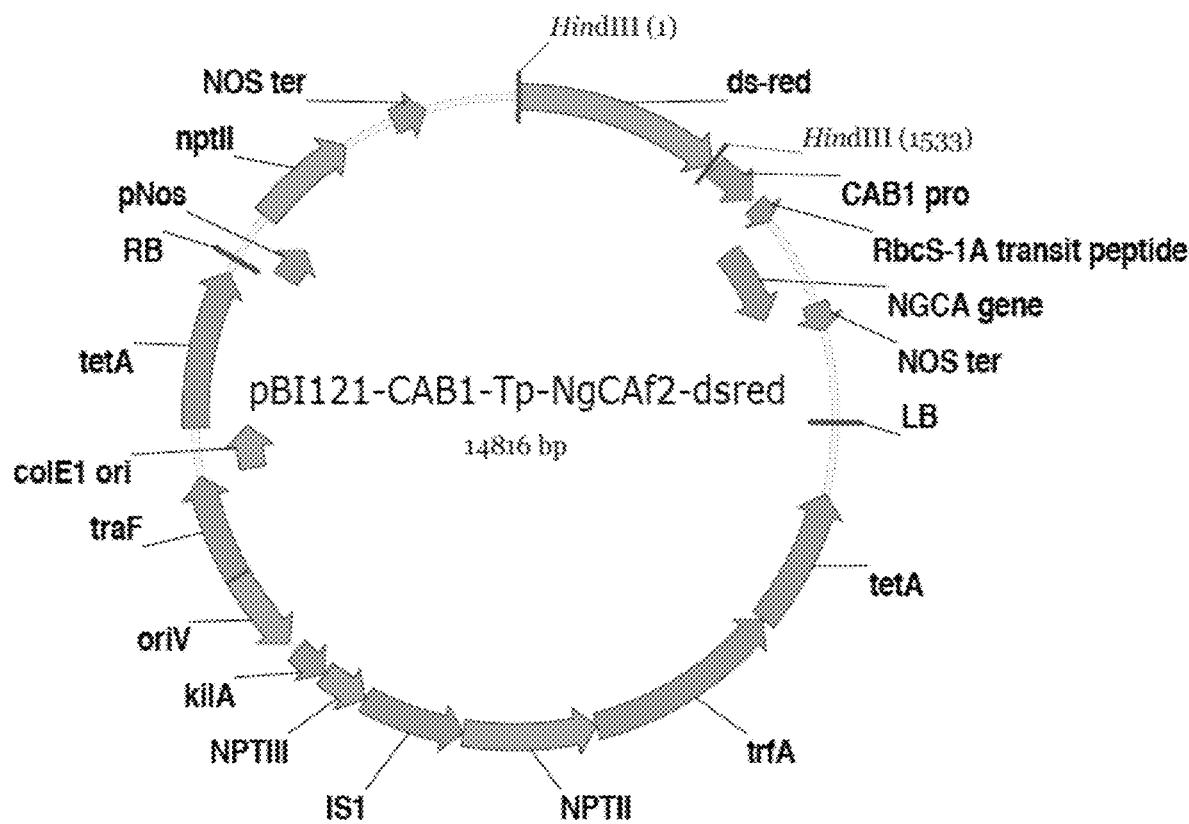


FIG. 10

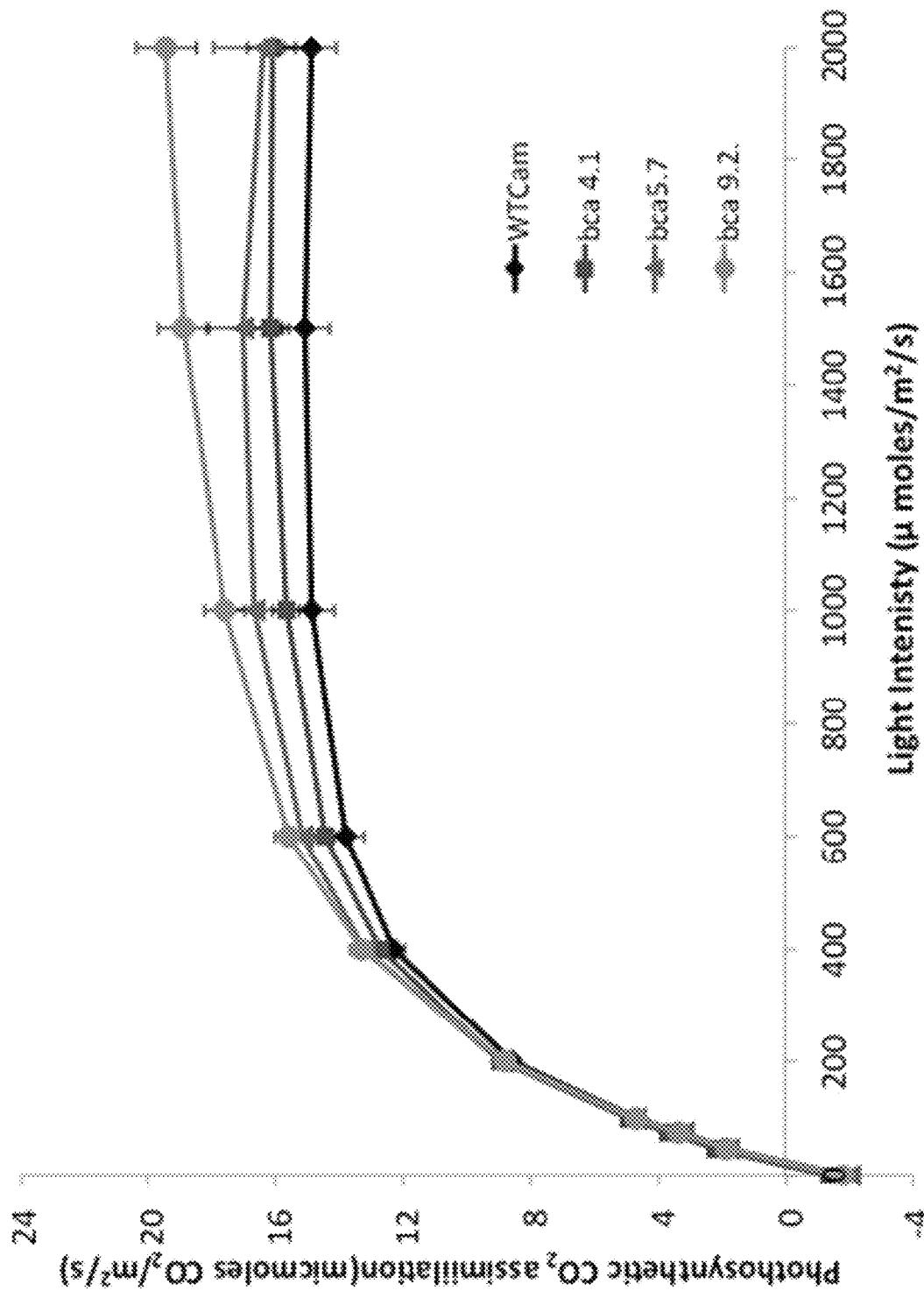


FIG. 11

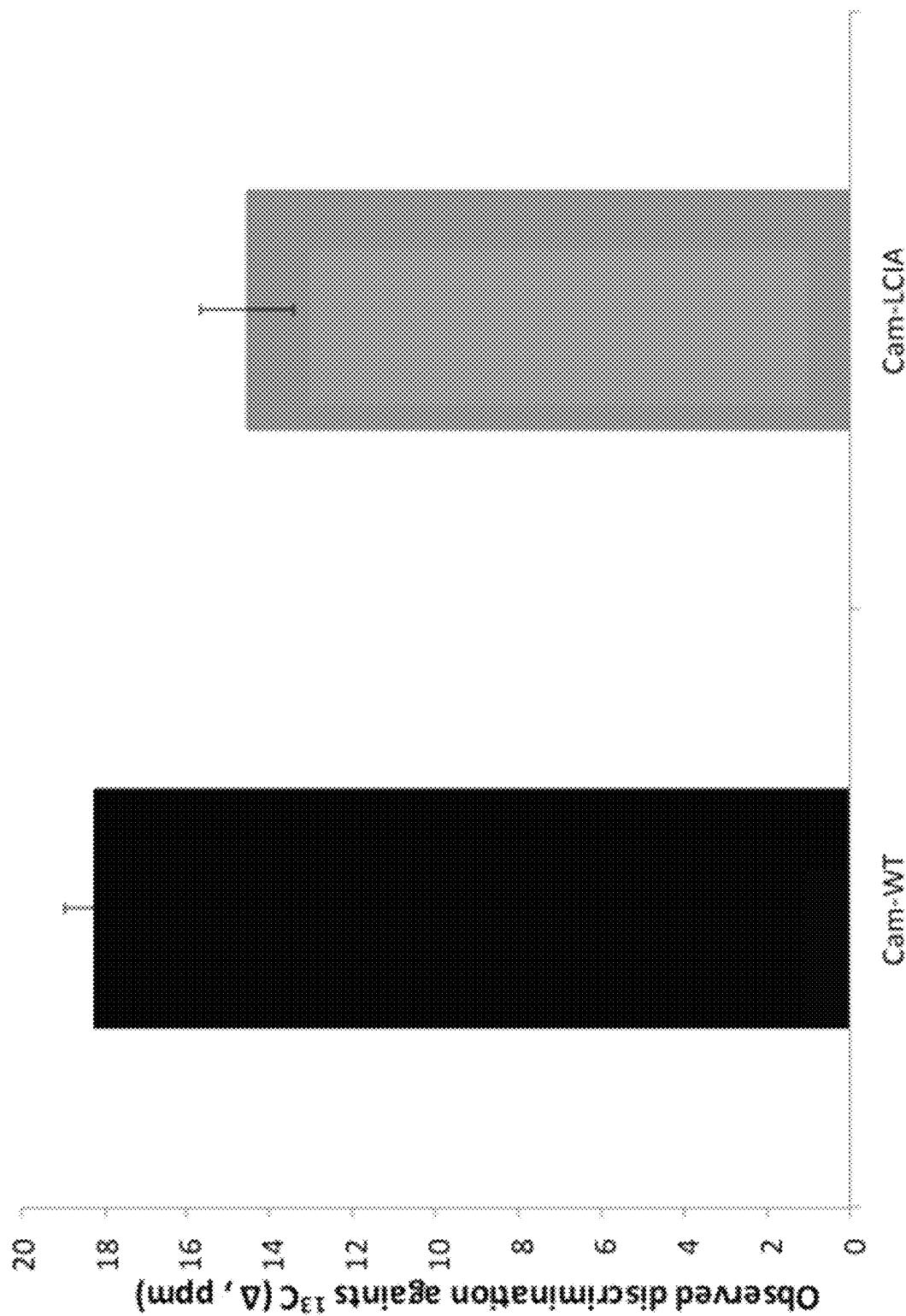


FIG. 12

1**CARBON FIXATION SYSTEMS IN PLANTS
AND ALGAE****CROSS-REFERENCE TO RELATED
APPLICATIONS**

This application is a continuation of U.S. patent application Ser. No. 16/358,331, titled "Improved Carbon Fixation Systems in Plants and Algae", filed on Mar. 3, 2019, which claims priority to U.S. patent application Ser. No. 15/411,854, entitled "Improved Carbon Fixation Systems in Plants and Algae", filed on Jan. 20, 2017, and issued on Mar. 3, 2019 as U.S. Pat. No. 10,233,458, which is a continuation of International Patent Application No. PCT/US2015/041617, entitled "Improved Carbon Fixation Systems in Plants and Algae", filed on Jul. 22, 2015, which claims priority to and the benefit of the filing of U.S. Provisional Patent Application No. 62/027,354, entitled "Carbon Fixation Systems in Plants and Algae", filed on Jul. 22, 2014, and the specification and claims thereof are incorporated herein by reference.

**STATEMENT REGARDING FEDERALLY
SPONSORED RESEARCH**

This invention was made with government support under grants Nos. DOE-CECO Prime No: DE-AR0000202, Sub No: 21018-N; DOE-CABS Prime No: DE-SC0001295, Sub No: 21017-NM NSF EF-1219603, NSF No:1219603. The U.S. government has certain rights in the invention.

SEQUENCE LISTING

The instant application contains a Sequence Listing which has been submitted electronically in ASCII format and is hereby incorporated by reference in its entirety. Said ASCII copy, created on Apr. 5, 2017, is named 040517_NMC0001-101-US_Sequence_Listing_ST25.txt and is 286 KB in size.

**INCORPORATION BY REFERENCE OF
MATERIAL SUBMITTED ON A COMPACT
DISC**

Not Applicable.

COPYRIGHTED MATERIAL

Not Applicable.

BACKGROUND

A major factor limiting photosynthetic efficiency is the competitive inhibition of CO₂ fixation by oxygen, due to lack of specificity of the enzyme RuBisCO. Incorporation of oxygen by RuBisCO is the first-dedicated step in photorespiration, a pathway that respires CO₂, compounding photosynthetic inefficiency. Overall, photorespiration reduces photosynthetic productivity by as much as 50% [1]. To date, attempts to engineer reduced oxygenase activity in RuBisCO have been largely unsuccessful.

Significantly, the cyanobacteria, eukaryotic microalgae, and C4 plants have evolved mechanisms to reduce photorespiration by concentrating CO₂ near RuBisCO, competitively inhibiting oxygenase activity and leading to substantial increases in yield and water use efficiency per unit

2

carbon fixed. However, carbon concentrating systems (CCMs) are not operational in the vast majority of plant species (i.e., C3 plants).

Attempts to reconstitute functional CCMs in C3 plants have been previously attempted by us and others, mainly focusing on engineering pathways that are directly involved in facilitating CO₂ transport into leaf chloroplasts. Note, for example, PCT International Publication WO 2012/125737; Sage and Sage (2009) Plant and Cell Physiol. 50(4):756-772; Zhu et al. (2010) J Intergr. Plant Biol. 52(8):762-770; Furbank et al. (2009) Funct. Plant Biol. 36(11):845-856; Weber and von Caemmerer (2010) Curr. Opin. Plant Biol.; Price (2013) J. Exp. Bot. 64(3):753-68; and U.S. Patent Application Publication No. 2013/0007916 A1.

However, ATP and NADPH production through light harvesting and electron transfer steps must be coordinated with carbon assimilation and additional energy requiring steps including CCM systems to prevent photoinhibition and to improve growth. Additionally, assimilatory flux and storage rates can limit carbon fixation due to feedback inhibition when sink demand is not matched to source capacity [2].

Thus, there is a critical need to improve plant productivity through integrated systems engineering approaches that balance source/sink interactions with energy and reductant production to develop energy-requiring, artificial CCMs that can effectively mimic those found in nature.

BRIEF SUMMARY OF THE INVENTION

Accordingly, in response to this need, the present disclosure provides methods for elevating cyclic electron transfer activity, improving carbon concentration, and enhancing carbon fixation in C3 and C4 plants, and algae, and producing biomass or other products from C3 or C4 plants, and algae, selected from among, for example, starches, oils, fatty acids, lipids, cellulose or other carbohydrates, alcohols, sugars, nutraceuticals, pharmaceuticals, fragrance and flavoring compounds, and organic acids, as well as transgenic plants produced thereby. These methods and transgenic plants and algae encompass the expression, or overexpression, of various combinations of genes that improve carbon concentrating systems in plants and algae, such as bicarbonate transport proteins, carbonic anhydrase, light driven proton pump, cyclic electron flow regulators, etc. Thus, among its various embodiments, the present disclosure provides the following:

A first embodiment of the present invention provides for a transgenic plant or alga, comprising within its genome, and expressing or overexpressing, a combination of heterologous nucleotide sequences encoding an ATP dependent bicarbonate anion transporter localized to the plasma membrane and a cyclic electron transfer modulator protein. The cyclic electron transfer modulator protein may be selected from a PGRL1 protein (for example SEQ ID NO:3), a PGR5 protein (for example SEQ ID NO:1), a leaf FNR1 protein (for example SEQ ID NO:96), a leaf FNR2 protein (for example SEQ ID NO:97), a Fd1 protein (for example SEQ ID NO:95), or any combination thereof and for example the ATP dependent bicarbonate anion transporter localized to the plasma membrane may be a HLA3 protein (for example SEQ ID NO:77). The transgenic plant or alga described may further comprise within its genome, and expressing or overexpressing the heterologous nucleotide sequence encoding a bicarbonate anion transporter protein localized to the chloroplast envelope. The transgenic plant or alga described herein may further comprise within its genome, and expressing or overexpressing the heterologous nucleotide sequence

a carbonic anhydrase protein. In a preferred embodiment, the cyclic electron transfer modulator protein is a PGR5 protein, in another preferred embodiment the cyclic electron transfer modulator protein is Fd1 protein, in yet another preferred embodiment, in still another preferred embodiment the cyclic electron transfer modulator protein is leaf FNR1, in a further preferred embodiment the cyclic electron transfer modulator protein is PGRL1. In a preferred embodiment the heterologous nucleotide sequences of the transgenic plant or alga encode i) a PGR5 protein, and a HLA3 protein; or ii) a PGR5 protein, a HLA3 protein and a PGRL1 protein or a PGR5 protein, a HLA3 protein, and a LCIA protein or a PGR5 protein, a HLA3 protein, a PGRL1 protein, a LCIA protein, and a BCA or HCA2 protein. In another preferred embodiment the heterologous nucleotide sequences of the transgenic plant or alga of wherein encode a PGR5 protein, a HLA3 protein, a LCIA protein and a BCA or optionally a HCA2 protein. The transgenic plant or alga as described wherein the PGR5 protein has an amino acid sequence at least 80% identical to SEQ ID NO:1; the HLA3 protein has an amino acid sequence at least 80% identical to SEQ ID NO:77; the PGRL1 protein has an amino acid sequence at least 80% identical to SEQ ID NO:3; the LCIA protein has an amino acid sequence at least 80% identical to SEQ ID NO:18; and/or the BCA protein has an amino acid sequence at least 80% identical to SEQ ID NO:21. Alternatively, the sequence identity/sequence similarity is about 75%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 100% to those specifically disclosed which includes for example proteins without a transit peptide sequence and the functional protein.

A second embodiment provides for a transgenic plant or alga, comprising within its genome, and expressing or overexpressing, a combination of heterologous nucleotide sequences encoding:

LCIA protein and BCA protein or HCA protein is provided. In a preferred embodiment the heterologous nucleotide sequences encode transgenic plant or alga wherein the LCIA protein has an amino acid sequence at least 80% identical to SEQ ID NO:18; and/or the BCA protein has an amino acid sequence at least 80% identical to SEQ ID NO:21 and the HCA protein has an amino acid sequence at least 80% identical to SEQ ID NO:19. Alternatively, the sequence identity/sequence similarity is about 75%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 100% to those specifically disclosed which include for example proteins without a transit peptide sequence and the functional protein.

A third embodiment provides for a transgenic plant or alga, comprising within its genome, and expressing or overexpressing, a combination of heterologous nucleotide sequences encoding an ATP dependent bicarbonate anion transporter localized to the plasma membrane (for example SEQ ID NO:77), a bicarbonate anion transporter localized to the chloroplast envelope (for example SEQ ID NO:18), a carbonic anhydrase, a proteorhodopsin protein targeted to thylakoid membranes (for example SEQ ID NO:98), and a R carotene monooxygenase protein (for example SEQ ID NO:100). In another preferred embodiment the proteorhodopsin comprises a chloroplast transit peptide selected from among a psbX stop-transfer trans-membrane domain fused to its C-terminus, a DNAJ transit peptide, a CAB transit peptide, a PGR5 transit peptide, and a psaD transit peptide. In another preferred embodiment the β -carotene monooxygenase is expressed under the control of a promoter selected from among an ethanol inducible gene promoter and a green tissue/leaf-specific promoter selected from among CAB and

rbcS. The proteorhodopsin may comprise an amino acid substitution selected from among L219E/T206S, M79T, and M79Y, and combinations thereof.

The carbonic anhydrase of the first, second, or third embodiment may be a BCA or optionally a HCA2 protein. The bicarbonate anion transporter localized to the chloroplast envelope of the first, second and third embodiment may be a LCIA protein. The ATP dependent bicarbonate anion transporter localized to the plasma membrane of the first and third embodiments may be HLA3.

A fourth embodiment provides for a method of making a transgenic plant or alga of a first embodiment wherein said method comprises expressing, or overexpressing, in a C₃ plant, a C₄ plant, or an alga, a combination of heterologous nucleotide sequences encoding an ATP dependent bicarbonate anion transporter localized to the plasma membrane and a cyclic electron transfer modulator protein. The cyclic electron transfer modulator protein may be selected from a PGRL1 protein, a PGR5 protein, a FNR1 protein, a FNR2 protein (leaf-form isotopes), a Fd1 protein, or any combination thereof and wherein the ATP dependent bicarbonate anion transporter localized to the plasma membrane is a HLA3 protein. The heterologous nucleotide sequences of the fourth embodiment further encoding a bicarbonate anion transporter protein localized to the chloroplast envelope for example the bicarbonate anion transporter protein is LCIA. Additionally, the heterologous nucleotide sequences encode a carbonic anhydrase protein for example a BCA protein or optionally a HCA2 protein. In a preferred embodiment the cyclic electron transfer modulator protein is a PGR5 protein and optionally a PGRL1 protein and or combination thereof.

and optionally a t-BCA1 protein and/or combination thereof.

A fifth embodiment provides a method of making a transgenic plant or alga as described in a second embodiment, wherein said method comprises expressing, or over-expressing, in a C3 plant, a C4 plant, or an alga, a combination of heterologous nucleotide sequences encoding a LCIA protein and a BCA protein or optionally a HCA protein.

protein.

A sixth embodiment provides a method of making a transgenic plant or alga of a third embodiment wherein said method comprises expressing, or overexpressing, in a C₃ plant, a C₄ plant, or an alga, a combination of heterologous nucleotide sequences encoding an ATP dependent bicarbonate anion transporter localized to the plasma membrane, a bicarbonate anion transporter, a carbonic anhydrase, a proteorhodopsin protein targeted to thylakoid membranes, and a R carotene monooxygenase protein. In a preferred embodiment the proteorhodopsin comprises a chloroplast transit peptide selected from among a psbX stop-transfer transmembrane domain fused to its C-terminus, a DNAJ transit peptide, a CAB transit peptide, a PGR5 transit peptide, and a psaD transit peptide. In another preferred embodiment the β-carotene monooxygenase is expressed under the control of a promoter selected from among an ethanol inducible gene promoter and a green tissue/leaf-specific promoter selected from among CAB and rbcS. In a preferred embodiment the proteorhodopsin comprises an amino acid substitution selected from among L219E/T206S, M79T, and M79Y, and combinations thereof. In another preferred embodiment the ATP dependent bicarbonate anion transporter localized to the plasma membrane is HLA3.

The transgenic plant of an embodiment disclosed herein may be a C3 plant or a C4 plant such as a transgenic oilseed plant or a transgenic food crop plant which may include the 65 genera *Brassica* (e.g., rapeseed/canola (*Brassica napus*; *Brassica carinata*; *Brassica nigra*; *Brassica oleracea*), Camelina, *Miscanthus*, and *Jatropha*; Jojoba (*Simmondsia*

chinensis), coconut; cotton; peanut; rice; safflower; sesame; soybean; mustard other than *Arabidopsis*; wheat; flax (lin-seed); sunflower; olive; corn; palm; palm kernel; sugarcane; castor bean; switchgrass; *Borago officinalis*; *Echium plantagineum*; *Cuphea hookeriana*; *Cuphea pulcherrima*; *Cuphea lanceolata*; *Ricinus communis*; *Coriandrum sativum*; *Crepis alpina*; *Vernonia galamensis*; *Momordica charantia*; and *Crambe abyssinica*, wheat, rice, maize (corn), barley, oats, sorghum, rye, and millet; peanuts, chickpeas, lentils, kidney beans, soybeans, lima beans; potatoes, sweet potatoes, and cassavas; soybeans, corn, canola, peanuts, palm, coconuts, safflower, cottonseed, sunflower, flax, olive, and safflower; sugar cane and sugar beets; bananas, oranges, apples, pears, breadfruit, pineapples, and cherries; tomatoes, lettuce, carrots, melons, strawberry, asparagus, broccoli, peas, kale, cashews, peanuts, walnuts, pistachio nuts, almonds; forage and turf grasses; alfalfa, clover; coffee, cocoa, kola nut, poppy; vanilla, sage, thyme, anise, saffron, menthol, peppermint, spearmint and coriander and preferably wheat, rice and canola. The transgenic alga of an embodiment disclosed herein may be selected from among a *Chlorella* species, a *Nannochloropsis* species, and a *Chlamydomonas* species. The heterologous nucleotide sequences are described in an embodiment may be codon-optimized for expression in said transgenic plant or alga. One aspect of the present invention provides for a transgenic plant or alga as described in an embodiment which exhibits enhanced CO₂ fixation compared to an otherwise identical control plant grown under the same conditions for example wherein CO₂ fixation is enhanced in the range of from about 10% to about 50% compared to that of an otherwise identical control plant grown under the same conditions.

A fourth embodiment provides for a part of said transgenic plant or alga of any embodiment described herein. For example, the part of said transgenic plant may be selected from among a protoplast, a cell, a tissue, an organ, a cutting, an explant, a reproductive tissue, a vegetative tissue, biomass, an inflorescence, a flower, a sepal, a petal, a pistil, a stigma, a style, an ovary, an ovule, an embryo, a receptacle, a seed, a fruit, a stamen, a filament, an anther, a male or female gametophyte, a pollen grain, a meristem, a terminal bud, an axillary bud, a leaf, a stem, a root, a tuberous root, a rhizome, a tuber, a stolon, a corm, a bulb, an offset, a cell of said plant in culture, a tissue of said plant in culture, an organ of said plant in culture, a callus, propagation materials, germplasm, cuttings, divisions, and propagations.

A fifth embodiment provides for a progeny or derivative of said transgenic plant or alga of any embodiment described herein. For example, the progeny or derivatives may be selected from among clones, hybrids, samples, seeds, and harvested material thereof and may be produced sexually or asexually.

Another embodiment of the present invention provides a method of elevating CET activity in a C3 plant, C4 plant, or alga wherein said method comprises expressing, or overexpressing, in a C3 plant, a C4 plant, or an alga, a combination of heterologous nucleotide sequences encoding an ATP dependent bicarbonate anion transporter localized to the plasma membrane and cyclic electron transfer modulator protein.

Yet another embodiment provides a method of enhancing carbon fixation in a C3 plant, C4 plant, or alga wherein said method comprises expressing, or overexpressing, in a C3 plant, a C4 plant, or an alga, a combination of heterologous nucleotide sequences encoding an ATP dependent bicarbonate anion transporter localized to the plasma membrane and a cyclic electron transfer modulator protein.

Yet another method provides for a method of producing biomass or other products from a C3 plant, C4 plant, or an alga, wherein said products are selected from among starches, oils, fatty acids, triacylglycerols, lipids, cellulose or other carbohydrates, alcohols, sugars, nutraceuticals, pharmaceuticals, fragrance and flavoring compounds, and organic acids wherein said method comprises expressing, or overexpressing, in a C3 plant, a C4 plant, or an alga, a combination of heterologous nucleotide sequences encoding an ATP dependent bicarbonate anion transporter localized to the plasma membrane and a cyclic electron transfer modulator protein. This method further comprises growing said plant or alga and harvesting said biomass or recovering said product from said plant or alga. Another aspect of the present invention provides for biomass or other product produced from a plant or alga selected from among starches, oils, fatty acids, lipids, cellulose or other carbohydrates, alcohols, sugars, nutraceuticals, pharmaceuticals, fragrance and flavoring compounds, and organic acids, made by a method of any one of the method of making a transgenic plant or alga embodiments herein.

Another embodiment provides a method of elevating cyclic electron transfer (CET) activity in a C3 plant, C4 plant, or alga wherein said method comprises expressing, or overexpressing, in a C3 plant, a C4 plant, or an alga, a combination of heterologous nucleotide sequences encoding an ATP dependent bicarbonate anion transporter localized to the plasma membrane, a bicarbonate anion transporter, a carbonic anhydrase, a proteorhodopsin protein targeted to thylakoid membranes; and a R carotene monooxygenase protein.

Another embodiment provides a method of enhancing carbon fixation in a C3 plant, C4 plant, or alga wherein said method comprises expressing, or overexpressing, in a C3 plant, a C4 plant, or an alga, a combination of heterologous nucleotide sequences encoding an ATP dependent bicarbonate anion transporter localized to the plasma membrane, a bicarbonate anion transporter, a carbonic anhydrase, a proteorhodopsin protein targeted to thylakoid membranes; and a R carotene monooxygenase protein.

Another embodiment provides for a method of producing biomass or other products from a C3 plant, C4 plant, or an alga, wherein said products are selected from among starches, oils, fatty acids, triacylglycerols, lipids, cellulose or other carbohydrates, alcohols, sugars, nutraceuticals, pharmaceuticals, fragrance and flavoring compounds, and organic acids wherein said method comprises expressing, or overexpressing, in a C3 plant, a C4 plant, or an alga, a combination of heterologous nucleotide sequences encoding an ATP dependent bicarbonate anion transporter localized to the plasma membrane, a bicarbonate anion transporter, a carbonic anhydrase, a proteorhodopsin protein targeted to thylakoid membranes; and a β carotene monooxygenase protein. The method further comprises growing said plant or alga and harvesting said biomass or recovering said product from said plant or alga.

Another embodiment provides for use of a construct comprising one or more nucleic acids encoding

- a) a PGR5 protein, and a HLA3 protein;
- b) a PGR5 protein, a HLA3 protein and a PGRL1 protein;
- c) a PGR5 protein, a HLA3 protein, and a LCIA protein;
- d) a PGR5 protein, a HLA3 protein, a LCIA protein and a BCA or HCA2 protein;
- e) a PGR5 protein, a HLA3 protein, a PGRL1 protein and a LCIA protein;
- f) a PGR5 protein, a HLA3 protein, a PGRL1 protein, a LCIA protein, and a BCA or HCA2 protein;

- g) a PGR5 protein, a HLA3 protein, and a BCA or HCA2 protein; or
- h) a PGRL1 protein, a HLA3 protein, a PGRL1 protein, and a BCA or HCA2 protein

for

- i) making a transgenic plant or alga of a first embodiment;
- ii) elevating CET activity in a C3 plant, C4 plant, or alga;
- iii) enhancing carbon fixation in a C3 plant, C4 plant, or alga; or
- iv) producing biomass or other products from a C3 plant, C4 plant, or an alga, wherein said products are selected from among starches, oils, fatty acids, triacylglycerols, lipids, cellulose or other carbohydrates, alcohols, sugars, nutraceuticals, pharmaceuticals, fragrance and flavoring compounds, and organic acids.

Another embodiment provides for use of a construct comprising one or more nucleic acids encoding

- a) a LCIA protein and a BCA or HCA2 protein;

for

- i) making a transgenic plant or alga of a second embodiment;
- ii) elevating CET activity in a C3 plant, C4 plant, or alga;
- iii) enhancing carbon fixation in a C3 plant, C4 plant, or alga; or
- iv) producing biomass or other products from a C3 plant, C4 plant, or an alga, wherein said products are selected from among starches, oils, fatty acids, triacylglycerols, lipids, cellulose or other carbohydrates, alcohols, sugars, nutraceuticals, pharmaceuticals, fragrance and flavoring compounds, and organic acids.

One aspect of the present invention provides for a transgenic plant or alga, comprising within its genome, and expressing or overexpressing, a combination of heterologous nucleotide sequences encoding:

1. i) a PGRL1 protein, a PGR5 protein, and a HLA3 protein; or
- ii) a PGRL1 protein, a PGR5 protein, a HLA3 protein, a LCIA protein, and a BCA or HCA2 protein; or
- iii) a Fd1 protein, a HLA3 protein, a LCIA protein, and a BCA or HCA2 protein; or
- iv) a leaf FNR1 protein, a HLA3 protein, a LCIA protein, and a BCA or HCA2 protein; or
- v) a proteorhodopsin protein targeted to thylakoid membranes, a HLA3 protein, a LCIA protein, a BCA or HCA2 protein, and a β-carotene monooxygenase.

2. The transgenic plant or alga of 1, wherein said proteorhodopsin comprises a chloroplast transit peptide selected from among a psbX stop-transfer trans-membrane domain fused to its C-terminus, a DNAJ transit peptide, a CAB transit peptide, a PGR5 transit peptide, and a psaD transit peptide.

3. The transgenic plant or alga of 1 or 2, wherein said β-carotene monooxygenase is expressed under the control of a promoter selected from among an ethanol inducible gene promoter and a green tissue/leaf-specific promoter selected from among CAB and rbcS.

4. The transgenic plant or alga of any one of 1-3, wherein said proteorhodopsin comprises an amino acid substitution selected from among L219E/T206S, M79T, and M79Y, and combinations thereof.

5. The transgenic plant of any one of 1-4, which is a C3 plant or a C4 plant.

6. The transgenic plant of any one of 1-5, which is a transgenic oilseed plant or a transgenic food crop plant.

7. The transgenic oilseed plant of 6, which is selected from among plants of the genera *Brassica* (e.g., rapeseed/canola (*Brassica napus*; *Brassica carinata*; *Brassica nigra*;

Brassica oleracea), *Camelina*, *Misanthus*, and *Jatropha*; *Jojoba* (*Simmondsia chinensis*), coconut; cotton; peanut; rice; safflower; sesame; soybean; mustard other than *Arabidopsis*; wheat; flax (linseed); sunflower; olive; corn; palm; palm kernel; sugarcane; castor bean; switchgrass; *Borago officinalis*; *Echium plantagineum*; *Cuphea hookeriana*; *Cuphea pulcherrima*; *Cuphea lanceolata*; *Ricinus communis*; *Coriandrum sativum*; *Crepis alpina*; *Vernonia galamensis*; *Momordica charantia*; and *Crambe abyssinica*.

10. The transgenic alga of any one of 1-5, which is selected from among *Chlorella* sp., *Nannochloropsis* sp., and *Chlamydomonas* sp.

11. The transgenic plant or alga of any one of 1-8, wherein said heterologous nucleotide sequences are codon-optimized for expression in said transgenic plant or alga.

12. The transgenic plant or alga of any one of 1-9, which exhibits enhanced CO₂ fixation compared to an otherwise identical control plant grown under the same conditions.

13. The transgenic plant or alga of 10, wherein CO₂ fixation is enhanced in the range of from about 10% to about 50% compared to that of an otherwise identical control plant grown under the same conditions.

14. A part of said transgenic plant or alga of any one of 1-11.

15. The part of said transgenic plant of 12, which is selected from among a protoplast, a cell, a tissue, an organ, a cutting, an explant, a reproductive tissue, a vegetative tissue, biomass, an inflorescence, a flower, a sepal, a petal, a pistil, a stigma, a style, an ovary, an ovule, an embryo, a receptacle, a seed, a fruit, a stamen, a filament, an anther, a male or female gametophyte, a pollen grain, a meristem, a terminal bud, an axillary bud, a leaf, a stem, a root, a tuberous root, a rhizome, a tuber, a stolon, a corm, a bulb, an offset, a cell of said plant in culture, a tissue of said plant in culture, an organ of said plant in culture, a callus, propagation materials, germplasm, cuttings, divisions, and propagations.

16. Progeny or derivatives of said transgenic plant or alga of any one of 1-11.

17. The progeny or derivatives of 14, which is selected from among clones, hybrids, samples, seeds, and harvested material thereof.

18. The progeny of 14 or 15, which is produced sexually.

19. The progeny of 14 or 15, which is produced asexually.

20. Another aspect of the present invention provides for a method selected from among:

18. i) making a transgenic plant or alga of any one of 1-11;
- ii) elevating CET activity in a C3 plant, C4 plant, or alga;
- iii) enhancing carbon fixation in a C3 plant, C4 plant, or alga; and

21. iv) producing biomass or other products from a C3 plant, C4 plant, or alga, wherein said products are selected from among starches, oils, fatty acids, triacylglycerols, lipids, cellulose or other carbohydrates, alcohols, sugars, nutraceuticals, pharmaceuticals, fragrance and flavoring compounds, and organic acids,

wherein said method comprises expressing, or overexpressing, in a C3 plant, a C4 plant, or an alga, a combination of heterologous nucleotide sequences encoding:

22. a) a PGRL1 protein, a PGR5 protein, and a HLA3 protein; or

23. b) a PGRL1 protein, a PGR5 protein, a HLA3 protein, a LCIA protein, and a BCA or HCA2 protein; or

24. c) a Fd1 protein, a HLA3 protein, a LCIA protein, and a BCA or HCA2 protein; or

25. d) a leaf FNR1 protein, a HLA3 protein, a LCIA protein, and a BCA or HCA2 protein; or

e) a proteorhodopsin protein targeted to thylakoid membranes, a HLA3 protein, a LCIA protein, a BCA or HCA2 protein, and a β -carotene monooxygenase.

19. The method of 18, wherein step iv) further comprises growing said plant or alga and harvesting said biomass or recovering said product from said plant or alga.

20. The method of 18 or 19, wherein said proteorhodopsin comprises a chloroplast transit peptide selected from among a psbX stop-transfer trans-membrane domain fused to its C-terminus, a DNAJ transit peptide, a CAB transit peptide, a PGR5 transit peptide, and a psaD transit peptide.

21. The method of any one of 18-20, wherein said β -carotene monooxygenase is expressed under the control of a promoter selected from among an ethanol inducible gene promoter and a green tissue/leaf-specific promoter selected from among CAB and rbcS.

22. The method of any one of 18-21, wherein said proteorhodopsin comprises an amino acid substitution selected from among L219E/T206S, M79T, and M79Y, and combinations thereof.

23. The method of any one of 18-22, wherein said transgenic plant is a C3 plant, a C4 plant, or an alga.

24. The method of any one of 18-23, wherein said transgenic plant is a transgenic oilseed plant or a transgenic food crop plant.

25. The method of 24, wherein said transgenic oilseed plant is selected from among plants of the genera *Brassica* (e.g., rapeseed/canola (*Brassica napus*; *Brassica carinata*; *Brassica nigra*; *Brassica oleracea*), Camelina, *Misanthus*, and Jatropha; Jojoba (*Simmondsia chinensis*), coconut; cotton; peanut; rice; safflower; sesame; soybean; mustard other than *Arabidopsis*; wheat; flax (linseed); sunflower; olive; corn; palm; palm kernel; sugarcane; castor bean; switchgrass; *Borago officinalis*; *Echium plantagineum*; *Cuphea hookeriana*; *Cuphea pulcherrima*; *Cuphea lanceolata*; *Ricinus communis*; *Coriandrum sativum*; *Crepis alpina*; *Vernonia galamensis*; *Momordica charantia*; and *Crambe abyssinica*.

26. The method of any one of 18-23, wherein said alga is selected from among *Chlorella* sp., *Nannochloropsis* sp., and *Chlamydomonas* sp.

27. The method of any one of 18-26, wherein said heterologous nucleotide sequences are codon-optimized for expression in said transgenic plant or alga.

28. The method of any one of 18-27, wherein said transgenic plant or alga exhibits enhanced CO₂ fixation compared to an otherwise identical control plant or alga grown under the same conditions.

29. The method of 28, wherein CO₂ fixation is enhanced in the range of from about 10% to about 50% compared to that of an otherwise identical control plant or alga grown under the same conditions.

Another aspect of the present invention provides for a transgenic plant or alga made by the method of any one of 18-29.

Yet another aspect of the present invention provides for a biomass or other product from a plant or alga, selected from among starches, oils, fatty acids, lipids, cellulose or other carbohydrates, alcohols, sugars, nutraceuticals, pharmaceuticals, fragrance and flavoring compounds, and organic acids, made by the method of any one of 18-29.

In addition to the various embodiments listed above, in the Examples below, and in the claims, this disclosure further variously encompasses the presently disclosed and claimed CCM protein combinations in further combinations with the genes and proteins focusing on engineering pathways that are directly involved in facilitating CO₂ transport

into leaf chloroplasts, disclosed and claimed in the inventors' previous application PCT International Publication WO 2012/125737. The present disclosure encompasses any combination of genes disclosed herein with any combination of genes disclosed in WO 2012/125737 and in Tables D1-D9 to improve carbon concentrating systems (CCMs) in plants and algae.

Table D1 represents different classes of α -CAs found in mammals.

Table D2-D4 represents representative species, Gene bank accession numbers, and amino acid sequences for various species of suitable CA genes.

Table D5 represents the codon optimized DNA sequence for chloroplast expression in *Chlamydomonas reinhardtii*. In Table D5, the underlines sequences represent restriction sites, and bases changed to optimize chloroplast expression are listed in lower case. Table D6 provides a breakdown of the number and type of each codon optimized.

Representative species and Gene bank accession numbers for various species of bicarbonate transporter are listed below in Tables D8-D9.

Further scope of the applicability of the presently disclosed embodiments will become apparent from the detailed description and drawing(s) provided below. However, it should be understood that the detailed description and specific examples, while indicating preferred embodiments of this disclosure, are given by way of illustration only since various changes and modifications within the spirit and scope of these embodiments will become apparent to those skilled in the art from this detailed description.

BRIEF DESCRIPTION OF THE SEQUENCE LISTING

The disclosure can be more fully understood from the following detailed description and the accompanying Sequence Listing, which form a part of this application.

The sequence descriptions summarize the Sequence Listing attached hereto. The Sequence Listing contains standard symbols and format used for nucleotide and amino acid sequence data comply with the rules set forth in 37 C.F.R. § 1.822.

BRIEF DESCRIPTION OF THE DRAWINGS

The above and other aspects, features, and advantages of the present disclosure will be better understood from the following detailed descriptions taken in conjunction with the accompanying drawing(s), all of which are given by way of illustration only, and are not limitative of the presently disclosed embodiments, in which:

FIG. 1. Model of the *Chlamydomonas* CCM showing the localization of inorganic carbon transporters (HLA3, LCIA) and carbonic anhydrase (CAH: CAH1, CAH3, and CAH6) [S]), and Rubisco. LCIB is an essential protein for CCM in *Chlamydomonas*. Its exact function is unknown.

FIG. 2.(A-B) Growth phenotypes of VVT and HLA3 transgenic (T3) *Arabidopsis* initially grown on MS media (plus nitrate, NO₃⁻). (B) MS media (plus ammonium (NH₄⁺) and sucrose) or in soil (ammonium only). X indicates plants died. Numbers refer to plant lines.

FIG. 3.(A-B) Growth phenotypes of VVT and HCA-II transgenic (T1) *Arabidopsis* 4 weeks after germination. (B) Growth phenotype of VVT *Arabidopsis* (Col-0, left) and the BCA transgenic (T3) (right).

FIG. 4. Photosynthetic assimilation rate of CO₂ in three transgenic lines (P1, P5, P6) of *Arabidopsis* expressing BCA

11

(bacterial carbonic anhydrase) measured using a LICOR 6400 gas analyzer. These lines showed ~30% increase in their photosynthetic efficiency when compared to WT *Arabidopsis* (Col-0).

FIG. 5.(A-C) Growth phenotypes of WT and LCIA transgenic (T1) *Arabidopsis* plants four weeks after germination. (B) Four-week-old WT (left 4 plants) and independent transgenic Camelina (right 4 plants) expressing LCIA. (C) CO₂-dependent photosynthetic rates of WT and LCIA transgenic Camelina.

FIG. 6. Phenotype of HLA3 transgenics grown on nitrate. Energy charge and reductive potential of WT and HLA3 transgenic *Arabidopsis*. Adenylate, nucleotide cofactors, and inorganic phosphate levels measured as nmole/gFW for plants grown on nitrate. Values are averages±SE.

FIG. 7. Photosynthetically active radiation in proteorhodopsin relative to plant-based chlorophyll [49].

FIG. 8. Plasmid pB110-CAB-PGR5-NOS (Example 1).

FIG. 9. Plasmid pB110-HLA3-pgr5-dsred (Example 1).

FIG. 10. Plasmid pBI 121-CAB1-Tp-NgCAF2-dsred (Example 1).

FIG. 11 illustrates light response curves of Camelina BCA lines.

FIG. 12 illustrates expression of LCIA in Camelina vs WT.

DETAILED DESCRIPTION OF SEVERAL EMBODIMENTS

The following detailed description is provided to aid those skilled in the art in practicing the various embodiments of the present disclosure described herein, including all the methods, uses, compositions, etc., described herein. Even so, the following detailed description should not be construed to unduly limit the present disclosure, as modifications and variations in the embodiments herein discussed may be made by those of ordinary skill in the art without departing from the spirit or scope of the present discoveries.

The present disclosure is explained in greater detail below. This disclosure is not intended to be a detailed catalog of all the different ways in which embodiments of this disclosure can be implemented, or all the features that can be added to the instant embodiments. For example, features illustrated with respect to one embodiment may be incorporated into other embodiments, and features illustrated with respect to a particular embodiment may be deleted from that embodiment. In addition, numerous variations and additions to the various embodiments suggested herein will be apparent to those skilled in the art in light of the instant disclosure, which variations and additions do not depart from the scope of the instant disclosure. Hence, the following specification is intended to illustrate some particular embodiments of the disclosure, and not to exhaustively specify all permutations, combinations, and variations thereof.

Any feature, or combination of features, described herein is(are) included within the scope of the present disclosure, provided that the features included in any such combination are not mutually inconsistent as will be apparent from the context, this specification, and the knowledge of one of ordinary skill in the art. Additional advantages and aspects of the present disclosure are apparent in the following detailed description and claims.

The contents of all publications, patent applications, patents, and other references mentioned herein are incorporated

12

by reference herein in their entirety. In case of conflict, the present specification, including explanations of terms, will control.

Definitions

The following definitions are provided to aid the reader in understanding the various aspects of the present disclosure. Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by those of ordinary skill in the art to which the disclosure pertains.

As used herein and in the appended claims, the singular forms “a”, “an”, and “the” include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to “a plant” includes a plurality of such plants, reference to “a cell” includes one or more cells and equivalents thereof known to those skilled in the art, and so forth. Similarly, the word “or” is intended to include “and” unless the context clearly indicates otherwise. Hence “comprising A or B” means including A, or B, or A and B. Furthermore, the use of the term “including”, as well as other related forms, such as “includes” and “included”, is not limiting.

The term “about” as used herein is a flexible word with a meaning similar to “approximately” or “nearly”. The term “about” indicates that exactitude is not claimed, but rather a contemplated variation. Thus, as used herein, the term “about” means within 1 or 2 standard deviations from the specifically recited value, or ±a range of up to 20%, up to 15%, up to 10%, up to 5%, or up to 4%, 3%, 2%, or 1% compared to the specifically recited value.

The term “comprising” as used in a claim herein is open-ended, and means that the claim must have all the features specifically recited therein, but that there is no bar on additional features that are not recited being present as well. The term “comprising” leaves the claim open for the inclusion of unspecified ingredients even in major amounts. The term “consisting essentially of” in a claim means that the invention necessarily includes the listed ingredients, and is open to unlisted ingredients that do not materially affect the basic and novel properties of the invention. A “consisting essentially of” claim occupies a middle ground between closed claims that are written in a closed “consisting of” format and fully open claims that are drafted in a “comprising” format. These terms can be used interchangeably herein if, and when, this may become necessary. Furthermore, the use of the term “including”, as well as other related forms, such as “includes” and “included”, is not limiting.

“BCA” refers to bacterial carbonic anhydrase.

“CCMs” and the like refer to carbon concentrating systems.

“CET” refers to cyclic electron transfer.

“LET” refers to linear electron transfer.

“WT” refers to wild-type.

“Cyclic electron transfer modulator protein” refers to any protein natural or synthetic that improves the separation of charge across the thylakoid membrane resulting in improved photophosphorylation with the production of chemical energy. Examples of such modulators are the PGR5 and PRGL1 reductases, however improved proteins in the electron transport chain such as cytochromes, ATPases, ferredoxin-NADP reductase, NAD(P)H-plastoquinone reductase, and the like are also CET modulator proteins.

Unless otherwise stated, nucleic acid sequences in the text of this specification are given, when read from left to right, in the 5' to 3' direction. Nucleic acid sequences may be

13

provided as DNA or as RNA, as specified; disclosure of one necessarily defines the other, as is known to one of ordinary skill in the art and is understood as included in embodiments where it would be appropriate. Nucleotides may be referred to by their commonly accepted single-letter codes. Unless otherwise indicated, amino acid sequences are written left to right in amino to carboxyl orientation, respectively. Amino acids may be referred to herein by either their commonly known three letter symbols or by the one-letter symbols recommended by the IUPAC-IUM Biochemical Nomenclature Commission. It is further to be understood that all base sizes or amino acid sizes, and all molecular weight or molecular mass values, given for nucleic acids or polypeptides are approximate, and are provided for description purposes and are not to be unduly limiting.

Regarding disclosed ranges, the endpoints of all ranges directed to the same component or property are inclusive and independently combinable (e.g., ranges of "up to about 25 wt. %, or, more specifically, about 5 wt. % to about 20 wt. %," is inclusive of the endpoints and all intermediate values of the ranges of "about 5 wt. % to about 25 wt. %," etc.). Numeric ranges recited with the specification are inclusive of the numbers defining the range and include each integer within the defined range.

As used herein, "altering level of production" or "altering level of expression" means changing, either by increasing or decreasing, the level of production or expression of a nucleic acid sequence or an amino acid sequence (for example a polypeptide, an siRNA, a miRNA, an mRNA, a gene), as compared to a control level of production or expression.

"Conservative amino acid substitutions": It is well known that certain amino acids can be substituted for other amino acids in a protein structure without appreciable loss of biochemical or biological activity. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence substitutions can be made in a protein sequence, and, of course, its underlying DNA coding sequence, and nevertheless obtain a protein with like properties. Thus, various changes can be made in the amino acid sequences disclosed herein, or in the corresponding DNA sequences that encode these amino acid sequences, without appreciable loss of their biological utility or activity.

Proteins and peptides biologically functionally equivalent to the proteins and peptides disclosed herein include amino acid sequences containing conservative amino acid changes in the fundamental amino acid sequence. In such amino acid sequences, one or more amino acids in the fundamental sequence can be substituted, for example, with another amino acid(s), the charge and polarity of which is similar to that of the native amino acid, i.e., a conservative amino acid substitution, resulting in a silent change.

It should be noted that there are a number of different classification systems in the art that have been developed to describe the interchangeability of amino acids for one another within peptides, polypeptides, and proteins. The following discussion is merely illustrative of some of these systems, and the present disclosure encompasses any of the "conservative" amino acid changes that would be apparent to one of ordinary skill in the art of peptide, polypeptide, and protein chemistry from any of these different systems.

As disclosed in U.S. Pat. No. 5,599,686, certain amino acids in a biologically active peptide, polypeptide, or protein can be replaced by other homologous, isosteric, and/or isoelectronic amino acids, wherein the biological activity of the original molecule is conserved in the modified peptide,

14

polypeptide, or protein. The following list of amino acid replacements is meant to be illustrative and is not limiting:

5	Original Amino Acid	Replacement Amino Acid(s)
	Ala	Gly
	Arg	Lys, ornithine
	Asn	Gln
10	Asp	Glu
	Glu	Asp
	Gln	Asn
	Gly	Ala
	Ile	Val, Leu, Met, Nle (norleucine)
	Leu	Ile, Val, Met, Nle
	Lys	Arg
	Met	Leu, Ile, Nle, Val
	Phe	Tyr, Trp
	Ser	Thr
	Thr	Ser
	Trp	Phe, Tyr
	Tyr	Phe, Trp
20	Val	Leu, Ile, Met, Nle

In another system, substitutes for an amino acid within a fundamental sequence can be selected from other members of the class to which the naturally occurring amino acid belongs. Amino acids can be divided into the following four groups: (1) acidic amino acids; (2) basic amino acids; (3) neutral polar amino acids; and (4) neutral non-polar amino acids. Representative amino acids within these various groups include, but are not limited to: (1) acidic (negatively charged) amino acids such as aspartic acid and glutamic acid; (2) basic (positively charged) amino acids such as arginine, histidine, and lysine; (3) neutral polar amino acids such as glycine, serine, threonine, cysteine, cystine, tyrosine, asparagine, and glutamine; (4) neutral nonpolar (hydrophobic) amino acids such as alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine.

Conservative amino acid changes within a fundamental peptide, polypeptide, or protein sequence can be made by substituting one amino acid within one of these groups with another amino acid within the same group.

Some of the other systems for classifying conservative amino acid interchangeability in peptides, polypeptides, and proteins applicable to the sequences of the present disclosure include, for example, the following:

Functionally defining common properties between individual amino acids by analyzing the normalized frequencies of amino acid changes between corresponding proteins of homologous organisms (Schulz, G. E. and R. H. Schirmer 1979) Principles of Protein Structure (Springer Advanced Texts in Chemistry), Springer-Verlag). According to such analyses, groups of amino acids can be defined where amino acids within a group exchange preferentially with each other, and therefore resemble each other most in their impact on overall protein structure;

Making amino acid changes based on the hydrophobicity index of amino acids as described by Kyte and Doolittle (1982) J. Mol. Biol. 157(1):105-32. Certain amino acids can be substituted by other amino acids having a similar hydrophobic index or score and still result in a protein with similar biological activity, i.e., still obtain a biological functionally equivalent protein. In making such changes, the substitution of amino acids whose hydrophobic indices are within +2 is preferred, those that are within +1 are particularly preferred, and those within +0.5 are even more particularly preferred;

Substitution of like amino acids on the basis of hydrophilicity. U.S. Pat. No. 4,554,101 states that the greatest

15

local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein. As detailed in this patent, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate (+3.0.+0.1); glutamate (+3.0.+0.1); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0); threonine (-0.4); proline (-0.5.+0.1); alanine (-0.5); histidine (-0.5); cysteine (-1.0); methionine (-1.3); valine (-1.5); leucine (-1.8); isoleucine (-1.8); tyrosine (-2.3); phenylalanine (-2.5); tryptophan (-3.4). Betts and Russell ((2003), "Amino Acid Properties and Consequences of Substitutions", Bioinformatics for Geneticists, Michael R. Barnes and Ian C. Gray, Eds., John Wiley & Sons, Ltd, Chapter 14, pp. 289-316) review the nature of mutations and the properties of amino acids in a variety of different protein contexts with the purpose of aiding in anticipating and interpreting the effect that a particular amino acid change will have on protein structure and function. The authors point out that features of proteins relevant to considering amino acid mutations include cellular environments, three-dimensional structure, and evolution, as well as the classifications of amino acids based on evolutionary, chemical, and structural principles, and the role for amino acids of different classes in protein structure and function in different contexts. The authors note that classification of amino acids into categories such as those shown in FIG. 14.3 of their review, which involves common physico-chemical properties, size, affinity for water (polar and non-polar; negative or positive charge), aromaticity and aliphaticity, hydrogen-bonding ability, propensity for sharply turning regions, etc., makes it clear that reliance on simple classifications can be dangerous, and suggests that alternative amino acids could be engineered into a protein at each position. Criteria for interpreting how a particular mutation might affect protein structure and function are summarized in section 14.7 of this review, and include first inquiring about the protein, and then about the particular amino acid substitution contemplated.

Biologically/enzymatically functional equivalents of the proteins and peptides disclosed herein can have 10 or fewer conservative amino acid changes, more preferably seven or fewer conservative amino acid changes, and most preferably five or fewer conservative amino acid changes, i.e., 10, 9, 8, 7, 6, 5, 4, 3, 2, or 1 conservative amino acid changes. The encoding nucleotide sequence (e.g., gene, plasmid DNA, cDNA, codon-optimized DNA, or other synthetic DNA) will thus have corresponding base substitutions, permitting it to code for the biologically functionally equivalent form of protein or peptide. Due to the degeneracy of the genetic code, i.e., the existence of more than one codon for most of the amino acids naturally occurring in proteins, other DNA (and RNA) sequences that contain essentially the same genetic information as these nucleic acids, and which encode the same amino acid sequence as that encoded by these nucleic acids, can be used in the methods disclosed herein. This principle applies as well to any of the other nucleotide sequences disclosed herein.

"Control" or "control level" means the level of a molecule, such as a polypeptide or nucleic acid, normally found in nature under a certain condition and/or in a specific genetic background. In certain embodiments, a control level of a molecule can be measured in a cell or specimen that has not been subjected, either directly or indirectly, to a treatment. A control level is also referred to as a wildtype or a basal level. These terms are understood by those of ordinary skill in the art. A control plant, i.e. a plant that does not contain a recombinant DNA that confers (for instance) an

16

enhanced trait in a transgenic plant, is used as a baseline for comparison to identify an enhanced trait in the transgenic plant. A suitable control plant may be a non-transgenic plant of the parental line used to generate a transgenic plant. A control plant may in some cases be a transgenic plant line that comprises an empty vector or marker gene, but does not contain the recombinant DNA, or does not contain all of the recombinant DNAs, in the test plant.

The terms "enhance", "enhanced", "increase", or "increased" refer to a statistically significant increase. For the avoidance of doubt, these terms generally refer to about a 5% increase in a given parameter or value, about a 10% increase, about a 15% increase, about a 20% increase, about a 25% increase, about a 30% increase, about a 35% increase, about a 40% increase, about a 45% increase, about a 50% increase, about a 55% increase, about a 60% increase, about a 65% increase, about 70% increase, about a 75% increase, about an 80% increase, about an 85% increase, about a 90% increase, about a 95% increase, about a 100% increase, or more over the control value. These terms also encompass ranges consisting of any lower indicated value to any higher indicated value, for example "from about 5% to about 50%", etc.

"Expression" or "expressing" refers to production of a functional product, such as, the generation of an RNA transcript from an introduced construct, an endogenous DNA sequence, or a stably incorporated heterologous DNA sequence. A nucleotide encoding sequence may comprise intervening sequence (e.g., introns) or may lack such intervening non-translated sequences (e.g., as in cDNA). Expressed genes include those that are transcribed into mRNA and then translated into protein and those that are transcribed into RNA but not translated (for example, siRNA, transfer RNA, and ribosomal RNA). The term may also refer to a polypeptide produced from an mRNA generated from any of the above DNA precursors. Thus, expression of a nucleic acid fragment, such as a gene or a promoter region of a gene, may refer to transcription of the nucleic acid fragment (e.g., transcription resulting in mRNA or other functional RNA) and/or translation of RNA into a precursor or mature protein (polypeptide), or both.

An "expression cassette" refers to a nucleic acid construct, which when introduced into a host cell, results in transcription and/or translation of a RNA or polypeptide, respectively.

The term "genome" as it applies to a plant cells encompasses not only chromosomal DNA found within the nucleus, but organelle DNA found within subcellular components (e.g., mitochondrial, plastid) of the cell. As used herein, the term "genome" refers to the nuclear genome unless indicated otherwise. However, expression in a plastid genome, e.g., a chloroplast genome, or targeting to a plastid genome such as a chloroplast via the use of a plastid targeting sequence, is also encompassed by the present disclosure.

The term "heterologous" refers to a nucleic acid fragment or protein that is foreign to its surroundings. In the context of a nucleic acid fragment, this is typically accomplished by introducing such fragment, derived from one source, into a different host. Heterologous nucleic acid fragments, such as coding sequences that have been inserted into a host organism, are not normally found in the genetic complement of the host organism. As used herein, the term "heterologous" also refers to a nucleic acid fragment derived from the same organism, but which is located in a different, e.g., non-native, location within the genome of this organism. Thus, the organism can have more than the usual number of

copy(ies) of such fragment located in its(their) normal position within the genome and in addition, in the case of plant cells, within different genomes within a cell, for example in the nuclear genome and within a plastid or mitochondrial genome as well. A nucleic acid fragment that is heterologous with respect to an organism into which it has been inserted or transferred is sometimes referred to as a "transgene."

A "heterologous" PGRL1 protein or CAB transit peptide protein-encoding nucleotide sequence, etc., can be one or more additional copies of an endogenous PGRL1 protein or CAB transit peptide protein-encoding nucleotide sequence, or a nucleotide sequence from another plant or other source. PGRL1 is a putative ferredoxin-plastoquinone reductase involved in photosynthetic cyclic electron flow. Furthermore, these can be genomic or non-genomic nucleotide sequences. Non-genomic nucleotide sequences encoding such proteins and peptides include, by way of non-limiting examples, mRNA; synthetically produced DNA including, for example, cDNA and codon-optimized sequences for efficient expression in different transgenic plants algae reflecting the pattern of codon usage in such plants; nucleotide sequences encoding the same proteins or peptides, but which are degenerate in accordance with the degeneracy of the genetic code; which contain conservative amino acid substitutions that do not adversely affect their activity, etc., as known by those of ordinary skill in the art.

The term "homology" describes a mathematically based comparison of sequence similarities which is used to identify genes or proteins with similar functions or motifs. The nucleic acid and protein sequences of the present invention can be used as a "query sequence" to perform a search against public databases to, for example, identify other family members, related sequences, or homologs. The term "homologous" refers to the relationship between two nucleic acid sequence and/or proteins that possess a "common evolutionary origin", including nucleic acids and/or proteins from superfamilies (e.g., the immunoglobulin superfamily) in the same species of animal, as well as homologous nucleic acids and/or proteins from different species of animal (for example, myosin light chain polypeptide, etc.; see Reeck et al., (1987) Cell, 50:667). Such proteins (and their encoding nucleic acids) may have sequence homology, as reflected by sequence similarity, whether in terms of percent identity or by the presence of specific residues or motifs and conserved positions. The methods disclosed herein contemplate the use of the presently disclosed nucleic and protein sequences, as well as sequences having sequence identity and/or similarity, and similar function.

"Host cell" means a cell which contains a vector and supports the replication and/or expression of the vector. Host cells may be prokaryotic cells such as *E. coli*, or eukaryotic cells such as yeast, insect, amphibian, or mammalian cells. Alternatively, the host cells are monocotyledonous or dicotyledonous plant cells.

The term "introduced" means providing a nucleic acid (e.g., an expression construct) or protein into a cell. "Introduced" includes reference to the incorporation of a nucleic acid into a eukaryotic or prokaryotic cell where the nucleic acid may be incorporated into the genome of the cell, and includes reference to the transient provision of a nucleic acid or protein to the cell. "Introduced" includes reference to stable or transient transformation methods, as well as sexually crossing. Thus, "introduced" in the context of inserting a nucleic acid fragment (e.g., a recombinant DNA construct/expression construct) into a cell, can mean "transfection" or "transformation" or "transduction", and includes reference

to the incorporation of a nucleic acid fragment into a eukaryotic or prokaryotic cell where the nucleic acid fragment may be incorporated into the genome of the cell (e.g., chromosome, plasmid, plastid, or mitochondrial DNA), converted into an autonomous replicon, or transiently expressed (e.g., transfected mRNA).

The term "isolated" refers to a material such as a nucleic acid molecule, polypeptide, or small molecule, that has been separated from the environment from which it was obtained. It can also mean altered from the natural state. For example, a polynucleotide or a polypeptide naturally present in a living animal is not "isolated" but the same polynucleotide or polypeptide separated from the coexisting materials of its natural state is "isolated", as the term is employed herein. Thus, a polypeptide or polynucleotide produced and/or contained within a recombinant host cell is considered isolated. Also intended as "isolated polypeptides" or "isolated nucleic acid molecules", etc., are polypeptides or nucleic acid molecules that have been purified, partially or substantially, from a recombinant host cell or from a native source.

As used herein, "nucleic acid" or "nucleotide sequence" means a polynucleotide (or oligonucleotide), including single or double-stranded polymers of deoxyribonucleotide or ribonucleotide bases, and unless otherwise indicated, encompasses naturally occurring and synthetic nucleotide analogues having the essential nature of natural nucleotides in that they hybridize to complementary single-stranded nucleic acids in a manner similar to naturally occurring nucleotides. Nucleic acids may also include fragments and modified nucleotide sequences. Nucleic acids disclosed herein can either be naturally occurring, for example genomic nucleic acids, or isolated, purified, non-genomic nucleic acids, including synthetically produced nucleic acid sequences such as those made by solid phase chemical oligonucleotide synthesis, enzymatic synthesis, or by recombinant methods, including for example, cDNA, codon-optimized sequences for efficient expression in different transgenic plants reflecting the pattern of codon usage in such plants, nucleotide sequences that differ from the nucleotide sequences disclosed herein due to the degeneracy of the genetic code but that still encode the protein(s) of interest disclosed herein, nucleotide sequences encoding the presently disclosed protein(s) comprising conservative (or non-conservative) amino acid substitutions that do not adversely affect their normal activity, PCR-amplified nucleotide sequences, and other non-genomic forms of nucleotide sequences familiar to those of ordinary skill in the art.

The protein-encoding nucleotide sequences, and promoter nucleotide sequences used to drive their expression, disclosed herein can be genomic or non-genomic nucleotide sequences. Non-genomic nucleotide protein-encoding sequences and promoters include, for example, naturally-occurring mRNA, synthetically produced mRNA, naturally-occurring DNA, or synthetically produced DNA. Synthetic nucleotide sequences can be produced by means well known in the art, including by chemical or enzymatic synthesis of oligonucleotides, and include, for example, cDNA, codon-optimized sequences for efficient expression in different transgenic plants and algae reflecting the pattern of codon usage in such organisms, variants containing conservative (or non-conservative) amino acid substitutions that do not adversely affect their normal activity, PCR-amplified nucleotide sequences, etc.

"A PGRL1 protein", "a PGR5 protein", "a HLA3 protein", "a CAB transit peptide", "a PGR5 transit peptide", or

any other protein or peptide presently broadly disclosed and utilized in any of the CCM methods and plants and algae disclosed herein refers to a protein or peptide exhibiting enzymatic/functional activity similar or identical to the enzymatic/functional activity of the specifically named protein or peptide. Enzymatic/functional activities of the proteins and peptides disclosed herein are described below. "Similar" enzymatic/functional activity of a protein or peptide can be in the range of from about 75% to about 125% or more of the enzymatic/functional activity of the specifically named protein or peptide when equal amounts of both proteins or peptides are assayed, tested, or expressed as described below under identical conditions, and can therefore be satisfactorily substituted for the specifically named proteins or peptides in the present enhanced CCM methods and transgenic plants and algae.

"Nucleic acid construct" or "construct" refers to an isolated polynucleotide which can be introduced into a host cell. This construct may comprise any combination of deoxyribonucleotides, ribonucleotides, and/or modified nucleotides. This construct may comprise an expression cassette that can be introduced into and expressed in a host cell.

"Operably linked" refers to a functional arrangement of elements. A first nucleic acid sequence is operably linked with a second nucleic acid sequence when the first nucleic acid sequence is placed in a functional relationship with the second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter effects the transcription or expression of the coding sequence. The control elements need not be contiguous with the coding sequence, so long as they function to direct the expression thereof. Thus, for example, intervening untranslated yet transcribed sequences can be present between a promoter and the coding sequence and the promoter can still be considered "operably linked" to the coding sequence.

The terms "plant" or "plants" that can be used in the present methods broadly include the classes of higher and lower plants amenable to transformation techniques, including angiosperms (monocotyledonous and dicotyledonous plants), gymnosperms, ferns, and unicellular and multicellular algae. The term "plant" also includes plants which have been modified by breeding, mutagenesis, or genetic engineering (transgenic and non-transgenic plants). It includes plants of a variety of ploidy levels, including aneuploid, polyploid, diploid, haploid, and hemizygous. The plant may be in any form including suspension cultures, embryos, meristematic regions, callus tissue, gametophytes, sporophytes, pollen, microspores, whole plants, shoot vegetative organs/structures (e.g. leaves, stems and tubers), roots, flowers and floral organs/structures, seed (including embryo, endosperm, and seed coat) and fruit, plant tissue (e.g. vascular tissue, ground tissue, and the like) and cells, and progeny of same.

Embodiments of the present disclosure also include parts of plants or algae, which can be selected from among a protoplast, a cell, a tissue, an organ, a cutting, an explant, a reproductive tissue, a vegetative tissue, biomass, an inflorescence, a flower, a sepal, a petal, a pistil, a stigma, a style, an ovary, an ovule, an embryo, a receptacle, a seed, a fruit, a stamen, a filament, an anther, a male or female gametophyte, a pollen grain, a meristem, a terminal bud, an axillary bud, a leaf, a stem, a root, a tuberous root, a rhizome, a tuber, a stolon, a corm, a bulb, an offset, a cell of said plant in culture, a tissue of said plant in culture, an organ of said plant in culture, a callus, propagation materials, germplasm, cuttings, divisions, and propagations.

Other embodiments include progeny or derivatives of transgenic plants and algae disclosed herein selected, for example, from among clones, hybrids, samples, seeds, and harvested material. Progeny can be asexually or sexually produced by methods well known in the art.

Useful C3 and C4 Plants

Plants to which the methods disclosed herein can be advantageously applied include both C3 and C4 plants, including "food crop" and "oilseed" plants, as well as algae.

10 Food Crop Plants

The term "food crop plant" refers to plants that are either directly edible, or which produce edible products, and that are customarily used to feed humans either directly, or indirectly through animals. Non-limiting examples of such plants include:

1. Cereal crops: wheat, rice, maize (corn), barley, oats, sorghum, rye, and millet;
2. Protein crops: peanuts, chickpeas, lentils, kidney beans, soybeans, lima beans;
- 20 3. Roots and tubers: potatoes, sweet potatoes, and cassavas;
4. Oil crops: soybeans, corn, canola, peanuts, palm, coconuts, safflower, cottonseed, sunflower, flax, olive, and safflower;
5. Sugar crops: sugar cane and sugar beets;
6. Fruit crops: bananas, oranges, apples, pears, breadfruit, pineapples, and cherries;
7. Vegetable crops and tubers: tomatoes, lettuce, carrots, melons, asparagus, etc.
- 30 8. Nuts: cashews, peanuts, walnuts, pistachio nuts, almonds;
9. Forage and turf grasses;
10. Forage legumes: alfalfa, clover;
11. Drug crops: coffee, cocoa, kola nut, poppy;
- 35 12. Spice and flavoring crops: vanilla, sage, thyme, anise, saffron, menthol, peppermint, spearmint, coriander.

In certain embodiments of this disclosure, the food crop plants are soybean, canola, tomato, potato, cassava, wheat, rice, oats, lettuce, broccoli, beets, sugar beets, beans, peas, kale, strawberry, and peanut.

40 "Oilseed Plants", "Oil Crop Plants", "Biofuels Crops", "Energy Crops"

The terms "oilseed plant" or "oil crop plant", and the like, to which the present methods and compositions can also be applied, refer to plants that produce seeds or fruit with oil content in the range of from about 1 to 2%, e.g., wheat, to about 20%, e.g., soybeans, to over 40%, e.g., sunflowers and rapeseed (canola). These include major and minor oil crops, as well as wild plant species which are used, or are being investigated and/or developed, as sources of biofuels due to their significant oil production and accumulation.

Exemplary oil seed or oil crop plants useful in practicing the methods disclosed herein include, but are not limited to, plants of the genera *Brassica* (e.g., rapeseed/canola (*Brassica napus*; *Brassica carinata*; *Brassica nigra*; *Brassica oleracea*), Camelina, *Misanthus*, and *Jatropha*; Jojoba (*Simmondsia chinensis*), coconut; cotton; peanut; rice; safflower; sesame; soybean; mustard; wheat; flax (linseed); sunflower; olive; corn; palm; palm kernel; sugarcane; castor bean; switchgrass; *Borage officinalis*; *Echium plantagineum*; *Cuphea hookeriana*; *Cuphea pulcherrima*; *Cuphea lanceolata*; *Ricinus communis*; *Coriandrum sativum*; *Crepis alpina*; *Vernonia galamensis*; *Momordica charantia*; and *Crambe abyssinica*).

60 65 A non-limiting example of a tuber that accumulates significant amounts of reserve lipids is the tuber of *Cyperus esculentus* (chufa or tigernuts), which has been proposed as

an oil crop for biofuel production. In the case of chufa, use of a constitutive or tuber-specific promoter would be useful in the methods disclosed herein.

Useful Algae

Algae useful in practicing various methods of the present disclosure include members of the following divisions: Chlorophyta and Heterokontophyta.

In certain embodiments, useful algae include members of the following classes: Chlorophyceae, Bacillariophyceae, Eustigmatophyceae, and Chrysophyceae. In certain embodiments, useful algae include members of the following genera: *Nannochloropsis*, *Chlorella*, *Dunaliella*, *Scenedesmus*, *Selenastrum*, *Oscillatoria*, *Phormidium*, *Spirulina*, *Amphora*, and *Ochromonas*. In one embodiment, members of the genus *Chlorella* are preferred.

Some algal species of particular interest include, without limitation: Bacillariophyceae strains, Chlorophyceae, Cyanophyceae, Xanthophyceae, Chrysophyceae, *Chlorella*, *Cryptothecodinium*, *Schizocystium*, *Nannochloropsis*, *Ulnenia*, *Dunaliella*, *Cyclotella*, *Navicula*, *Nitzschia*, *Cyclorella*, *Phaeodactylum*, and *Thaustochytrid*.

Non-limiting examples of algae species that can be used with the methods of the present disclosure include, for example, *Achnanthes orientalis*, *Agmenellum* spp., *Amphiprora hyaline*, *Amphora coffeeiformis*, *Amphora coffeeiformis* var. *linea*, *Amphora coffeeiformis* var. *punctata*, *Amphora coffeeiformis* var. *taylori*, *Amphora coffeeiformis* var. *tenuis*, *Amphora delicatissima*, *Amphora delicatissima* var. *capitata*, *Amphora* sp., *Anabaena*, *Ankistrodesmus*, *Ankistrodesmus falcatus*, *Boekelovia hooglandii*, *Borodinella* sp., *Botryococcus braunii*, *Botryococcus sudeticus*, *Bracteococcus minor*, *Bracteococcus medionucleatus*, *Carteria*, *Chaetoceros gracilis*, *Chaetoceros muelleri*, *Chaetoceros muelleri* var. *subsalsum*, *Chaetoceros* sp., *Chlamydomas perigranulata*, *Chlorella anitrita*, *Chlorella antarctica*, *Chlorella aureoviridis*, *Chlorella Candida*, *Chlorella capsulate*, *Chlorella desiccate*, *Chlorella ellipoidea*, *Chlorella emersonii*, *Chlorella fusca*, *Chlorella fusca* var. *vacuolata*, *Chlorella glucotropha*, *Chlorella infusionum*, *Chlorella infusionum* var. *actophila*, *Chlorella infusionum* var. *auxenophila*, *Chlorella kessleri*, *Chlorella lobophora*, *Chlorella luteoviridis*, *Chlorella luteoviridis* var. *aureoviridis*, *Chlorella luteoviridis* var. *lutescens*, *Chlorella miniata*, *Chlorella minutissima*, *Chlorella mutabilis*, *Chlorella nocturna*, *Chlorella ovalis*, *Chlorella parva*, *Chlorella photophilica*, *Chlorella pringsheimii*, *Chlorella protothecoides*, *Chlorella protothecoides* var. *acidicola*, *Chlorella regularis*, *Chlorella regularis* var. *minima*, *Chlorella regularis* var. *umbricata*, *Chlorella reisiglii*, *Chlorella saccharophila*, *Chlorella saccharophila* var. *ellipoidea*, *Chlorella salina*, *Chlorella simplex*, *Chlorella sorokiniana*, *Chlorella* sp., *Chlorella sphaerica*, *Chlorella stigmatophora*, *Chlorella vanniellii*, *Chlorella vulgaris*, *Chlorella vulgaris* fo. *tertia*, *Chlorella vulgaris* var. *autotrophica*, *Chlorella vulgaris* var. *viridis*, *Chlorella vulgaris* var. *vulgaris*, *Chlorella vulgaris* var. *vulgaris* fo. *tertia*, *Chlorella vulgaris* var. *vulgaris* fo. *viridis*, *Chlorella xanthella*, *Chlorella zofingiensis*, *Chlorella trebouxioides*, *Chlorella vulgaris*, *Chlorococcum infusionum*, *Chrysosphaera* sp., *Cricosphaera* sp., *Cryptothecodinium cohnii*, *Cryptomonas* sp., *Cyclotella cryptica*, *Cyclotella meneghiniana*, *Cyclotella* sp., *Chlamydomonas moewusii*, *Chlamydomonas reinhardtii*, *Chlamydomonas* sp., *Dunaliella* sp., *Dunaliella bardawil*, *Dunaliella bioculata*, *Dunaliella granulata*, *Dunaliella maritime*, *Dunaliella minuta*, *Dunaliella parva*, *Dunaliella peircei*, *Dunaliella primolecta*, *Dunaliella salina*, *Dunaliella terricola*, *Dunaliella tertio-*

lecta, *Dunaliella viridis*, *Dunaliella tertiolecta*, *Eremosphaera viridis*, *Eremosphaera* sp., *Ellipsoidon* sp., *Euglena* spp., *Franceia* sp., *Fragilaria crotonensis*, *Fragilaria* sp., *Gleocapsa* sp., *Gloeothamnion* sp., *Haematococcus pluvialis*, 5 *Hymenomonas* sp., *Isochrysis* aff. *galbana*, *Isochrysis galbana*, *Lepocinclis*, *Micractinium*, *Micractinium*, *Monoraphidium minutum*, *Monoraphidium* sp., *Nannochloropsis* sp., *Nannochloropsis* salina, *Nannochloropsis* sp., *Navicula acceptata*, *Navicula biskanterae*, *Navicula pseudo-tenelloides*, *Navicula pelliculosa*, *Navicula saprophila*, *Navicula* sp., *Nephrochloris* sp., *Nephroselmis* sp., *Nitzchia communis*, *Nitzschia alexandrina*, *Nitzschia closterium*, *Nitzschia communis*, *Nitzschia dissipata*, *Nitzschia frustulum*, *Nitzschia hantzschiana*, *Nitzschia inconspicua*, *Nitzschia intermedia*, *Nitzschia microcephala*, *Nitzschia pusilla*, *Nitzschia pusilla elliptica*, *Nitzschia pusilla monoensis*, *Nitzschia quadrangular*, *Nitzschia* sp., *Ochromonas* sp., 10 *Oocystis parva*, *Oocystis pusilla*, *Oocystis* sp., *Oscillatoria limnetica*, *Oscillatoria* sp., *Oscillatoria subbrevis*, *Parachlorella kessleri*, *Pascheria acidophila*, *Pavlova* sp., *Phaeodactylum tricornutum*, *Phagus*, *Phormidium*, *Platymonas* sp., *Pleurochrysis carterae*, *Pleurochrysis dentate*, *Pleurochrysis* sp., *Prototheca wickerhamii*, *Prototheca stagnora*, *Prototheca portoricensis*, *Prototheca moriformis*, *Prototheca zoppii*, *Pseudochlorella aquatica*, *Pyramimonas* sp., *Pyrobothrys*, *Rhodococcus opacus*, *Sarcinoid chrysophyte*, *Scenedesmus armatus*, *Schizochytrium*, *Spirogyra*, *Spirulina platensis*, *Stichococcus* sp., *Synechococcus* sp., *Synechocystis*, *Tagetes erecta*, *Tagetes patula*, *Tetradron*, *Tetraselmis* sp., *Tetraselmis suecica*, *Thalassiosira weissflogii*, and *Viridiella fridericiiana*.

In certain embodiments of this disclosure, the algae are species of *Chlorella*, *Nannochloropsis*, and *Chlamydomonas* listed above.

Exemplary food crop plant include wheat, rice, maize (corn), barley, oats, sorghum, rye, and millet; peanuts, chickpeas, lentils, kidney beans, soybeans, lima beans; potatoes, sweet potatoes, and cassavas; soybeans, corn, canola, peanuts, palm, coconuts, safflower, cottonseed, sunflower, flax, olive, and safflower; sugar cane and sugar beets; bananas, oranges, apples, pears, breadfruit, pineapples, and cherries; tomatoes, lettuce, carrots, melons, strawberry, asparagus, broccoli, peas, kale, cashews, peanuts, walnuts, pistachio nuts, almonds; forage and turf grasses; alfalfa, clover; coffee, cocoa, kola nut, poppy; vanilla, sage, thyme, anise, saffron, menthol, peppermint, spearmint and coriander and preferably wheat, rice and canola.

The terms “peptide”, “poly peptide”, and “protein” are used to refer to polymers of amino acid residues. These terms are specifically intended to cover naturally occurring biomolecules, as well as those that are recombinantly or synthetically produced, for example by solid phase synthesis.

The term “promoter” or “regulatory element” refers to a region or nucleic acid sequence located upstream or downstream from the start of transcription and which is involved in recognition and binding of RNA polymerase and/or other proteins to initiate transcription of RNA. Promoters need not be of plant or algal origin. For example, promoters derived from plant viruses, such as the CaMV35S promoter, or from other organisms, can be used in variations of the embodiments discussed herein. Promoters useful in the present methods include, for example, constitutive, strong, weak, tissue-specific, cell-type specific, seed-specific, inducible, repressible, and developmentally regulated promoters.

A skilled person appreciates that a promoter sequence can be modified to provide for a range of expression levels of an

operably linked heterologous nucleic acid molecule. Less than the entire promoter region can be utilized and the ability to drive expression retained. However, it is recognized that expression levels of mRNA can be decreased with deletions of portions of the promoter sequence. Thus, the promoter can be modified to be a weak or strong promoter. A promoter is classified as strong or weak according to its affinity for RNA polymerase (and/or sigma factor); this is related to how closely the promoter sequence resembles the ideal consensus sequence for the polymerase. Generally, by "weak promoter" is intended a promoter that drives expression of a coding sequence at a low level. By "low level" is intended levels of about 1/10,000 transcripts to about 1/100,000 transcripts to about 1/500,000 transcripts. Conversely, a strong promoter drives expression of a coding sequence at a high level, or at about 1/10 transcripts to about 1/100 transcripts to about 1/1,000 transcripts. The promoter of choice is preferably excised from its source by restriction enzymes, but can alternatively be PCR-amplified using primers that carry appropriate terminal restriction sites. It should be understood that the foregoing groups of promoters are non-limiting, and that one skilled in the art could employ other promoters that are not explicitly cited herein.

The term "purified" refers to material such as a nucleic acid, a protein, or a small molecule, which is substantially or essentially free from components which normally accompany or interact with the material as found in its naturally occurring environment, and/or which may optionally comprise material not found within the purified material's natural environment. The latter may occur when the material of interest is expressed or synthesized in a non-native environment. Nucleic acids and proteins that have been isolated include nucleic acids and proteins purified by standard purification methods. The term also encompasses nucleic acids and proteins prepared by recombinant expression in a host cell as well as chemically synthesized nucleic acids.

"Recombinant" refers to a nucleotide sequence, peptide, polypeptide, or protein, expression of which is engineered or manipulated using standard recombinant methodology. This term applies to both the methods and the resulting products. As used herein, a "recombinant construct", "expression construct", "chimeric construct", "construct" and "recombinant expression cassette" are used interchangeably herein.

As used herein, the phrase "sequence identity" or "sequence similarity" is the similarity between two (or more) nucleic acid sequences, or two (or more) amino acid sequences. Sequence identity is frequently measured as the percent of identical nucleotide or amino acid residues at corresponding positions in two or more sequences when the sequences are aligned to maximize sequence matching, i.e., taking into account gaps and insertions.

One of ordinary skill in the art will appreciate that sequence identity ranges are provided for guidance only. It is entirely possible that nucleic acid sequences that do not show a high degree of sequence identity can nevertheless encode amino acid sequences having similar functional activity. It is understood that changes in nucleic acid sequence can be made using the degeneracy of the genetic code to produce multiple nucleic acid molecules that all encode substantially the same protein. Means for making this adjustment are well-known to those of skill in the art. When percentage of sequence identity is used in reference to amino acid sequences it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g., charge or hydrophobicity) and therefore do not

change the functional properties of the molecule. Where sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences which differ by such conservative substitutions are said to have "sequence similarity" or "similarity". Means for making this adjustment are well-known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity.

"Percentage of sequence identity" is determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison and multiplying the result by 100 to yield the percentage of sequence identity.

Sequence identity (or similarity) can be readily calculated by known methods, including but not limited to those described in: Computational Molecular Biology, Lesk, A. M., ed., Oxford University Press, New York, 1988; Biocomputing: Informatics and Genome Projects, Smith, D. W., ed., Academic Press, New York, 1993; Computer Analysis of Sequence Data, Part I, Griffin, A. M., and Griffin, H. G., eds., Humana Press, New Jersey, 1994; Sequence Analysis in Molecular Biology, von Heijne, G., Academic Press, 1987; and Sequence Analysis Primer, Gribskov, M. and Devereux, J., eds., M Stockton Press, New York, 1991; and Carillo, H., and Lipman, D., SIAM J. Applied Math., 48: 1073 (1988). Methods to determine identity are designed to give the largest match between the sequences tested. Moreover, methods to determine identity are codified in publicly available computer programs. Optimal alignment of sequences for comparison can be conducted, for example, by the local homology algorithm of Smith & Waterman, by the homology alignment algorithms, by the search for similarity method or, by computerized implementations of these algorithms (GAP, BESTFIT, PASTA, and TFASTA in the GCG Wisconsin Package, available from Accelrys, Inc., San Diego, Calif., United States of America), or by visual inspection. See generally, (Altschul, S. F. et al., J. Mol. Biol. 215: 403-410 (1990) and Altschul et al. Nucl. Acids Res. 25: 3389-3402 (1997)).

One example of an algorithm that is suitable for determining percent sequence identity and sequence similarity is the BLAST algorithm, which is described in (Altschul, S., et al., NCBI NLM NIH Bethesda, Md. 20894; & Altschul, S., et al., J. Mol. Biol. 215: 403-410 (1990)). Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold. These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumula-

tive scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always >0) and N (penalty score for mismatching residues; always <0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value, the cumulative score goes to zero or below due to the accumulation of one or more negative-scoring residue alignments, or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff & Henikoff (1989) Proc. Natl. Acad. Sci. USA 89:10915).

In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin & Alt-schul, Proc. Nat'l. Acad. Sci. USA 90: 5873-5877 (1993)). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. BLAST searches assume that proteins can be modeled as random sequences. However, many real proteins comprise regions of nonrandom sequences which may be homopolymeric tracts, short-period repeats, or regions enriched in one or more amino acids. Such low-complexity regions may be aligned between unrelated proteins even though other regions of the protein are entirely dissimilar. A number of low-complexity filter programs can be employed to reduce such low-complexity alignments. For example, the SEG (Wooten and Federhen, Comput. Chem., 17: 149-163 (1993)) and XNU (Claverie and States, Comput. Chem., 17: 191-201 (1993)) low-complexity filters can be employed alone or in combination.

The constructs and methods disclosed herein encompass nucleic acid and protein sequences having sequence identity/sequence similarity at least about 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 100% to those specifically and/or sequences having the same or similar function for example if a protein or nucleic acid is identified with a transit peptide and the transit peptide is cleaved leaving the protein sequence without the transit peptide then the sequence identity/sequence similarity is compared to the protein with and/or without the transit peptide.

A “transgenic” organism, such as a transgenic plant, is a host organism that has been stably or transiently genetically engineered to contain one or more heterologous nucleic acid fragments, including nucleotide coding sequences, expression cassettes, vectors, etc. Introduction of heterologous nucleic acids into a host cell to create a transgenic cell is not limited to any particular mode of delivery, and includes, for example, microinjection, floral dip, adsorption, electroporation, vacuum infiltration, particle gun bombardment, whisksers-mediated transformation, liposome-mediated delivery, *Agrobacterium*-mediated transfer, the use of viral and retroviral vectors, etc., as is well known to those skilled in the art.

Conventional techniques of molecular biology, recombinant DNA technology, microbiology, and chemistry useful in practicing the methods of the present disclosure are

- described, for example, in Green and Sambrook (2012) Molecular Cloning: A Laboratory Manual, Fourth Edition, Cold Spring Harbor Laboratory Press; Ausubel et al. (2003 and periodic supplements) Current Protocols in Molecular Biology, John Wiley & Sons, New York, N.Y.; Amberg et al. (2005) Methods in Yeast Genetics: A Cold Spring Harbor Laboratory Course Manual, 2005 Edition, Cold Spring Harbor Laboratory Press; Roe et al. (1996) DNA Isolation and Sequencing: Essential Techniques, John Wiley & Sons; J. M. Polak and James O'D. McGee (1990) In Situ Hybridization: Principles and Practice; Oxford University Press; M. J. Gait (Editor) (1984) Oligonucleotide Synthesis: A Practical Approach, IRL Press; D. M. J. Lilley and J. E. Dahlberg (1992) Methods in Enzymology: DNA Structure Part A: Synthesis and Physical Analysis of DNA, Academic Press; and Lab Ref: A Handbook of Recipes, Reagents, and Other Reference Tools for Use at the Bench, Edited by Jane Roskams and Linda Rodgers (2002) Cold Spring Harbor Laboratory Press; Burgess and Deutscher (2009) Guide to Protein Purification, Second Edition (Methods in Enzymology, Vol. 463), Academic Press. Note also U.S. Pat. Nos. 8,178,339; 8,119,365; 8,043,842; 8,039,243; 7,303,906; 6,989,265; US20120219994A1; and EP1483367B1. The entire contents of each of these texts and patent documents are herein incorporated by reference.

Preliminary Results: Transgenic Plants Expressing Algal CCM Genes

Previously, reconstitution of a functional inorganic CCM in C3 plants to suppress photo-respiration and enhance photosynthesis was proposed. In WO 2012/125737, it was hypothesized that expression of a minimum of three algal CCM proteins would be sufficient to elevate internal plastid CO₂ concentrations high enough to suppress photorespiration. These three algal CCM genes included the *Chlamydomonas* plasma membrane-localized and ATP-dependent bicarbonate transporter, HLA3; the chloroplast envelope localized bicarbonate anion transporter, LCIA; and a chloroplast stromal-localized carbonic anhydrase (HCA-II) to accelerate conversion of bicarbonate into CO₂. These genes have individually been shown to be important to the CCM in prior studies ([3-5]). To test this hypothesis, we generated multiple independent transgenic *Arabidopsis* and Camelina plants expressing each CCM gene as a single gene construct, as well as a stacked 3-gene construct. The expression of each gene was controlled by the light-regulated Cab1 gene promoter [6].

The results of phenotypic analyses of *Arabidopsis* and Camelina plants transformed with the single CCM gene constructs were as follows:

HLA3 *Arabidopsis* transgenics varied in their phenotypes, but generally had reduced growth phenotypes relative to wild-type (WT) plants (FIG. 6). When the same plasmid was used to transform Camelina, no viable seeds were recovered from any transformation event after multiple attempts, indicating that HLA3 expression was likely toxic to Camelina.

With respect to carbonic anhydrase (CA) transgenics, we expressed a human carbonic anhydrase-2 (HCA2 (SEQ ID NO:17)) or a bacterial *Neisseria gonorrhoeae* carbonic anhydrase (BCA SEQ ID NO: 4) in the chloroplast stroma [7]. We choose these CAs because each has a turnover number (Kcat=106 sec-1) that is approximately 10x faster than plant/algal Cas. In both *Arabidopsis* and Camelina, we observed phenotypes that were either similar to WT (HCA2) or substantially larger (BCA) than WT plants (FIG. 3B).

Transgenic *Arabidopsis* plants expressing the LCIA gene were substantially impaired in growth (FIG. 5A). In contrast, Camelina LCIA transgenics grew better than WT, had

up to 25% higher photosynthetic rates at ambient CO₂ concentrations, and had reduced CO₂ compensation points (FIG. 5B).

The fact that expression of individual CCM genes impaired growth in C3 plants suggested that additional traits may need to be expressed or silenced to achieve optimal photosynthetic performance.

To determine if we could reconstitute a fully functional CCM complex in C3 plants, we transformed *Arabidopsis* and Camelina with a triple-gene CCM construct in which the expression of the HLA3, CA, and LCIA genes was driven by the green-tissue specific Cab1 promoter. In both *Arabidopsis* and Camelina there was either a substantial impairment in growth, or the plants did not survive (results not shown).

Thus, co-expression of the HLA3 gene with any other CCM gene(s) impaired growth even in plants in which expression of the other CCM genes, e.g., LCIA in Camelina, or BCA in *Arabidopsis*, enhanced growth. These results indicated that HLA3 expression was problematic.

Since the HLA3 protein catalyzes active bicarbonate transport and is the first-dedicated step in the engineered CCM, we re-focused our efforts on trying to determine why HLA3 expression was toxic to plants and how to mitigate its effects. We considered two possible hypotheses for HLA3 toxicity: 1) expression of the HLA3 ABC-transporter increases ATP demand (1 ATP/COO for photosynthesis by 25% and depletes cytoplasmic ATP levels [3-5,8] and 2) elevated bicarbonate levels in HLA3 transgenic plants negatively impact cytoplasmic pH levels. With respect to the latter hypothesis, it is noteworthy that unlike cyanobacteria, plants have robust cytoplasmic CA activity, potentially mitigating the effects of elevated bicarbonate levels on cytoplasmic pH.

The Role of ATP Demand and Cyclic Electron Transfer Activity in CCMs

In contrast to air-grown algae (4 ATP/2 NADPH/CO₂) and C4 plants (5 ATP/2 NADPH/CO₂) which have increased ATP demands for photosynthesis, C3 plants (3 ATP/2 NADPH/CO₂) have limited capacity to generate additional ATP for each electron transferred [8-10]. Increasing ATP demand by 25% per carbon fixed in HLA3 transgenic plants, therefore, could deplete cytoplasmic ATP levels as well as alter the redox state of the cell [8,10]. One mechanism to increase ATP synthesis for each light-driven electron transferred is by cyclic electron transfer (CET) activity. Light-driven CET is catalyzed by photosystem I (PSI) mediated charge separation leading to the reduction of ferredoxin (fd) and the PGR5 protein. The PGR5 protein reduces and protonates plastoquinone (PQ). PQH₂ is then oxidized by the cytochrome b6f complex (Cyt b6f). Protons released from the oxidation of PQH₂ drive ATP synthesis. The electron transfer cycle is completed by the reduction of plastocyanin (PC) by Cyt b6f, which in turn is oxidized by the PSI primary donor P700⁺. Significantly, molecular studies have demonstrated that genes encoding proteins functional in CET are substantially overexpressed (4-10 \times) in C4 plants and air-grown algae relative to related C3 species or high CO₂ grown algae [9,11-17]. These CET genes include: the Proton Gradient Regulation Genes PGR5 and PGRL1, and certain members of the Fd and ferredoxin NADP reductase (FNR) gene families [8-15]; Accession Nos.: PGR5:NM_126585; PGRL1: NM_179091; Fd: AtFd1: At1g10960; AtFd2:At1g60950; FNR: LFNR1:At5g66190;

LFRN2: At1g20020) [15]. The sequence for the PRG5 protein with the transit peptide amino acid sequence underlined is provided as

(SEQ ID NO: 1)
MAAASISAIG CNQTLIGTSF YGGWGSSISG EDYQTMLSKT
VAPPQQARVS RKAIRAVPMKVNNEGKGLF APLVVVTRNL
VGKKRQNQLR GKAIALHSQV ITEFCKSIGA
DAKQRQGLIRAKKNGERLG FL.

The transit peptide is cleaved to produce the functional PGR5 protein.

To test the hypothesis that ATP depletion in HLA3 transgenics resulted in growth impairment, we compared the phenotypes of VVT and HLA3 transgenics grown on nitrate which would require more linear electron transport (LET) to facilitate the reduction of nitrate. Significantly, the additional ATP produced by LET is not required for conversion of nitrate to ammonium and thus total ATP levels are expected to increase. In contrast, plants grown on ammonium do not require additional LET. Finally, we also grew transgenics on ammonium with sucrose which would presumably provide additional ATP via respiration [15,17]. We hypothesized that growth on nitrate or ammonium with sucrose would provide additional ATP that could potentially drive HLA3 activity.

As shown in FIG. 2B, none of the *Arabidopsis* HLA3 transgenics (4 independent lines) grew in the presence of ammonium, but all HLA3 lines were rescued when grown on ammonium with sucrose. Furthermore, plants grown on ammonium plus sucrose were phenotypically similar to VVT (FIG. 2B). In contrast, all HLA3 plants grown on nitrate survived, but some lines (#9, #20) had substantially impaired growth phenotypes. Identical results were observed for the germination and growth of VVT and HLA3 transgenic seeds on MS media agar plates using either nitrate (HLA3 transgenics survived) or ammonium (HLA3 transgenics died) as the sole nitrogen source (results not shown). Based on these observations, we propose that increased ATP synthesis associated with nitrate-driven LET and/or sucrose metabolism reduces the depletion of cytoplasmic ATP levels in HLA3 transgenics and rescues them.

This interpretation was corroborated by comparative metabolite analyses of leaf energy charge (EC) status (ATP), inorganic phosphate levels, and leaf reductive potential (RP) of VVT and HLA3 transgenic *Arabidopsis* grown on nitrate. As shown in FIG. 6, HLA3 transgenics grown on nitrate had reduced EC and RP ratios relative to WT. Energy charge is defined as $([ATP]+1/2[ADP])/([ATP]+[ADP]+[AMP])$. The reduction potential is a measurement of the capacity of the system to gain or lose electrons.

Significantly, inorganic phosphate levels were two-fold higher in HLA3 line #20, while the NADH level was two-fold lower than WT.

These results are consistent with the hypothesis that HLA3 expression places increased ATP demand on plants. This increased ATP demand in HLA3 transgenics may be met in part via NAD(P)H oxidation via the malate/oxaloacetate redox shunt between the mitochondria and chloroplasts [16].

LCIA Phenotype Depends on Plant Species

As previously indicated, LCIA expression in transgenic *Arabidopsis* resulted in plants with severely depressed growth phenotypes (FIG. 5A). In contrast, transgenic Camelina expressing LCIA had increased growth rates as well as

higher CO₂-dependent photosynthetic rates relative to WT (FIG. 5B). We propose that the substantially greater carbon sink-strength of Camelina relative to *Arabidopsis* accounts for the enhanced growth phenotype observed in Camelina LCIA plants. In support of this hypothesis, we observed that Camelina LCIA transgenics had higher CO₂-dependent rates of photosynthesis and lower CO₂ compensation points (40 vs. 53 ppm CO₂) than WT plants indicative of facilitated inorganic carbon uptake by LCIA (FIG. 5C).

Overview: Enhancing photosynthetic carbon fixation by increasing ATP production and limiting CO₂ diffusion out of artificial CCM lines; Strategies for facilitating CET and ATP synthesis in C3 plants

Prior attempts to subvert the limitations of photosynthesis have focused on engineering RuBisCO throughput and specificity [35] by introduction of engineered and non-native forms of the enzyme [36], through alterations in the regenerative capacity of the Calvin cycle [37,38] or by engineering photorespiratory bypasses [39]. These studies produced mixed results, thus advocating for a more comprehensive systems-level approach to enhance and/or redirect photosynthetic carbon flux.

As evidenced by our prior work described above, we postulate that both the carbon assimilatory steps and the light-based generation of ATP and NAPDH must be considered to develop a competent CCM with significantly improved photosynthetic capacity. To demonstrate proof of concept, an *Arabidopsis* line that contains a functional CCM that includes mechanisms to adjust ATP levels to meet transporter demand will be generated.

Enhancing CET and ATP Synthesis to Support HLA3-Dependent Bicarbonate Uptake

To exploit the expression of an algal CCM in C3 plants requires that we meet the additional energy demands required to actively transport inorganic carbon. As previously discussed in the section entitled “The role of ATP demand and cyclic electron transfer activity in CCMs”, C4 plants and algae have robust CET activity, and overexpress a variety of genes involved in CET [13,16,40-45] compared to C3 plants.

Several strategies are identified in the following examples, to increase ATP synthesis to support HLA3-dependent bicarbonate transport. Several of these strategies focus on elevating CET activity in C3 plants. Another approach involves the expression of a green photon-driven bacterial proton pump in thylakoids to supplement proton-driven ATP synthesis. Each approach is designed to complement existing CCM lines in *Arabidopsis*, Camelina, and potato we have created, and are evaluated based upon measured adenylate levels, plant biomass production, and photosynthetic measurements of carbon assimilation. The materials and methods employed in the examples below are for illustrative purposes only, and are not intended to limit the practice of the present embodiments thereto. Any materials and methods similar or equivalent to those described herein as would be apparent to one of ordinary skill in the art can be used in the testing or practice of the present embodiments, i.e., the materials, methods, and examples are illustrative only and not intended to be limiting.

Example 1: Enhancing CET Based on Overexpressing the Proton Gradient Regulatory Proteins PGR5 and PGRL1 in C3 Plants

Enhancing CET is based on overexpressing the proton gradient regulatory proteins PGR5 and/or PGRL1 which have previously been shown to be important to CET [37].

It has recently been demonstrated that the PGRL1 protein has antimycin A-sensitive (AA), ferredoxin-plastoquinone reductase (FQR) activity [46]. In *Chlamydomonas*, PGRL1 is part of the Cytb6f/PSI supercomplex which mediates CET. Significantly, PGRL1 forms homodimers as well as heterodimers with PGR5 via redox active cysteine residues. Under high-light conditions, thioredoxinred reduces PGRL1 dimers present in grana stacks, increasing the abundance of PGRL1 monomers and enhancing CET [47]. Mutational studies have shown that the PGR5 protein is required for Fd oxidation and PGRL1 reduction, but not for PQ reduction. In addition, it has been shown that PGRL1/PGR5 heterodimers are more active in CET than PGRL1 monomers. In C4 plants PGR5 and PGRL1 expression levels are elevated (4×) relative to C3 plants [9]. Similarly, PGR5 expression is up-regulated in air-grown *Chlamydomonas* (active CCM and HLA3 activity) relative to high CO₂ (low CCM) grown cells [16,43]. Significantly, overexpression of PGRL1 and PGR5 has also been shown to increase AA-sensitive CET in transgenic *Arabidopsis* [48]. One embodiment of the present invention provides for an overexpression of PGRL1 gene (SEQ ID NO:106) and PGR5 gene with chloroplast targeting sequence (SEQ ID NO:2) with HLA3 gene (SEQ ID NO:12) or with HLA3 gene (SEQ ID NO:12) and LCIA gene (SEQ ID NO:16) and BCA gene codon optimized for expression in *Arabidopsis* (SEQ ID NO:4) to yield substantially increased photosynthetic rates, particularly in plants with enhanced sink strength (Camelina and potato for example). Co-expression of the PGR5 gene (SEQ ID NO:2) along with the HLA3 gene (SEQ ID NO:12) in Camelina rescued the HLA3 gene and it was no longer lethal. These results indicate that the PGR5 gene is enabling the production of sufficient ATP to meet the demands of the HLA3 gene product.

HLA3 (SEQ ID NO:12) and PGR5 (SEQ ID NO:2) are introduced as a double construct into *Arabidopsis* or Camelina, by *Agrobacterium*-mediated Ti plasmid transformation using, for example, plasmid pB110-HLA3-pgr5-dsred (FIG. 9). Since PGR5 protein (SEQ ID NO:1) is naturally targeted to the thylakoid membranes, no additional targeting sequences are introduced. Similarly, since HLA3 protein (SEQ ID NO:77) is naturally targeted to the chloroplast envelope, no additional targeting sequences are added. HLA3 is codon optimized for plant expression.

In one embodiment, the expression of each protein is driven by the light sensitive leaf-specific CAB1 promoter (SEQ ID NO:7) and Nos terminator (SEQ ID NO:9) (FIG. 9).

The BCA gene (AAW89307; SEQ ID NO:4), under the control of CAB1 promoter, is introduced to *Arabidopsis* by *Agrobacterium*-mediated Ti plasmid transformation by floral dip method using the construct shown in FIG. 10.

As a visual marker, the plasmid also includes a gene for expression of fluorescent DsRed protein under the control of CVMV promoter and Nos terminator (FIG. 10).

Plants are transformed by vacuum infiltration method (Lu and Kang (February, 2008) Plant Cell Rep. 27(2):273-8), and will be screened for biomass yield parameters (including plant weight, height, branching and seed yield) and photosynthetic efficiency measured as CO₂ absorption with the aid of a LiCor 6400 gas exchange analyzer.

The PGRL1 gene from *Arabidopsis* (NM_179091 SEQ ID NO:3) will be subcloned into pCambia1301-based binary plasmid under control of the CAB1 promoter (SEQ ID NO:7) and Nos terminator (SEQ ID NO:9). The plasmid will also carry a gene for hygromycin selection marker. *Agrobacterium*-mediated transformation takes place by the stan-

dard floral dip method followed by germination of seeds on hygromycin to select for transformants. The expression of PRGL1 will be confirmed by RT-PCR, and the resulting transgenic plant lines will be crossed with HLA3/PGR5 plants and screened for biomass yield and photosynthesis rate (CO_2 fixation).

Example 2: Determining if Fd1 Gene Overexpression can Support Algal CCM and Increased Photosynthetic Rates

It has recently been demonstrated that specific members of the ferredoxin (Fd) gene family facilitate CET. Overexpression of pea ferredoxin1 (Fd1) enhanced CET at the expense of LET in tobacco [16,40].

Therefore, another embodiment of the present invention provides enhancing ATP production and titrating the expression of the pea Fd1 gene in the three model C3 plants with and without co-expression of the CCM genes to determine if Fd1 overexpression can support the algal CCM and increased photosynthetic rates. Earlier results demonstrated that Fd1 overexpression slightly impaired Linear Electron Transfer (LET), resulting in a stunted phenotype [40]. We expect that the additional ATP demand in HLA3 transgenics, however, will mitigate these effects.

Fd1 gene (At1g10960) will be introduced by *Agrobacterium*-mediated Ti plasmid transformation. Fd1 gene will be subcloned into pCambia1301-based binary plasmid under control of CAB1 promoter (SEQ ID NO:7) and Nos terminator (SEQ ID NO:9). The plasmid will also carry a gene for hygromycin selection as a marker. *Agrobacterium*-mediated transformation takes place by the standard floral dip method, followed by germination of seeds on hygromycin to select for transformants. The expression of FD1 (SEQ ID NO:93) will be confirmed by real time QPCR, and the resulting plant lines exhibiting different levels of FD1 expression will be crossed with CCM-expressing plants and screened for biomass yield and photosynthesis rate with the aid of a LiCor 6400 CO_2 -gas exchange analyzer.

Example 3: Overexpression of Unique Ferredoxin NADP Reductase (FNR) Gene Family Members Associated with CET

Yet another embodiment is based on overexpression of unique ferredoxin NADP reductase (FNR) gene family members associated with CET. Leaf FNR (LFNR) catalyzes the reduction of Fd and is involved in both LET and CET [15]. It was recently demonstrated that there are three LFNR gene family members expressed in maize leaves: Accession Nos. BAA88236 (LFNR1), BAA88237 (LFNR2), and ACF85815 (LFNR3).

LFNR-1 was shown to be localized to thylakoid membranes and associated with Cytb6f complexes. LFNR2 was present in thylakoids and stroma associated with Cytb6f complexes. LFNR3 was soluble and not associated with Cytb6f complexes.

Significantly, when plants were grown with nitrate instead of ammonium, expression of LFNR1 and LFNR2 was elevated but not that of LFNR3. In contrast, studies using *Arabidopsis* LFNR1 knock out mutants demonstrated that PGA-dependent oxygen evolution (which requires additional ATP) is more negatively affected than is nitrate-dependent oxygen evolution (no additional ATP demand),

suggesting that LFNR1 may play a role in regulating CET [15]. However, this interpretation remains equivocal.

To determine if CET activity and HLA3 mediated inorganic carbon uptake can be altered by differential expression of LFNR1, we will both over-express (CAB1 promoter (SEQ ID NO:7)) and under-express (LFNR1 RNAi) LFNR1 in transgenic *Arabidopsis* to determine the impact of altered LFNR1 expression on functional CCM activity.

For overexpression of the LFNR1, the gene (At5g66190) will be introduced by *Agrobacterium*-mediated Ti plasmid transformation by floral dipping. The LFNR1 gene will be subcloned into pCambia1301-based binary plasmid under control of the CAB1 promoter (SEQ ID NO:7) and Nos terminator (SEQ ID NO:9). The plasmid will also carry a gene for hygromycin selection as a marker. The expression of LFNR1 will be confirmed by real time QPCR, the resulting plant lines will be crossed with CCM-expressing plants, and screened for biomass yield and photosynthesis rate with the aid of a LiCor 6400 CO_2 -gas exchange analyzer.

For downregulation of the LFNR1 levels, an RNAi construct containing a partial sequence of the LFNR1 (At5g66190 or BAA88236) and reverse complementary sequence of LFNR1 will be subcloned into pCambia1301-based binary plasmid under control of the CAB1 promoter (SEQ ID NO:7) and Nos terminator (SEQ ID NO:9). The plasmid will also carry a gene for hygromycin selection as a marker. The reduced level of LFNR1 expression will be confirmed by real time QPCR.

The resulting lines will be crossed with CCM-expressing lines to generate double mutants. Those mutants will be screened for biomass yield parameters (including plant weight, height, branching and seed yield) and photosynthetic efficiency measured as CO_2 absorption with the aid of a LiCor 6400 gas exchange analyzer.

Example 4: Facilitated Vectoral Proton Transport Using Proteorhodopsin (PR)

In yet another embodiment green photons, not absorbed by chlorophyll, to drive proton transport across thylakoids by expressing modified PR [49] will be employed to enhance ATP synthesis (FIG. 7).

PR is a seven-helix transmembrane-spanning protein similar to bacteriorhodopsin that contains retinal in its active site. Green light-driven cis-trans isomerization of retinal drives vectoral proton transfer across the membrane [50-55]. Significantly, it has been demonstrated that a functional PR could be expressed in a respiration-impaired mutant of *E. coli* when supplemented with exogenous all-trans retinal [56]. More recently, hydrogen production was shown to increase nearly two-fold in PR-expressing *E. coli* when cells were exposed to increasing light intensities (70 to 130 μE), indicating that PR can efficiently absorb light even at low intensities [57]. To the best of our knowledge, retinal complementation of other rhodopsins has not been reported. Significantly, PR-expressing *E. coli* respiratory mutants generated sufficient proton-motive force to support ATP synthesis levels, leading to enhanced cell viability and motility when transgenics were exposed to sunlight as the only energy source.

These results suggest that targeting PR to the thylakoid membrane using appropriate targeting sequences (e.g., nuclear-encoded, N-terminal, light harvesting complex signal sequences) and supplementation with exogenous retinal or retinal derived from β -carotene cleavage) could drive additional ATP synthesis. One concern is that the optical

cross section of retinal is small and light harvesting by PR is not supplemented by antenna complexes. This constraint may be overcome in part by overexpressing PR in thylakoids. Regardless, the additional proton gradient necessary to support HLA3 activity is substantially less than that required to support overall CO₂ fixation. The best achievable PR expression levels will be determined empirically using different gene promoters, e.g., psaD (SEQ ID NO:10), rbcS (SEQ ID NO:11), and cab1 (SEQ ID NO:7), to drive its expression.

Generation of Improved PR and its Functional Reconstitution in Chloroplasts

PR (AF279106), for example (SEQ ID NO:98), will be introduced into *Arabidopsis*, Camelina, and potato by Ti plasmid transformation and targeted to the thylakoid membrane using the DNAJ transit peptide (At5g21430, SEQ ID NO: 22) or psbX stop-transfer trans-membrane domain (At2g06520 SEQ ID NO:23) fused to the C-terminus of PR [58], or transit peptides from nuclear encoded chloroplast proteins such as CAB (SEQ ID NO:13), PGR5 (SEQ ID NO:14), and psaD (SEQ ID NO:15). Reconstitution with exogenous retinal will be carried out in a manner similar to strategies described for *E. coli*, except that retinal will be painted on the surface of the leaf [56] to demonstrate proof of concept. Retinal reconstitution will be followed by monitoring the absorption of the thylakoid membranes at 540 nm [59].

If exogenously applied retinal is not incorporated into PR, we will express low levels of a plant codon-optimized β-carotene monooxygenase for example (SEQ ID NO:100) in plastids to cleave a small fraction of β-carotene to generate retinal. Non-limiting examples of β-carotene monooxygenases that can be used include, for example, mouse, human, zebra fish, and rat enzymes (Accession Nos. AW044715, AK001592, AJ290390, and NM_053648, respectively). Alternatively, if β-carotene levels are severely depleted, we will transiently express β-carotene monooxygenase under the control of a transient inducible promoter such as an ethanol inducible gene promoter. This is available as an EcoRI/PstI fragment from Syngenta-Construct: pJL67-5S::AlcR/AlcA::GUS in pMLBART (Weigel World, Max Planck Institute for Developmental Biology, Tubingen, Germany) for periods of time sufficient to fully saturate PR [60,61]. Operation of a functional retinal photocycle in PR will be confirmed by transient absorption spectroscopy [62].

Alternatively, promoters such as the green tissue/leaf-specific promoters such as the CAB (At3g54890 SEQ ID NO:7) and rbcS (At5g38420 SEQ ID NO:11) promoters can be used, for example see SEQ ID NO:5 for the BCA protein with a rbc-1a transit peptide. As the skilled person will be well aware, various promoters may be used to promote the transcription of the nucleic acid of the invention, i.e. the nucleic acid which when transcribed yields an RNA molecule that modulates the expression and/or activity of a protein according to the invention. Such promoters include for example constitutive promoters, inducible promoters (e.g. light inducible promoters, stress-inducible promoters, drought-inducible promoters, hormone-inducible promoters, chemical-inducible promoters, etc.), tissue-specific promoters, developmentally regulated promoters and the like.

Thus, a plant expressible promoter can be a constitutive promoter, i.e. a promoter capable of directing high levels of expression in most cell types (in a spatio-temporal independent manner). Examples of plant expressible constitutive promoters include promoters of bacterial origin, such as the octopine synthase (OCS) and nopaline synthase (NOS) promoters from *Agrobacterium*, but also promoters of viral

origin, such as that of the cauliflower mosaic virus (CaMV) 35S transcript (Hapster et al., 1988, Mol. Gen. Genet. 212: 182-190) or 19S RNAs genes (Odell et al., 1985, Nature. 6; 313(6005):810-2; U.S. Pat. No. 5,352,605; WO 84/02913; Benfey et al., 1989, EMBO J. 8:2195-2202), the enhanced 2x35S promoter (Kay et al., 1987, Science 236:1299-1302; Datla et al. (1993), Plant Sci 94:139-149) promoters of the cassava vein mosaic virus (CsVMV; WO 97/48819, U.S. Pat. No. 7,053,205), 2xCsVMV (WO2004/053135) the circovirus (AU 689 311) promoter, the sugarcane bacilliform badnavirus (ScBV) promoter (Samac et al., 2004, Transgenic Res. 13(4):349-61), the figwort mosaic virus (FMV) promoter (Sanger et al., 1990, Plant Mol Biol. 14(3):433-43), the subterranean clover virus promoter No 4 or No 7 (WO 96/06932) and the enhanced 35S promoter as described in U.S. Pat. Nos. 5,164,316, 5,196,525, 5,322,938, 5,359,142 and 5,424,200. Among the promoters of plant origin, mention will be made of the promoters of the promoter of the *Arabidopsis thaliana* histone H4 gene (Chabouté et al., 1987), the ubiquitin promoters (Holtorf et al., 1995, Plant Mol. Biol. 29:637-649, U.S. Pat. No. 5,510,474) of Maize, Rice and sugarcane, the Rice actin 1 promoter (Act-1, U.S. Pat. No. 5,641,876), the histone promoters as described in EP 0 507 698 A1, the Maize alcohol dehydrogenase 1 promoter (Adh-1) (from the world wide web at patentlens.net/daisy/promoters/242.html)).

A variety of plant gene promoters that regulate gene expression in response to environmental, hormonal, chemical, developmental signals, and in a tissue-active manner can be used for expression of a sequence in plants. Choice of a promoter is based largely on the phenotype of interest and is determined by such factors as tissue (e.g., seed, fruit, root, pollen, vascular tissue, flower, carpel, etc.), inducibility (e.g., in response to heat, cold, drought, light etc.), timing, developmental stage, and the like.

Promoters that can be used to practice this invention include those that are green tissue specific such as the promoter of light harvesting complex protein 2 (Sakamoto et al. Plant Cell Physiology, 1991, 32(3): 385-393) or the promoter of the cytosolic fructose-1, 6-bisphosphatase from rice (Si et al. Acta Botanica Sinica 45: 3(2003): 359-364). Alternative embodiments include light inducible promoters such as promoters of the plant ribulose-biscarboxylase/oxygenase (Rubisco) small subunit promoter (U.S. Pat. No. 4,962,028; WO99/25842) from *Zea mays* and sunflower. Also the small subunit promoter from *Chrysanthemum* may be used, combined or not combined with the use of the respective terminator (Outchkourov et al., Planta, 216: 1003-1012, 2003).

Additional promoters that can be used to practice this invention are those that elicit expression in response to stresses, such as the RD29 promoters that are activated in response to drought, low temperature, salt stress, or exposure to ABA (Yamaguchi-Shinozaki et al., 2004, Plant Cell, Vol. 6, 251-264; WO12/101118), but also promoters that are induced in response to heat (e.g., see Ainley et al. (1993) Plant Mol. Biol. 22: 13-23), light (e.g., the pea rbcS-3A promoter, Kuhlemeier et al. (1989) Plant Cell 1: 471-478, and the maize rbcS promoter, Schaffner and Sheen (1991) Plant Cell 3: 997-1012); wounding (e.g., wun1, Siebertz et al. (1989) Plant Cell 1: 961-968); pathogens (such as the PR-I promoter described in Buchel et al. (1999) Plant Mol. Biol. 40: 387-396, and the PDF 1.2 promoter described in Manners et al. (1998) Plant Mol. Biol. 38: 1071-1080), and chemicals such as methyl jasmonate or salicylic acid (e.g., see Gatz (1997) Annu. Rev. Plant Physiol. Plant Mol. Biol. 48: 89-108). In addition, the timing of the expression can be

controlled by using promoters such as those acting at senescence (e.g., see Gan and Amasino (1995) *Science* 270: 1986-1988); or late seed development (e.g., see Odell et al. (1994) *Plant Physiol.* 106: 447-458).

Use may also be made of salt-inducible promoters such as the salt-inducible NHX1 promoter of rice landrace Pokkali (PKN) (Jahan et al., 6th International Rice Genetics symposium, 2009, poster abstract P4-37), the salt inducible promoter of the vacuolar H⁺-pyrophosphatase from *Thellungiella halophila* (TsVP1) (Sun et al., *BMC Plant Biology* 2010, 10:90), the salt-inducible promoter of the *Citrus sinensis* gene encoding phospholipid hydroperoxide isoform gpx1 (Avsian-Kretchmer et al., *Plant Physiology* July 2004 vol. 135, p 1685-1696).

In alternative embodiments, tissue-specific and/or developmental stage-specific promoters are used, e.g., promoter that can promote transcription only within a certain time frame of developmental stage within that tissue. See, e.g., Blazquez (1998) *Plant Cell* 10:791-800, characterizing the *Arabidopsis* LEAFY gene promoter. See also Cardon (1997) *Plant J* 12:367-77, describing the transcription factor SPL3, which recognizes a conserved sequence motif in the promoter region of the *A. thaliana* floral meristem identity gene API; and Mandel (1995) *Plant Molecular Biology*, Vol. 29, pp 995-1004, describing the meristem promoter eIF4. Tissue specific promoters which are active throughout the life cycle of a particular tissue can be used. Other promoters that can be used to express the nucleic acids of the invention include, a leaf-specific promoter (see, e.g., Busk (1997) *Plant J*. 11:1285 1295, describing a leaf-specific promoter in maize); a tomato promoter active during fruit ripening, senescence and abscission of leaves, a guard-cell preferential promoter e.g. as described in PCT/EP12/065608, and, to a lesser extent, of flowers can be used (see, e.g., Blume (1997) *Plant J*. 12:731 746); the Blec4 gene from pea, which is active in epidermal tissue of vegetative and floral shoot apices of transgenic alfalfa making it a useful tool to target the expression of foreign genes to the epidermal layer of actively growing shoots or fibers; the ovule-specific BELI gene (see, e.g., Reiser (1995) *Cell* 83:735-742, GenBank No. U39944); and/or, the promoter in Klee, U.S. Pat. No. 5,589,583, describing a plant promoter region is capable of conferring high levels of transcription in meristematic tissue and/or rapidly dividing cells. Further tissue specific promoters that may be used according to the invention include, promoters active in vascular tissue (e.g., see Ringli and Keller (1998) *Plant Mol. Biol.* 37: 977-988), carpels (e.g., see Ohl et al. (1990) *Plant Cell* 2. In alternative embodiments, plant promoters which are inducible upon exposure to plant hormones, such as auxins, are used to express the nucleic acids used to practice the invention. For example, the invention can use the auxin-response elements EI promoter fragment (AuxREs) in the soybean (*Glycine max* L.) (Liu (1997) *Plant Physiol.* 115:397-407); the auxin-responsive *Arabidopsis* GST6 promoter (also responsive to salicylic acid and hydrogen peroxide) (Chen (1996) *Plant J*. 10: 955-966); the auxin-inducible parC promoter from tobacco (Sakai (1996) 37:906-913); a plant biotin response element (Streit (1997) *Mol. Plant Microbe Interact.* 10:933-937); and, the promoter responsive to the stress hormone abscisic acid (ABA) (Sheen (1996) *Science* 274:1900-1902). Further hormone inducible promoters that may be used include auxin-inducible promoters (such as that described in van der Kop et al. (1999) *Plant Mol. Biol.* 39: 979-990 or Baumann et al., (1999) *Plant Cell* 11: 323-334), cytokinin-inducible promoter (e.g., see Guevara-Garcia (1998) *Plant Mol. Biol.* 38: 743-753), promoters responsive to gibberellin (e.g., see

Shi et al. (1998) *Plant Mol. Biol.* 38: 1053-1060, Willmott et al. (1998) *Plant Molec. Biol.* 38: 817-825) and the like.

In alternative embodiments, nucleic acids used to practice the invention can also be operably linked to plant promoters which are inducible upon exposure to chemicals reagents which can be applied to the plant, such as herbicides or antibiotics. For example, the maize In2-2 promoter, activated by benzenesulfonamide herbicide safeners, can be used (De Veylder (1997) *Plant Cell Physiol.* 38:568-577); application of different herbicide safeners induces distinct gene expression patterns, including expression in the root, hydathodes, and the shoot apical meristem. Coding sequence can be under the control of, e.g., a tetracycline-inducible promoter, e.g., as described with transgenic tobacco plants containing the *Avena sativa* L. (oat) arginine decarboxylase gene (Masgrau (1997) *Plant J*. 11:465-473); or, a salicylic acid-responsive element (Stange (1997) *Plant J*. 11:1315-1324). Using chemically- {e.g., hormone- or pesticide} induced promoters, i.e., promoter responsive to a chemical which can be applied to the transgenic plant in the field, expression of a polypeptide of the invention can be induced at a particular stage of development of the plant. Use may also be made of the estrogen-inducible expression system as described in U.S. Pat. No. 6,784,340 and Zuo et al. (2000, *Plant J*. 24: 265-273) to drive the expression of the nucleic acids used to practice the invention.

In alternative embodiments, a promoter may be used whose host range is limited to target plant species, such as corn, rice, barley, wheat, potato or other crops, inducible at any stage of development of the crop.

In alternative embodiments, a tissue-specific plant promoter may drive expression of operably linked sequences in tissues other than the target tissue. In alternative embodiments, a tissue-specific promoter that drives expression preferentially in the target tissue or cell type, but may also lead to some expression in other tissues as well, is used.

According to the invention, use may also be made, in combination with the promoter, of other regulatory sequences, which are located between the promoter and the coding sequence, such as transcription activators ("enhancers"), for instance the translation activator of the tobacco mosaic virus (TMV) described in Application WO 87/07644, or of the tobacco etch virus (TEV) described by Carrington & Freed 1990, *J. Virol.* 64: 1590-1597, for example.

Other regulatory sequences that enhance the expression of the nucleic acid of the invention may also be located within the chimeric gene. One example of such regulatory sequences is introns. Introns are intervening sequences present in the pre-mRNA but absent in the mature RNA following excision by a precise splicing mechanism. The ability of natural introns to enhance gene expression, a process referred to as intron-mediated enhancement (IME), has been known in various organisms, including mammals, insects, nematodes and plants (WO 07/098042, p 11-12). IME is generally described as a posttranscriptional mechanism leading to increased gene expression by stabilization of the transcript. The intron is required to be positioned between the promoter and the coding sequence in the normal orientation. However, some introns have also been described to affect translation, to function as promoters or as position and orientation independent transcriptional enhancers (Chaubet-Gigot et al., 2001, *Plant Mol Biol.* 45(1):17-30, p 27-28).

Examples of genes containing such introns include the 5' introns from the rice actin 1 gene (see U.S. Pat. No. 5,641,876), the rice actin 2 gene, the maize sucrose synthase gene (Clancy and Hannah, 2002, *Plant Physiol.* 130(2):918-

29), the maize alcohol dehydrogenase-1 (Adh-1) and Bronze-1 genes (Callis et al. 1987 *Genes Dev.* 1(10):1183-200; Mascarenhas et al. 1990, *Plant Mol Biol.* 15(6):913-20), the maize heat shock protein 70 gene (see U.S. Pat. No. 5,593,874), the maize shrunken 1 gene, the light sensitive 1 gene of *Solanum tuberosum*, and the heat shock protein 70 gene of *Petunia hybrida* (see U.S. Pat. No. 5,659,122), the replacement histone H3 gene from alfalfa (Keleman et al. 2002 *Transgenic Res.* 11(1):69-72) and either replacement histone H3 (histone H3.3-like) gene of *Arabidopsis thaliana* (Chaubet-Gigot et al., 2001, *Plant Mol Biol.* 45(1):17-30).

Other suitable regulatory sequences include 5' UTRs. As used herein, a 5' UTR, also referred to as a leader sequence, is a particular region of a messenger RNA (mRNA) located between the transcription start site and the start codon of the coding region. It is involved in mRNA stability and translation efficiency. For example, the 5' untranslated leader of a petunia chlorophyll a/b binding protein gene downstream of the 35S transcription start site can be utilized to augment steady-state levels of reporter gene expression (Harpster et al., 1988, *Mol Gen Genet.* 212(1):182-90). WO95/006742 describes the use of 5' non-translated leader sequences derived from genes coding for heat shock proteins to increase transgene expression.

The chimeric gene may also comprise a 3' end region, i.e. a transcription termination or polyadenylation sequence, operable in plant cells. As a transcription termination or polyadenylation sequence, use may be made of any corresponding sequence of bacterial origin, such as for example the nos terminator of *Agrobacterium tumefaciens*, of viral origin, such as for example the CaMV 35S terminator, or of plant origin, such as for example a histone terminator as described in published Patent Application EP 0 633 317 A1. The polyadenylation region can be derived from the natural gene, from a variety of other plant genes, or from T-DNA. The 3' end sequence to be added may be derived from, for example, the nopaline synthase or octopine synthase genes, or alternatively from another plant gene, or less preferably from any other eukaryotic gene.

The expression and targeting of proteorhodopsin to the thylakoid membranes will take advantage of the green energy spectrum that is inaccessible to chlorophyll. An increase in the amount of ATP is expected under photosynthesis conditions, from proton gradient generated both by the photosystems and the proteorhodopsin pump. Under conditions of inhibition of electron transfer through the photosystems, we should be able to observe a steady rate of ATP synthesis well above the basal rate through the activity of the proteorhodopsin proton pump.

Under normal pH conditions, protons are pumped into the bacterial periplasmic space by PR [50]. The photo-driven retinal cycle begins with photoisomerization of all trans-retinal to 13-cis retinal. The resulting conformational change poises the system for transfer of a proton from the Schiff base (SB; pKa~11) to the counter ion, Asp 97 (pKa~7.5). The proton is transferred to the lumen via a proton-conducting channel, and the SB is reprotonated from the cytoplasm. The mechanism of proton release in PR is not as well understood as in bacteriorhodopsin (BR); however, the main events of the photocycle are expected to be similar to those of BR. One potential challenge for pumping protons by PR in thylakoid membranes is the pH gradient-dependent reversibility of proton transfer by PR. At periplasmic pHs, <5.5, proton flow in PR is reversed, potentially depleting the proton gradient and impairing ATP synthesis. Thus, at the luminal pH of thylakoids (4.5), reversed proton transduction via PR is possible. One of the critical residues

involved in reversible proton flow is Asp97, which acts as the proton acceptor from retinal. The pKa of Asp97 in PR is ~7.5, while the pKa of its counterpart in BR is ~2.5. Due to the extremely low pKa of the counter ion, BR is able to retain its forward pumping activity at pHs as low as 3.5. The ability of PR to act as a proton pump in the thylakoid membrane thus entails maintaining the pumping efficiency at low pH conditions prevailing in the lumen. We propose that vectorial pumping of protons into the thylakoid lumen can be achieved by lowering the pKa of Asp97 and/or by protecting the SB from the luminal pH through rational, site-specific mutagenesis. The electrostatic environment around the SB in PR is presumably maintained by the counter ions, Asp97, Asp227 (analogous to BR Asp212), Arg94 (analogous to BR Arg82) and His75. In BR, the low pKa of Asp85 is attributed to its strong hydrogen bonding interactions with Thr89 and Arg82 [53,54]. Since, interactions that reduce the pKa of Asp97 will promote proton-pumping activity at low external pH, mutation of Met79 to a residue that can hydrogen bond to His75 and Asp212, like Tyr or Thr, will be explored. These mutations are proposed by overlaying the structures of BR and PR, and identifying residues which are in a position to effect the desired behavior. Finally, the ability of a modified PR to work as an efficient H⁺ pump at acidic pHs will also entail shielding the SB from the extracellular environment. To this end, a L219E/T206S mutant will be generated, wherein E219 and S206 will form a Glu-Ser gate regulating vectorial proton transfer as occurs in BR.

To determine if any transgenes alter CET or ATP synthesis activity, we will compare the dark reduction kinetics of the photosystem I primary donor, P700+ in VVT and transgenic plants, with and without dibromothymoquinone (DBMIB), an inhibitor of Cytb6f-mediated CET. Dark P700+ reduction kinetics are expected to be faster in plants with more active CET. In addition, we will assess the amplitude of the After Glow (AG) thermoluminescence band (~40° C.) associated with CET activity [11,14,16,43,63]. Pool sizes of ATP will also be assessed in VVT and transgenic plants by mass spectroscopy.

Referring now to FIG. 11, additional transgenic Camelina lines were produced that expressed the BCA gene (SEQ ID NO:4) in the chloroplast stroma. These lines were produced using the *Agrobacterium*-mediated transformation procedures as described previously. Three lines were evaluated for their ability to accumulate biomass and provide improved photosynthetic rates. Wildtype Camelina and the BCA mutant lines were not significantly different at lower light levels (0-400 umol/m²/s) in their ability to assimilate carbon dioxide. However, as light intensity increased the BCA transformants showed between 10 and 30% higher accumulation of CO₂ at 2000 umoles/m²/s than wildtype. The BCA line 9.2 was the highest while lines BCA 4.1 and BCA 5.7 were both about 10% higher than wildtype. This improved ability to assimilate CO₂ was reflected in two of the lines (BCA-5.7 and BCA-9.2) into increased biomass accumulation, with these lines having about 15% greater biomass accumulation than wildtype. The BCA-4.1 line did not show improved biomass accumulation compared to control.

Referring now to FIG. 12, the ability of the chloroplast enveloped localized bicarbonate transporter bicarbonate transporter (LCIA) protein to transport bicarbonate and improve the capture of inorganic carbon by transgenic Camelina was determined following the method of Farquhar and colleagues (1989). LCIA transgenic Camelina were produced using the *Agrobacterium*-mediated transformation processed described previously. A LCIA expressing mutant line (CAM-LCIA) was compared to wildtype Camelina

(Cam-WT) for the observed discrimination of the stable isotope ^{13}C . This carbon isotope discrimination is expressed as the difference between the ^{13}C in the air and in a plant which has been previously exposed to $^{13}\text{CO}_2$, the carbon isotope discrimination is symbolized by A and expressed in parts per million (ppm) and is described by Farquhar and colleagues (1989). In the LCIA transgenic lines, the observed discrimination by the plant was 20% less than that

observed in the wildtype. This indicates that the insertion of LCIA provides the plant the ability to better accumulate and retain inorganic carbon than the wildtype plant and shows decreased “leakiness” vs wildtype. Reference for ^{13}C discrimination: Carbon isotope discrimination and photosynthesis, G. D. Farquhar, J. R. Ehleringer and K. T. Hubick. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 1989, 40, 503-537.

TABLE D1

Isoenzyme	Kcat (s ⁻¹)	Km (mM)	Kcat/ Km (M ⁻¹ s ⁻¹)	Ki (nM)	Subcellular localization	Tissue I organ localization
hCAI	2×10^5	4.0	5.0×10^7	250	cytosol	E, GI
hCAII	$1.4 \times 10^{6\circ}$	9.3	1.5×10^8	12	cytosol	E, eye, GI, BO, K, L, T, B
hCAIII	1.0×10^4	33.3	3.0×10^5	2×10^5	cytosol	SM, A
hCAIV	1.0×10^6	21.5	5.1×10^7	74	membrane	K, L, P, B, C, H
hCAVA	2.9×10^5	10.0	2.9×10^7	63	mitochondria	Li
hCAVB	9.5×10^5	9.7	9.8×10^7	54	mitochondria	H, SM, P, K, SC, GI
hCAVI	3.4×10^5	6.9	4.9×10^7	11	secreted	G
hCAVII	9.5×10^5	11.4	8.3×10^1	2.5	cytosol	CNS
hCAVIII					cytosol	CNS
hCAIX	3.8×10^5	6.9	5.5×10^7	25	transmembrane	TU, GI
hCAX					cytosol	CNS
hCAXI					cytosol	CNS
hCAXII	4.2×10^5	12.0	3.5×10^7	5.7	transmembrane	R, I, RE, eye, TU
hCAXIII	1.5×10^5	13.8	1.1×10^7	16	cytosol	K, B, L, GI, RE
hCAXIV	3.1×10^5	7.9	3.9×10^7	41	transmembrane	K, B, L
hCAXV	4.7×10^5	14.2	3.3×10^7	72	membrane	K

H = Human;
M = Mouse;
hCAVIII, X, and XI are devoid of catalytic activity.
E = Erythrocytes;
GI = GI tract;
BO = Bone osteoclasts;
K = kidney;
L = Lung;
T = testis;
B = brain;
SM = skeletal muscle;
A = Adipocytes;
P = pancreas;
C = colon;
H = heart;
Li = liver;
SC = spinal cord;
G = salivary and mammary gland;
R = renal;
I = intestinal;
TU = tumors,
RE = Reproductive

TABLE D2

Exemplary Type II Carbonic Anhydrases				
Organism	Sequence	Accession Number	SEQ. ID.	NO.
Human	MSHHWGYGKH NGPEHWHKDF PIAKGERQSP VDIDTHTAKY DPSLKPPLSVS YDQATSLRIL NNGHAFNVER DDSQDKAVLK GGPLDGTYRL IQFHFHWGSL DGQGSEHTVD KKKYAAELHL VHWNTKYGDF GKAVQQPDGL AVLGFIFLKVG SAKPGLQKVV DVLDLSIKTKG KSADFTNFDP RGLLPESLDY WTYPGSLTPP PILLECVTWIV LKEPIVSSE QVLKPRKLNF NGEGEPEELM VDNWRPAQPL KNRQIKASFK	NP-000058.1	SEQ. ID. NO.	19
Macaca fascicularis (crab-eating macaque)	MSHHWGYGKH NGPEHWHKDF PIAKGQRQSP VDIDTHTAKY DPSLKPPLSVS YDQATSLRIL NNGHAFNVER DDSQDKAVIK GGPLDGTYRL IQFHFHWGSL DGQGSEHTVD KKKYAAELHL VHWNTKYGDF GKAVQQPDGL AVLGFIFLKVG SAKPGLQKVV DVLDLSIKTKG KSADFTNFDP RGLLPESLDY WTYPGSLTPP PILLECVTWIV LKEPIVSSE QMSKPRKLNF NGEGEPEELM VDNWRPAQPL KNRQIKASFK	BAE91302.1	SEQ. ID. NO.	24

TABLE D2-continued

Exemplary Type II Carbonic Anhydrases				
Organism	Sequence	Accession Number	SEQ.	ID. NO.
<i>Pan troglodytes</i>	MSHHWGYGKH NGPEHWKDF PIAKGERQSP VDIDTHTAKY DPSLKPLSVS YGQATSLRIL NNGHSFNVF DDSQDKAVLK GGPLDGTYRL IQFHFHWGSL DGQGSEHTVD KKKYAAELHL VHWNNTKYGDF GKAQQPDGL AVLGIFLKVG SAKPGLQKVV DVLDLSIKTKG KSADFTNFPD RGLLPESLDY WTYPGSLTTP PLLECVTWIV LKEPISVSSE QMLKFRKLNF NGEGEPEELM VDNWRPAQPL KNRQIKASFK	NP_001181853	SEQ.	ID. NO. 25
<i>Macaca mulatta</i>	MSHHWGYGKH NGPEHWKDF PIAKGERQSP VDINTHTAKY DPSLKPLSVS YDQATSLRIL NNGHSFNVF DDSQDKAVIK GGPLDGTYRL IQFHFHWGSL DGQGSEHTVD KKKYAAELHL VHWNNTKYGDF GKAQQPDGL AVLGIFLKVG SAKPGLQKVV DVLDLSIKTKG KSADFTNFPD RGLLPESLDY WTYPGSLTTP PLLECVTWIV LKEPISVSSE QMSKFRKLNF NGEGEPEELM VDNWRPAQPL KNRQIKASFK	NP_001182346	SEQ.	ID. NO. 26
<i>Pongo abelii</i>	MSHHWGYGKH NGPEHWKDF PIAKGERQSP VDIDTHTAKY DPSLKPLSVC YDQATSLRIL NNGHSFNVF DDSQDKAVLK GGPLDGTYRL IQFHFHWGSL DGQGSEHTVD KKKYAAELHL VHWNNTKYGDF GKAQQPDGL AVLGIFLKVG SAKPGLQKVV DVLDLSIKTKG KCADFTNFPD RGLLPESLDY WTYPGSLTTP PLLECVTWIV LKEPISVSSE QMLKFRKLNF NGEGEPEELM VDNWRPAQPL KKRRQIKASFK	XP_002819286	SEQ.	ID. NO. 27
<i>Callithrix jacchus</i>	MSHHWGYGKH NGPEHWKDF PIAKGERQSP VDIDTHTAKY DPSLKPLSVS YDQATSWRIL NNGHSFNVF DDSQDKAVLK GGPLDGTYRL IQFHFHWGST DGQGSEHTVD KKKYAAELHL VHWNNTKYGDF GKAQQPDGL AVLGIFLKVG SAKPGLQKVV DVLDLSIKTKG KSADFTNFPD RGLLPESLDY WTYPGSLTTP PLLESVTWIV LKEPISVSSE QILKFRKLNF SGEGEPEELM VDNWRPAQPL KNRQIKASFK	XP_002759086	SEQ.	ID. NO. 28
<i>Lemur catta</i>	MSHHWGYGKH NGPEHWKDF PIAKGERQSP VDINTGAAKH DPSLKPLSVY YEQATSRRIL NNGHSFNVF DDSQDKAVLK GGPLDGTYRL IQFHFHWGSL DGQGSEHTVD KKKYAAELHL VHWNNTKYGDF GKAQQPDGL AVLGIFLKVG SAKPGLQKVV DVLDLSIKTKG KSADFTNFPD RGLLPESLDY WTYLGSLTTP PLLECVTWIV LKEPISVSSE QMMKFRKLSF SGEGEPEELM VDNWRPAQPL KNRQIKASFK	ADD83028	SEQ.	ID. NO. 29
<i>Ailuropoda melanoleuca</i>	MAHHWGYGKH NGPEHWKDF PIAKGQRQSP VDIDTKAAIH DPALKALCPT YEAVSQRVI NNGHSFNVF DDSQDNAVLK GGPLDGTYRL IQFHFHWGSS DGQGSEHTVD KKKYAAELHL VHWNNTKYGDF GKAQQPDGL AVVGVLKVG GAKPGLQKVL DALDSIKTKG KSADFTNFPD RGLLPESLDY WTYPGSLTTP PLLECVTWIV LREPIVSSE QLLKFRRLNF NKEGEPEELM VDNWRPAQPL HNRQINASFK	XP_002916939	SEQ.	ID. NO. 30
<i>Eguus caballus</i>	MSHHWGYGQH NGPKHWKDF PIAKGQRQSP VDIDTKAAVH DAALKPLAVH YEAVSQRRI NNGHSFNVF DDSQDKAVLQ GGPLDGTYRL IQFHFHWGSS DGQGSEHTVD KKKYAAELHL VHWNNTKYGDF GKAQQPDGL AVVGVLKVG GAKPGLQKVL DVLDLSIKTKG KSADFTNFPD RGLLPESLDY WTYPGSLTTP PLLECVTWIV LREPIVSSE QLLKFRRLNF NAEGKPEDPM VDNWRPAQPL NSRQIRASFK	XP_001488540	SEQ.	ID. NO. 31
<i>Canis lupus familiaris</i>	MAHHWGYAKH NGPEHWKDF PIAKGERQSP VDIDTKAAVH DPALKSLCPC YDQAVSQRRI NNGHSFNVF DDSQDKTVLK GGPLDGTYRL IQFHFHWGSS DGQGSEHTVD KKKYAAELHL VHWNNTKYGEF GKAQQPDGL AVLGIFLKIG GANPGLQKIL DALDSIKTKG KSADFTNFPD RGLLPESLDY WTYPGSLTTP PLLECVTWIV LKEPISVSSE QMLKFRKLNF NKEGEPEELM MDNWRPAQPL HSRQINASFK	NP_001138642	SEQ.	ID. NO. 32
<i>Oryctolagus cuniculus</i>	MSHHWGYGKH NGPEHWKDF PIANGERQSP IDIDTNAAKH DPSLKPLRVC YEHPISRRRI NNGHSFNVF DDSHDKTVLK EGPLEGTYRL IQFHFHWGSS DGQGSEHTVN KKKYAAELHL VHWNNTKYGDF GKAQKHPDGL AVLGIFLKIG SATPGLQKVV DTLSSIKTKG KSVDFTNDPP RGLLPESLDY WTYPGSLTTP PLLECVTWIV LKEPITVSSE QMLKFRNLNF NKEAEPEEPM VDNWRPTQPL KGQVKASFK	NP_001182637	SEQ.	ID. NO. 33
<i>Ailuropoda melanoleuca</i>	GPEHWYWKDFP IAKGQRQSPV DIDTKAAHD PALKALCPTY EFB24165 SEQ. ID. NO. 34 EQAVSQRVIN NGHSFNVFED DSQDNAVLKG GPLTGTYRLI QGFHFHWGSSD GQGSEHTVDK KKYAAELHLV HWNTKYGDFG			

TABLE D2-continued

Exemplary Type II Carbonic Anhydrases			
Organism	Sequence	Accession Number	SEQ. ID. NO
	KAVQQPDGLA VLGIFLKIGD ARPGLOKVL ALDSIKTKKG SADFTNFDPR GLLPESLDW TYPGSLLTPP LLECVTIVL KEPIVSSEQ MLKFRRLNFN KEGEPEELMV DNWRPAQPLH NRQINASFK		
<i>Sus scrofa</i>	MSSHHWGYDKH NGPEHWHKDF PIAGKDRQSP VDINTSTAVH DPALKPLSLC YEQATSRIV NNGHSFNVF DDSQDKGVLE GGPLAGTYRL IQFHFHWGSS DGQGSEHTVD KKKYAAELHL VHWNTKYKDF GEAAQQPDGL AVLGVFLKIG NAQPGLOKIV DVLDISIKTKG KSVEFTGFDP RDLLPGSLDY WTYPGSLTTP PLLESVTWIV LREPISVSSG QMMKFRTLNF NKEGEPEHPM VDNWRPTQPL KNRQIRASFQ	XP_001927840.1	SEQ. ID. NO. 35
<i>Callithrix jacchus</i>	MSSHHWGYGKH NGPEHWHKDF PIAGKERQSP VDIDTHTAKY DPSLKLPLSVS YDQATSWRIL NNGHSFNVF DDSQDKAVLK GGPLDGTYRL IQLHLVHWNT KYGDFGKAAQ QPDGLAVLGI FLKVGSAKPG LQKVVDVLDS IKTGKKSADF TNFDPRGLLP ESLDYWTYPC SLTPPLLES VTWIVLKEPI SVSSEQILKF RKLNFSGEGE PEELMVDNWR PAQPLKNRQI KASFK	XP_002759087	SEQ. ID. NO. 36
<i>Mus musculus</i>	MSSHHWGYSKH NGPENWHKDF PIANGDRQSP VDIDTATAQH DPALQPLLIS YDKAASKSIV NNGHSFNVF DDSQDNAVLK GGPLSDSYRL IQFHFHWGSS DGQGSEHTVN KKKYAAELHL VHWNTKYGDF GKAVQQPDGL AVLGVFLKIG PASQGLQKVL EALHSIKTKG KRAAFANFDP CSLLPGMLDY WTYPGSLTTP PLLECVTIVL LKEPITVSSE QMSHFRTLNF NEEGDAEAM VDNWRPAQPL KNRRIKASFK	NP_033931	SEQ. ID. NO. 37
<i>Bos taurus</i>	MSSHHWGYGKH NGPEHWHKDF PIANGERQSP VDIDTKAVVQ DPALKPLALV YGEATSRMV NNGHSFNVF DDSQDKAVLK DGPLTGTYRL VQFHFHWGSS DDQGSEHTVD RKKYAAELHL VHWNTKYGDF GTAAQQPDGL AVVGFLKVG DANPALQKVL DALDISIKTKG KSTDFPNDDP GSLLPNVLDY WTYPGSLTTP PLLESVTWIV LKEPITVSQ QMLKFRTLNF NAEGEPELLM LANWRPAQPL KNRQVRGFPK	NP_848667	SEQ. ID. NO. 38
<i>Oryctolagus cuniculus</i>	GKHNHGPEHWH KDPFIANGER QSPIDIDTNA AKHDPSLKPL RVCYEHPIR RIINNGHSFN VEFDDSHDKT VLKEGPLEGT YRLIQFHFHW GSDDGQGSEH TVNKKKYYAE LHLVHWNTKY GDPGKAVKHP DGLAVLGIFL KIGSATPGLQ KVVDTLSSIK TKGKSVDFTD FDPRGLLPES LDYWTPGSL TPPPLLECVT WIVLKEPITV SSEQMLKPRN LNFNKEAEPE EP	AAA80531	SEQ. ID. NO. 39
<i>Rattus norvegicus</i>	MSSHHWGYSKS NGPENWHKEF PIANGDRQSP VDIDTGTAQH DPSLQPLLIC YDKVASKSIV NNGHSFNVF DDSQDFAVLK EGPLSGSYRL IQFHFHWGSS DGQGSEHTVN KKKYAAELHL VHWNTKYGDF GKAVQHPDGL AVLGVFLKIG PASQGLQKIT EALHSIKTKG KRAAFANFDP CSLLPGMLDY WTYPGSLTTP PLLECVTIVL LKEPITVSSE QMSHFRTLNF NSEGRAEELM VDNWRPAQPL KNRRIKASFK	NP062164	SEQ. ID. NO. 40

TABLE D3

Exemplary Type VII Carbonic Anhydrases			
Organism	Sequence	Accession Number	SEQ. ID. NO
Human	MSLSITNNGH SVQVDFNDSD DRTVVTGGPL EGPYRLKQFH FHWGKKHDVG SEHTVDGKSF PSELHLVHWN AKKYSTFGEA ASAPDGLAVV GVPLETGDDEH PSMNRLTDAL YMVRFKGKTA QFSCFNPKCL LPASRHWTY PGSLTTPLS ESVTIVLRE PICISEROMG KFRSLLFTSE DDERIHMVNN FRPPQPLKGR VVKASFRA		SEQ. ID. NO. 41
<i>Pongo abelii</i>	MTGHHGHWGYG QDDGPSHWK LYPIAQGDRQ SPINIISQAA VYSPLSLQPLIE LSYEACMLS ITNNNGHSVQV DFNDSDDRTV VTGGPLEGY RLKQFHFHWG KKHDVGSEHT VDGKSFPSL HLVHWNAKKY STFGEAASAP DGLAVVGVFL ETGDEHPSMN RLTDALYMR FKGTKAQFSCFNPKSLLPAS RHYWTPGSL TPPPLSESVT WIVLREPICI SERQMGKFRS LLFTSEDDER IHMVNNFRPP QPLKGRVVKA SFRA	XP_002826555	SEQ. ID. NO. 42

TABLE D3-continued

Exemplary Type VII Carbonic Anhydrases				
Organism	Sequence	Accession Number	SEQ.	ID. NO.
<i>Pan troglodytes</i>	MEFGLSPELS PSRCFKRLLR GSERGRSRSP NERTEPTGQV HGC GDGSGMT GHGWGYQD DGP SHWHKLY PIAQGDRQSP INI ISSLQAVY SP SLQPLELS YEACMSL SIT NNGHSVQVDF NDSDRTVVT CGPLEGPYRL KQFHFHWGKK HDVGSEHTVD GKS FSELHL VHWNAKKY STFGEAASAPDG LAVGVVFLET GDEHPSMNRL TDALYMRFK GTKAQFSCFN PKCLLPASRH YTWTYPSLTT PPLSESVTWI VLREPICISE RQMRKFRSLL FTSEDDERIH MVNNFRPPQP QPLKGRVVKA SFRA	XP_001143159.1	SEQ.	ID. NO. 43
<i>Callithrix jacchus</i>	MTGHHGWG YG QDDGPSHWK LYPIAQGDRQ SPINIISSQA VYSPSLQPL ELSYEACMSL ITNNGHSVQV DFNDSDRTV VTGGPLEG PY RLQFHFHWG KKHDVGSEHT VDGKSFPS HLVHWNAKKY STFGEAASAP DGLAVGVFVL ETGDEHPSMN RLTDALYMR FKGTKAQFSC FNPKCLLPAS WHYWTYPSL TTPPLSES VT WIVLREPICI SERQMGKFRS LLFTSEDDER VHMVNNFRPP QPLKGRVVKA SFRA	XP_002761099	SEQ.	ID. NO. 44
<i>Ailuropoda melanoleuca</i>	GPSQWHKLYP IAQGDRQSPI NIVSSQAVYS PSLKP LEPLY EACTLSNS NGHSVQVDFN DSDDR TVTG GPLDGYRLK QFH FHWG KKH SVGEHTV DGKSFPS SELHLV HWNAKKY STF GEAASAPDGL AVVGVL ETGDEHPSMN RLTDALYMR FKGTKAQFSC FNPKCLLPAS WHYWTYPSL TPLSES VT WIVLREPICI SERQMGKFRS LREPISI SERQMEKFRS LLFTSEDDER IHM VNNFRPPQPL KGRVVKA SFRA	EFB15849	SEQ.	ID. NO. 45
<i>Canis familiaris</i>	MTGHHCWG YG QNDEIQASLS PSLSTPAGPS QWHKLYPIAQ GDRQSPINIV SSQAVYSPSL KPLELSYEAC ISLSITNNGH SVQVDFNDS DRATV TG GPLDGYRLK FHWGKKH SVG SEHTV DGKSFPS SELHLV HWNAKKY STFGEA ASAPDGL AVGV GIFLETGDEH PSMNRLTDAL YMVRFKT KAQFSCFPKCL LPASRHYWTY PGSLTT PPLSES VT WIVLREPICI SERQMGKFRS KFRSLLFTSE EDERIH MVNNFRPP QPLKGRVVKA SFRA	XP_546892	SEQ.	ID. NO. 46
<i>Bos taurus</i>	MTGHHGWG YG QNDGPSHWK LYPIAQGDRQ SPINIVSSQA VYSPSLKPL ESYECS TSL SIANNGHSVQV DFNDSDRTV VS GGLDGPY RLQFHFHWG KKHDVGSEHT VDGKSFPS HLVHWNAKKY STFGEAASAP DGLAVGVFVL ETGDEHPSMN RLTDALYMR FKGTKAQFSC FNPKCLLPAS RHYWTYPSL TTPPLSES VT WIVLREPICI SERQMEKFRS LLFTSEDDER IHMVNNFRPP QPLKGRVVKA SFRA	XP_002694851	SEQ.	ID. NO. 47
<i>Rattus norvegicus</i>	MTVLWWPMIL EELMSKLRTG GPSNWHKLYP IAQGDRQSPI NI ISSLQAVYS PSLQPLELFY EACMSL SITN NGHSVQVDFN DSDDR TVTGAG GPLEG PY RLQFHFHWG KKHDVGSEHT VDGKSFPS KSFP SELHLV HWNAKKY STFGEAASAP DGLAVGVFVL ETGDEHPSMN DEHP S MNRLTDALYMR FKGTKAQFSC FNPKCLLPAS WHYWTYPSL TTPPLSES VT WIVLREPICI SERQMEKFRS LLFTSEDDER IHMVNNFRPP QPLKGRVVKA SFRA	EDL87229	SEQ.	ID. NO. 48
<i>Oryctolagus cuniculus</i>	MTGHHGWG YG QDDGRPSHWK HKLYPIAQGD RQSPINIVSS QAVYSPGLQ PLELSYEACTS LSIANNGHSV QVDFNDS DDR TVTGGLP EYLKQFHFHWG KKHDVGSEHT VDGKSFPS ELHLVHWNAK KYSTFGEAASAP DGLAVGVFVL FLETGNEHPS MNRLTDALYMR VRFKGTKAQFSC FNPKCLLPAS WHYWTYPSL SLTTPLSES VT WIVLREPICI SERQMEKFRS LLFTSEDDER ERHVHMVNNFRPP QPLKGRVVKA SFRA	XP_002711604	SEQ.	ID. NO. 49
<i>Mus musculus</i>	GQDDGPSNW H KLYPIAQGD QSPINIISSQ AVYSPSLQPL ELFYEACMSL SITNNGHSVQV DFNDSDRTV VVSGGPLEGP YRLQFHFHWG KKHDVGSEHT VDGKSFPS LHLVHWNAKK YSTFGEAAA PDGLAVGVFVL LETGDEHPSMN RLTDALYMR RFKDTKAQFS CFNP KCLLPAS SRHYWTYPSL TTPPLSES TWIVLREPICI SERQMEKFRS LLFTSEDDER RIHMVDNF RP PQPLKGRVVKA SFRA	AAG16230.1	SEQ.	ID. NO. 50
<i>Monodelphis domesticus</i>	MTGHHGWG YG QEDGPSEWHK LYPIAQGDRQ SPIDIVSSQA VYDP TLKPLV LAYECS MSL SIANNGHSV MV EFDDV DRTV VNGGPLDGPY RLQFHFHWG KKHDVGSEHT VDGKSFPS HLVHWNAKKY KTFAEEAAAP DGLAVGVFVL ETGDEHPSMN RLTDALYMR FKGTKAQFSC FNPKCLLPAS LSYWTYPSL TTPPLSES VT WIVLKEPITI SEKQMEKFRS LLFTSEDDER VRMVNNFRPP QPLKGRVVKA SFRA	XP_001364411.1	SEQ.	ID. NO. 51
<i>Gallus gallus</i>	MTGHHSW GY QDDGPAEWHK SYPIAQGNRQ SPIDIISAKA VYDP KLMPLV ISYESCTSLN ISNNGHSV MV EFEDIDD KTV	XP_414152.1	SEQ.	ID. NO. 52

TABLE D3-continued

Exemplary Type VII Carbonic Anhydrases			
Organism	Sequence	Accession Number	SEQ. ID. NO
	ISGGPFESPP RLKQFHFHWG AKHSEGSEHT IDGKPPCCE HLVHWNAKKY ATFGEAAAAP DGLAVGVFL EIGKEHANMN RLTDALYMVK FKGTKAQFRS FNPKCLLPLS LDYWTYLGSL TTPPLNESVI WVLKEPISI SEKQLEKFRM LLFTSEEDQK VQMNNFRPP QPLKGRTVRA SFKA		
Taeniopygia guttata	MTGQHSWGYG QADGPSEWHK AYPIAQGNRQ SPIDIDSARA VYDPLSLQPLL ISYESCSSLIS ISNTGHSMV EFEDTDDRTA ISGGPFQNPRL KQFHFHWG TTHSQGSEHT IDGKPPCCE HLVHWNARKY TTFGEEAAAAP DGLAVGVFL EIGKEHASMN RLTDALYMVK FKGTKAQFRG FNPKCLLPLS LDYWTYLGSL TTPPLNESVT WIVLKEPIRI SVKQLEKFRM LLFTGEEDQR IQMANNFRPP QPLKGRTVRA SFKA	XP_002190292.1	SEQ. ID. NO. 53

TABLE D4

Exemplary Type XIII Carbonic Anhydrases			
Organism	Sequence	Accession Number	SEQ. ID. NO
Human	MSRLSWGYRE HNGPIHWKEF FPIADGDQQS PIEIKTKEVK NP_940986.1 YDSSLRPLSI KYDPSSAKII SNSGHSFNVDF DDTEENKSVL RGGPLTGSYR LRQFHLHWGS ADDHGSEHIV DGVSYAAELH VVHWNSDKYP SFVEAAHEPD GLAVLGVFLQ IGEPNSQLQK ITDTLDLSIKE KGKQTRFTNF DLLSLLPPSW DYWTYPGSLT VPPLLESVTW IVLKQPINIS SQQLAKFRSL LCTAEGEAAA FLVSNHRPPQ PLKGRKVRAFH		SEQ. ID. NO. 54
Pan troglodytes	MSRLSWGYRE HNGPIHWKEF FPIADGDQQS PIEIKTKEVK XP_001169377.1 YDSSLRPLSI KYDPSSAKII SNSGHSFNVDF DDTEENKSVL RGGPLTGSYR LRQFHLHWGS ADDHGSEHIV DGVSYAAELH VVHWNSDKYP SFVEAAHEPD GLAVLGVFLQ IGEPNSQLQK ITDTLDLSIKE KGKQTRFTNF DLLSLLPPSW DYWTYPGSLT VPPLLESVTW IVLKQPINIS SQQLAKFRSL LCTAEGEAAA FLVSNHRPPQ PLKGRKVRAFH		SEQ. ID. NO. 55
Macaca mulatta	MSRLSWGYRE HNGPIHWKEF FPIADGDQQS PIEIKTKEVK XP_001095487.1 YDSSLRPLSI KYDPSSAKII SNSGHSFNVDF DDTEENKSVL RGGPLTGSYR LRQFHLHWGS ADDHGSEHIV DGVSYAAELH VVHWNSDKYP SFVEAAHEPD GLAVLGVFLQ IGEPNSQLQK ITDTLDLSIKE KGKQTRFTNF DLLSLLPPSW DYWTYPGSLT VPPLLESVTW IVLKQPINIS SQQLAKFRSL LCTAEGEAAA FLLNSNHRPPQ PLKGRKVRAFH		SEQ. ID. NO. 56
Oryctolagus cuniculus	MSRISWGYGE HNGPIHWNQF FPIADGDQQS PIEIKTKEVK XP_002710714.1 YDSSLRPLSI KYDPSSAKII SNSGHSFNVDF DDTEENKSVL RGGPLTGSYR LRQFHLHWGS ADDHGSEHIV DGVRYAAELH VVHWNSDKYP SFVEAAHEPD GLAVLGVFLQ IGEYNSQLQK ITDILDLSIKE KGKQTRFTNF DLLSLLPPSW DYWTYPGSLT VPPLLESVTW IVLKQPINIS SQQLAKFRSL LCSAEGESAA FLLSNHRPPQ PLKGRKVRAFH		SEQ. ID. NO. 57
Ailuropoda melanoleuca	MSRLSWGYGE HNGPIHWKEF FPIADGDQQS PIEIKTKEVK XP_002916937.1 YDSSLRPLSI KYDANSAKII SNSGHSFNVDF DDTEENKSVL RGGPLTGSYR LRQFHLHWGS ADDHGSEHIV DGVRYAAELH VVHWNSDKYP SFVEAAHEPD GLAVLGVFLQ IGEHNSQLQK ITDILDLSIKE KGKQTRFTNF DLLSLLPPSW DYWTYPGSLT VPPLLESVTW IVLKQPINIS SQQLATFRTL LCTAEGEAAA FLLSNHRPPQ PLKGRKVRAFH		SEQ. ID. NO. 58
Sus scrofa	MSRFSTWGYGE HNGPVHNEF FPIADGDQQS PIEIKTKEVK XP_001924497.1 YDSSLRPLSI KYDPSSAKII SNSGHSFNVDF DDTEENKSVL RGGPLTGSYR LRQFHLHWGS ADDHGSEHIV DGVKYAAELH VVHWNSDKYP SFVEAAHEPD GLAVLGVFLQ IGEHNSQQLQK ITDILDLSIKE KGKQTRFTNF DLLSLLPPSW DYWTYPGSLT VPPLLESVTW IILKQPINIS SQQLATFRTL LCTKEGEAAA FLLSNHRPLQ PLKGRKVRAFH		SEQ. ID. NO. 59
Callithrix jacchus	MSRLSWGYGE HNGPIHWNEF FPIADGDQQS PIEIKAKEVK XP_002759085.1 YDSSLRPLSI KYDPSSAKII SNSGHSFNVDF DDTEENKSVL RGGPLTGSYR LRQFHLHWGS ADDHGSEHIV DGVRYAAELH		SEQ. ID. NO. 60

TABLE D4-continued

Exemplary Type XIII Carbonic Anhydrases			
Organism	Sequence	Accession Number	SEQ. ID. NO
	VVHWNSEKYP SFVEAAHEPD GLAVLGFLQ IGEPNSQLQK IIDILDLSIKE KGKQIRFTNF DPLSLFPPSW DYWTYSGSLT VPPLLESVTW ILLKQPINIS SQQLAKFRSL LCTAEGEAAA FLLSNYRPPQ PLKGRKVRA S FR		
Rattus norvegicus	MARLSWGYDE HNGPIHWNEL FPIADGDQQS PIEIKTKEVK NP_001128465.1 SEQ. ID. NO. 61 YDSSLRPLSI KYDPASAKII SNSGHSFNVD FDDTEDKSVL RGGPLTGYSR LRQFHLHWGS ADDHGSEHVV DGVRYAAELH VVHWNSDKYP SFVEAAHESD GLAVLGFLQ IGEHNPQLQK ITDILDLSIKE KGKQTRFTNF DPLCLLPSSW DYWTYPGSLT VPPLLESVTW IVLKQPISIS SQQLARFRSL LCTAEGESAA FLLSNHRPPQ PLKGRRVRAS FY		
Mus musculus	MARLSWGYGE HNGPIHWNEL FPIADGDQQS PIEIKTKEVK NP_078771.1 SEQ. ID. NO. 62 YDSSLRPLSI KYDPASAKII SNSGHSFNVD FDDTEDKSVL RGGPLTGNYR LRQFHLHWGS ADDHGSEHVV DGVRYAAELH VVHWNSDKYP SFVEAAHESD GLAVLGFLQ IGEHNPQLQK ITDILDLSIKE KGKQTRFTNF DPLCLLPSSW DYWTYPGSLT VPPLLESVTW IVLKQPISIS SQQLARFRSL LCTAEGESAA FLLSNHRPPQ PLKGRRVRAS FY		
Canis familiaris	MPPRRHGPNT FLSAGTKGQQ NFWTKNQKSG PIHWNKFFP1 XP_544159 SEQ. ID. NO. 63 ADGDQSPIE IKTKEVKYDS SLRPLSIKYD ANSAKIISNS GHSFSVDFDD TEDKSVLRRG PLTGSYRLRQ FHLHWGSADD HGSEHVVVG RYAAELHVHV WNSDKYPSFV EAAHEPDGLA VLGVFLQIGE HNSQLQKITD ILDSIKEKGK QTRFTNFDPL SLLPPSWDYW TYPGSLTVPP LLESVTWIVL KQPINISSQQ LATFRILLCT AEGEAAAFLL SNHRPPQPLK GRKVRA SFH		
Eguus caballus	MSGPVHNEF FPIADGDQQS PIEIKTKEVK YDSSLRPLTI XP_001489984.2 SEQ. ID. NO. 64 KYDPSSAKII SNSGHSFSVG FDDTENKSVL RGGPLTGYSR LRQFHLHWGS ADDHGSEHVV DGVRYAAELH IVHWNSDKYP SFVEAAHEPD GLAVLGFLQ VGEHNPQLQK ITDTLDLSIKE KGKQTLFTNF DPLSLLPPSW DYWTYPGSLT VPPLLESVTW IILKQPINIS SQQLVKFRRTL LCTAEGETAA FLLSNHRPPQ PLKGRKVRA S FR		
Bos taurus	MSGFWGYGE RDGPVHNEF FPIADGDQQS PIEIKTKEVR XP_002692875.1 SEQ. ID. NO. 65 YDSSLRPLGI KYDASSAKII SNSGHSFNVD FDDTDDKSVL RGGPLTGYSR LRQFHLHWGS TDDHGSEHVV DGVRYAAELH VVHWNSDKYP SFVEAAHEPD GLAVLGIFLQ IGEHNPQLQK ITDILDLSIKE KGKQTRFTNF DPVCLLPPCR DYWTYPGSLT VPPLLESVTW IILKQPINIS SQQLAAFRTL LCSREGETAA FLLSNHRPPQ PLKGRKVRA S FR		
Monodelphis domestica	MSRLSWGYCE HNGPVHSEL FPIADGDYQS PIEINTKEVK XP_001366749.1 SEQ. ID. NO. 66 YDSSLRPLSI KYDPASAKII SNSGHSFSVD FDDSEDKSVL RGGPLIGTYR LRQFHLHWGS TDDQGSEHTV DGMKYAAELH VVHWNSDKYP SFVEAAHEPD GLAVLGIFLQ TGEHNLQMOK ITDILDLSIKE KGKQIRFTNF DPATLLPQSW DYWTYPGSLT VPPLLESVTW IVLKQPITIS SQQLAKFRSL LYTGEGEAAA FLLSNYRPPQ PLKGRKVRA S FR		

TABLE D4-continued

Exemplary Type XIII Carbonic Anhydrases			
Organism	Sequence	Accession Number	SEQ. ID. NO.
<i>Ornithorhynchus anatinus</i>	MKKGVGSFYE LAVNRWSVNV RVQIMIVESI TEPLLCGSRA XP_001507177.1 SEQ. ID. NO. 67 LALTLSPTQA LAVAPALALA VVQALALTVV QALALAVSPA LALSVAPALA LAVVQALALA VVQALALAVA QALALAVAAQ LALAVAAQALA LALPQALALT LPQALALTLS PTIALSVAPA LALAVAPALA LADSPALALA LARPHPSSGS SPALDCELVL FGDCHTVLLK WMRMGNYSSV SPLERNSSC PLGPIHWNEL FPIADGDRQS PIEIKTKEVK YDSSLRPLSI KYDPTSAKII SNSGHFSVDF DDDTEDKSVL RGGPLSGTYR LRQPHFHWS ADDHGSEHTV DGMESYSAELH VVHWNSDKYS SFVEAAHEPD GLAVLGIFLK RGEHNQLQK ITDILDAIKE KGKQMRFTNF DPLSLPLTR DYWTYPGSLT VPPLLESVIW IIFKQPISIS SQLAKFRNL LYTAEGEAAD FMLSNNHRPPQ PLKGRKVRS FRS		

TABLE D5

Exemplary CA II DNA expression constructs for chloroplast expression			
ATGTCCCAC ACTGGGGGTA CGGCCAACAC AACGGACCTG AGCACTGGCA TAAGGACTTC	SEQ. ID. NO. 94		
CCCATGGCCA AGGGAGAGCG CCAGTCCCCT GTTGACATCG ACACATCAC AGCCAAGTAT	(human cDNA sequence)		
GACCCTTCCC TGAAGCCCTG GTCTGTTCC TAGTATCAAG CAACTCCCT GAGGATCTC			
AACAATGGTC ATGCTTCAA CGTGAGGTTT GATGACTCTC AGGACAAAGC AGTGCCTAAG			
GGAGGACCCC TGGATGGCAC TTACAGATTG ATTCACTTC ACTTTCACTG GGGTTCACTT			
GATGGACAAG GTTCAGAGCA TACTGTGGAT AAAAGAAAT ATGCTGCAGA ACTTCACCTG			
GTCTACTGGA ACACCAAATA TGGAAGTTTT GGAAAGGCTG TGCAGCAACC TGATGGACTG			
GCCGTTCTAG TATTTTTTG GAAGGTTGGC AGCGCTAAC CGGGCCTTC GAAAGTTGTT			
GATGTGCTGG ATTCCATTAA AACAAAGGGC AAAGGTGCTG ACTTCACCAA CTTCGATCCT			
CGTGGCCCTC TTCTGAATC CTTGGATTAC TGGACCTACCC CAGGCTCACT GACCACCCCT			
CCTCTCTGG ATATGTGAC CTGATTGTTG CTCAGGAAAC CCATCAGCGT CAGCAGCGAG			
CAGGTGTTGAA AATTCCGAA ACTTAACTTC AATGGGGAGG GTGAACCCGA AGAACTGATG			
GTGGACAACT GGCGCCAGC TCAGCCACTG AAGAACAGGC AAATCAAAGC TTCTTCAAA			
TAA			
gaattcATGTctCATCATGGGtATGGtaAACACAAAtGGtCCTGAaCACTGGCATAAgACTT	SEQ. ID. NO. 108		
tCCaATTGCaAaGGtGAACTtAaCCTGTTGatAtTGACACTCATAcAGCtAAaTATGACC	(Optimized for		
CTTCttTaAAaCCatTaTCTGTTCaTATGATCAAGCAACTTCTtTacGtAttTaAAACATGGT	chloroplast		
CATGCTTTAatGtaGaaTTTGATGACTCTCAAGtAAAGCAGTatTaAAaGtGgtCCatTaGA	Expression)		
TGGTACTTACGtTTaATTCACTTCACTGGGTTCACTGATGGTCAAGGTTCAAGAAc			
ATACTGtaGATAAAAaAAATATGTCAGAAatTaCACTtaGTTCACTGGAAACACaAAATATGGt			
GATTTGGtAAAGCTGtaCAaCAACCTGATGGGtTaGctGTTTAGGTATTTTTaAaGTGTT			
tAgTGTAAACCAAGGtCTTCAAAAGTTGATGATGatTaGATTCAATTAAACAAAGtAAaA			
GTGCTGACTTtACTAAttTTCGATCCTGTTGatTaCTTCTGGAATCTTAgATTACTGGACATAt			
CCAGGtTCAtAACAAcCCTCTTtTaGATGTGtaAcGATGTTGatTaAAaGAACCAAT			
tAGtGtAAGtGtGAAcAAcAtTAaATTCCGTAACACTTAATTTCAATGgtGAAGGTGACCGa			
AGAAAtTAATGGTtGtAACTGGCgtCCAGCTCAcCAtTAAAAtcGtCAAATtAAAGCTtCa			
TTCAAATAAAGcatqc			

TABLE D6

TABLE D6-continued

Codons in Human CA II optimized for expression in chloroplast of <i>Chlamydomonas reinhardtii</i>				Codons in Human CA II optimized for expression in chloroplast of <i>Chlamydomonas reinhardtii</i>			
Amino acid	Total number	Number of codons that were optimized	No. of amino acids of each codon	55	Amino acid	Total number	Number of codons that were optimized
Ser(S)	18	12	TCT TCA AGT (7:5)	1:1:1	A(D)	19	3
Phe(F)	12	3	TIT TTC (8:4)	2:1	Ile(I)	9	4
Leu(L)	26	19	TIA CTT (21:5)	5:1	Met(M)	2	0
Val(V)	17	10	GTT GTA (8:9)	1:1	60 Gln(Q)	11	7
Pro(P)	17	6	CCT CCA (8:9)	3:4	Glu(E)	13	6
Thr(T)	12	5	ACT ACA (5:7)	2:3	Lys(K)	24	11
Ala(A)	13	3	GCT GCA (9:4)	2:1	Cys(C)	1	0
Tyr(Y)	8	2	TAT TAC (6:2)	2:1	Tfe(W)	7	0
His(H)	12	1	CAT CAC (6:6)	1:1	Gly(G)	22	17
Asn(N)	10	4	AAT AAC (7:3)	2.5 1	65 Arg(R)	7	5

TABLE D7

Exemplary algal bicarbonate transporter types				
Transport Type	Mechanism	Substrate affinity	Flux rate	Photosynthetic affinity ko.6
BicA	Na+ dependent	Low- medium	High	90-170 μ M HC ₃₋
SbtA	Na+ dependent	High	Low	<5 μ M HC ₃₋
	HC ₃₋ uptake			

TABLE D7-continued

Exemplary algal bicarbonate transporter types					
5	Transport Type	Mechanism	Substrate affinity	Flux rate	Photosynthetic affinity ko.6
10	BicA	Na+ dependent	Low- medium	High	90-170 μ M HC ₃₋
	SbtA	Na+ dependent	High	Low	<5 μ M HC ₃₋

TABLE D8

Exemplary plasma membrane localized Bicarbonate transporters				
Organism	Sequence	Accession Number	SEQ. ID.	NO.
<i>Chlamydomonas reinhardtii</i>	MLPGLGVILL VLPMQYFFGY KIVQIKLQNA KHVALRSAIM QEVLPAlKLV KYYAWEQFFE NQISKVRRREE IRLNFWNCVM KVINVACVFC VPPMTAFVIF TTYEQRARL VSSVAFTTLS LFNILRFPPLV VLPKALRAVS EANASLQLRLE AYLLEEVPSG TAAVKTPKNA PPGAVIENGV FHHPSNPNNW LHVPKFVEKP GQVVAVVGRI AAGKSSLVQA ILGNMVKEHG SFNVGGRISY VPQNPNWLQNL SLRDNVLFGE QFDENKYTDV IESCALTLLD QILSNGDQSK AGIRGVNFSG GQRQRVNLLAR CAYADADLVL LDNALSAVDH HTAAHIFDKC IKGLFSDKAV VLVTHQIEPM PRCDNVAIMD EGRCLYFGKW NEEAQHLLGK LLPITHLLHA AGSQEAPPAP KKKAEDKAGP QKSQSLQLTL APTSIGKPFK KPKDVQKLTA YQAAIYITWY GNLFLVGVCE FFFLAAQCSR QISDFWVRWW VNDEYKKFPV KGEQDSAATT FYCLIYLLLV GLFYIFMIFR GATFLWWVLLK SSETIRRKAL HNVLNAPMGF FLVTPVGDLL LNFTKDQDIM DENLPDAVHF MGIVGLILLA TTITVSVTIN FFAAFTGALI IMTLIMLSIY LPAATALKKA RAVSGGMLVG LVAEVLEGGLG VVQAFNQKEY FIEEAARRTN ITNSAVPNAE ALNLWLAFWC DFIGACLVGV VSAPAVGMAK DLGGATVGLA FSNIIQMLVF YTWWVRFISE SISLNFNSVEG MAYLADYVPH DGVFYDQRQK DGVAQIVLP DGNIVPAASK VQVVVDDAAL ARWPATGNIR FEDVWMQYRL DAPWALKGVT FKINDGEKVG AVGRTGSGKS TTLLALYRMF ELGKGRLIVD GVDIATLSK RLRTGLSIIP QEPVMFTGTV RSNLDPFGEF KDDAILWEVL KKVGLEDQAO HAGGLDGQVD GTGKKAWSLG QMQLVCLARA ALRAVPILCL DEATAAMDPH TEAIVQQTIK KVFDDDRIT IAHRRLDTIE SLMYEESP SK LLANRDSMFS KLVDKTGPAA AAALRKMAED FWSTRSAQGR NQ	EDP07736.1	SEQ. ID.	77
<i>Volvox carteri f. nagariensis</i>	MGTISHMPARG NDPTAGFFNK EAEGWMFKHV SEARKNGDID LDKMGMPPEN HAHEAYDMFA SNWAEMKLK DSGAKPSLVR ALRKSFGFLVY LLGGVFKCFW STEVITGAFY FVRSLLAHVN GIKDGRILYSK TVSGWCLMAG FTLDLAWLLGL SLQRMGYICM SVGIRARAAL VQAVTHKAFR LSSVRADQSA AIVNFVSSDI QKIYDGALEF HYLTWATPEA AAIALLGYL TNDSLPGLG VILLVLPLOY FFGYKIIQIK LQNAKHVALR SSILQEVLPA IKLVVKPLQY QFFEDIEISKI RREEMRLSFV NAMMKVINA CVFCVPPMTA FVIPTTYEFQ KARLVSGVAF TTLSLFNILR FPLVLPKAL RAVSEAHASL QRLEYSLLED VPQGTASGGK SSKSSAPGVH IDNAVYHHPN NPNWHLHVPR FDVRPGQVVA VVGRIGAGKS SLVQAILGNM VKEHGSQQVG GRISYVPQNP WIQNLSLIRDN VTFGEWDEA KYEAVIDACA LTMDLQILFQ GDQSKAGIRG VNFSGGQRQR VNLRACAYAD ADLVLNDNAL SAVDHHTAHH IFDKC1KGLF SDKAVVLLITH QIEFMPRCDA VAIMDEGRCL YFGKWNEESQ HLLGKLLPIT HLLHAAGSQE APPAAPKKD DKATPQKSQS LQLTLAPTSI GKPTQKDTKA APKLTAFKAA LIYTYYGNIL LVFVCFITPL AAQTCRQMSD FWVRRWWVNDE YKHFPKRTGV REESATKFYA LIYLLLVLGF YFTMVARGST FLWWVLRSS NIKKCALNNV LNAPMGFFLV TPVGDLNNF TKDQDIMDEN LPDAIHFMGI YGLLILLATTI TVSVTINFEG AFTGFLIIMT LIMLAIYLP AITALKKARAV SGQLVGLVA EVLEGlnvvq AFSKQEYFIE EAARRTDVTN AAVFNAESLN LWLAFWCDLI GASLUVGVSA FAVGLKDQLG AATVGLAFSN IIQMLVFYTW VVRFIAESIS LFNSVEAMAW	XP_002950646.1	SEQ. ID.	69

TABLE D8-continued

Exemplary plasma membrane localized Bicarbonate transporters				
Organism	Sequence	Accession Number	SEQ.	ID. NO.
	LADYVPKDGFI FYDQKQLDGV AKSITLPDGQ IVPATSKVQVVDDAALARW PATGNIRFED VWMQYRLDAA WALKGVTFKINDGEKVGAVG RTGSGKSTTL LALYRMFELG KGRILIDGVDTATLSLKRRL TGLSIIQPQE VMFTGTVRSN LDPPGFEFKDDSVLWEVLQKV GLEAQAOHQAG GLDGRVDTG GKAWSLQCMOLVCLARAALR AVPILCLDEA TAAMDPHTEQ VVQETIKKVFDDRRTTIAH RLDTIIESDK VLVMEEAGELK EFAPPAAQLLANRETMFSKLV DKTGPAAAAA LRKMADEHFS KSQARAAAQRH			
<i>Chlorella variabilis</i>	MVPLLAQRGR IRSQAPRTWH PDPPQPLHAER SRQCPGRGVR AAKRGGGSG GATHKSKKSK ELDEVAAFEQ LMCDWDDAFAADCYDNERAA RMARLAAEFG QHHGRGFVFV RSRLDKRSRKARNDSGASK FGAAAKALSV EQGTPLENNP QLHLLSWTAC YIASSQLDL GGLFSTQEGV LLPDGSGLLT DGSSGASGSNAADAVGELQR VLRGQDLSQL RGYVGAPPQA RPASGSDDDG SSTTGSNNGA AGEGESEVEEG TAMCGIIRRPE PESGELVULLSCKIGGKPAV GAELLAVAQA EDGKHAPGAS PDTRLCKEPSQS AFDLWSFG WMNKIVPAAAR RGEVEVADLP LPEAQQAEPYC EELNTNWEA AVQEAKKAGK EPKLMKVWLW TYKGDIVLAGIFKLMWSVVF ILGAYYFTRS ILMCIRTLEG KDSDIYDTEWKGVWLTGFFF LDAWLLGMML QRMAFNCLKV GIKARAALTTMIARKCYNMA HLTKDAAEA VGFVASDINK VFEGIQEVHYLWGAPVEAGA ILALLGTLVG VYCIGGVII CMVPLQYYFGYKIIKNNKIK NAPNVTTERWS IIQEBILPAMK LVKYYAWERFPEKHDADMRT RERHYMFWNA VVKTVNVMTMV FGVPPMVTFAVLVPPYELWHV DSSTSEPYIK PQTAFTMLSL FNVLRFPLVWLPKAMRCVSE ALRSVGNLK FLAEPVAPRQ DLEGKPGAQSKAVLRHEMD TSGFTLRRVPE FSVKAGELVA VVGRVGAGKS SILQAMLGNM QTASGLAKCQ HSASSCLPFL VEGTAHSGGR IAYVPQTAWC QNLSLRDNIT FGQPWDEAKY KQVIHACALELDSLAILAAGD QSKAGLRGIN LSGGQRQLRN LARCAYFDGD LVLLDNALSA VDHHTAHIF EHCVRGMFRD KATVLVTHQVEFLPQCDKVA IMDDGTCVYF GPWNAAAQQL LSKYLPASHL LAAGGNAEQP RDTKVVKK EETKKTEDAG KAKRVHSASLTLKSALWEYC WDARWIIFCL SLFFFLTAQA SRQLADYFIRWWTRDHYNKY GVLCIDEGDN PCGFLFYVQY YGILGLLCFI VLMAFRGAFL YTWSLGASYR QHEKSIHRVL YAPLGFFLTTPVGDLLVSFT KDQDVMDDAL PDALYYAGIY GLILLATAITVSVTIPLFSZ LAGGLFVVSC IMLAIYLPAA THLKKLRMGTSGDVVTIJAAL ALDGLGVIAQ YGKQAYFTTI TSQYVNDAHR ALFGAESLNL WLAFICDFFC ACMVLSVACF GIGQWSTLGS SSVGLAFSQS IQMLVFTW S IRLVAECIGL FGSAEKIAWL ANHTPOEAGS LDPPSLPGSC ETKAAPKKRG TAGKFLPPLKDEDLAIPTG GPKVPGWPR TGVLNFNQVV MKYAPHLPPA LRGVSPKVKS GDKVGVVGRT GSGKSTLLA LYRMFNLESGAITLDGIDIS TLTLQQLRRG LSVIPQEPTV FSGTVRTNLPFGEFGADAI LWEALRDCGL EEEQVKACGGL DAKLDGTGGN AWSIGQQQLM CLARAALKV PVLCLEDATA AMDPHTEAHV LEIIERIFSD RTMLTIAHRL DNVIIRSDLVV VMDAGQCEM GTPDELLANP _QSAFSQLVDK TGAASAAAALR KMAADFLLDER ARGQKLGKRP RPSLESHIC VAPSPLSLLS TLLFPFAFMA NVTALLLPKP VLSHAPVSSQ TVNTYIIRLN IQLQCNVLHP ATKEATWSSR RITFTTAHLSS SGSKPPPPLP PLTELPGRG LDWSSAGYRD GREAIPSPSA KYSAADYGAA GDGVTDDTQA LQVAVAAAE DDEGGVVYLG AGTFVLTQPL STAGSNVIR GAGEDATITF VPLPLSDVFP GTWSMDASGK VTSPWITRGGLFLAFSGRRTK SSDSSTLLAT VAGSVEQGAS VIPVDSTAEOF RLGGQWVRIII NDASTDASAG GGTLERGSSE VQESETMIAE GATGGGAGVR AQWTGVLHAF EPTVQCSGVE QLTIRFNHSM MAAHLAERGY NAIELEDVVD CWIRQVTILN ADNAIRLRT DHSTLGSQAC SGGGVVAVVP VVCRGLPSP ADTVGVTEL RWEPTDREVN GHHAITVSKG HANLVTTRPRI TAPFYHDISL EGGALLNVIS SGGGANLNLD LHRSGPWGNL FSQLGMLAA RPFDAGGRDG RGAHAGRQNT FWNLQPGDVA AAAPALQPSA AAGDARRLLV DGDSLLHAGT GQARLLRQLE ADDSAEPLLPSCEFGPLLN FVGGFAGELC KSSGWLVLGAGL PDDRPDLHAS QVTARLQHGA ADNKTHA	EFN52914.1	SEQ.	ID. NO. 70
<i>Synechococcus elongatus</i> PCC 7942.J	MDFLSNFLMD FVKQLQSPTL SFLIGGMVIA ACGSQQLIPE SICKIIVFML LTKIGLTGGM AIRNSNLTEM VLPALFSVAI GILIVIFIARY TLARMPKVKT VDAAITGGLF GAVSGSTMAA ALTLLEEQKI PYEAWAGALY PFMDIPALVT AIVVANIYLKKRKEAAFA SAQGAYSQDP VAAGDYSSSS DYPSSRREYAQQESGDHRVK IWPIVEESLQ GPALSAMLLG VALGLFARPE	ABB57505.1	SEQ.	ID. NO. 71

TABLE D8-continued

Exemplary plasma membrane localized Bicarbonate transporters			
Organism	Sequence	Accession Number	SEQ. ID. NO.
	SVYEGFYDPL FRGLLSILML VMGMMEAWSRI SELRKVAQWY VVYSIVAPLA HGFIAFGLGM IAHYATGFSM GGVVVLAVIA ASSSDISGPP TLRAGIPSAN PSAYIGASTA IGTPVAIGIA IPLFLGLAQT IGG		
<i>Synechocystis</i> sp. PCC 6803	MDFLSNFTLD FVGQLQSPTL AFLIGGMVIA ALGTQLVPE NP_441340 AISTIIIFVML LTKIGLTGGM AIRNSNLTEM LLPVAFSIVL GILIVIFIARY TLAKLPNVRT VDAIATGGLF GAVSGSTMAA ALTTLEEKI SYEAWAGALY PFMDIPALVT AIVVANIYLN KRKRKSAAAS IEESFSKQPV AAGDYGDQTD YPRTRQEYLS QQEPEDNRVK IWPIIEESLQ GPALSAMLLG LALGIFTKPE SVYEGFYDPL FRGLLSILML IMGMMEAWSRI GELRKVAQWY VVYSLIAPIV HGFIAFGLGM IAHYATGFSL GGVVVLAVIA ASSSDISGPP TLRAGIPSAN PSAYIGSSTA IGTPIAIGVC IPLFIGLAQT LGAG		SEQ. ID. No. 72
<i>Nostoc</i> sp. PCC 712	MDFFSLFVMD FVKQLQSPTL GFLIGGMVIA ALGSELIYPE NP_486174 AICQIIVFML LTKIGLTGGI AIRNSNLTEM VLPAASAVAV GVLVVIFIARY TLAKLPKVNT VDAIATGGLF GAVSGSTMAA ALTLLEEKI PYEAWAAALY PFMDIPALVT AIVVANIYLN KKKRSAAGEY LSKQSVAAAGE YPDQDYPSS RQEYLRKQQS ADNRVKIWIPI VKESLQGPAL SAMLLGIALG LFTQPESYK SFYDPLFRGL LSILMLVMGM EAWSRIGELR KVAQWYVVYS VVAPLVHGFI AFGLGMIAHY ATGFSLGGVV ILAVIAASSS DISGPTTLRA GIPSANPSAY IGASTAIGTP IAIGLAIPLF LGQAQAIIGGR		SEQ. ID. No. 73
<i>Cyanothece</i> sp. PCC 7425	MDFWSYFLMD FVKGQLQSPTL GFLIGGMVIA ALGSQLVPE YP_002485721 AICQIIVFML LTKIGLTGGM AIRNSNLTEM VLPAAFSIV GILIVIFIARY TLAKLPKVRT VDAIATGGLF GAVSGSTMAA ALTLLEEKI PYEAWAGALY PFMDIPALVT AIVVANIYLN KKKRRAESEA LSKQBYLGKQ SIVAGDYPAQ QDYPSTRQEY LSKQQGPENN RVKIWPIVQE SLQGPALSAM LLGVALGILT KPESVESFY DPLFRGLLSI LMVLVMGMEAW SRIGELKVA QWYVVYVSVA PVFHGLIAFG LGMFHAYTMG FSMGGVVLA VIASSSSDIS GPPTLRAGIP SANPSAYIGA STAIGTPIAI GLCIPFFIGL AQTLGGG		SEQ. ID. No. 74
<i>Microcysti</i> <i>aeruginosa</i> NIES-843	MDFFSLFVMD FIQQLQSPTL AFLIGGMIIA ALGSELVIPE YP_001661223 SICIIIFVML LTKIGLTGGI AIRNSNLTEM VLPMIFAVIV GIIVVVIFIARY TLANLPKVKV VDAIATGGLF GAVSGSTMAA GLTVLEEQKM PYEAWAGALY PFMDIPALVT AIVVANIYLN KKKRKEAAYD QESFSKQPVA AGNYSDQDY PSSRQEYLSQ QQPADNRVKI WPIIEESLRG PALSAMLLGL ALGIFTQPE VYKSFYDPLF RGLLSVLMLV MGMEAWSRVG ELRKVAQWYV VYSVIAFPVH GLIAFGLGM AHYATGFSWG GVVMLAVIAS SSSDISGPT LRAGIPSANP SAYIGASTAI GTPVAIGLCI PFFVGLAQL SGG		SEQ. ID. No. 75
<i>Anabaena</i> <i>variabilis</i> ATCC 29413	MDFVSLFVKD FIAQLQSPTL AFLIGGMIIA ALGSELVIPE YP_323532 SICIIIFVML LTKIGLTGGI AIRNSNLTEM VLPMIFAVIT GITIVFISRY TLAKLPKVKV VDAIATGGLF GAVSGSTMAA GLTVLEEQKM AYEAWAGALY PFMDIPALVT AIVVANIYLN KKKRKEAIVMS TEQPVAAQGDY PDQKDYPSSR QEYLSSQKGD EDNRVKIWIPI IEESLRGPAL SAMLLGLALG LFTQPESYK SFYDPAFRGL LSILMLVMGM EAWSRIGELR KVAQWYVVYS VVAPFVHGFI AFGLGMIAHY TMNFSMGGVV ILAVIAASSS DISGPTTLRA GIPSANPSAY IGASTAVGTP VAIGLCIPFF LGQAQAIIGG		SEQ. ID. No. 86
<i>Cyanothece</i> sp. PCC 880	MDFLSLFVKD FIIQLQSPTL AFLIGGMVIA ALGSELVIPE YP_002371470.1 SEQ. ID. No. 87 SICIIIFVML LTKIGLTGGI AIRNSNLTEM VLPMICAVIV GIVVVIFIARY TLAKLPKVNV VDAIATGGLF GAVSGSTMAA GLTVLEEQKI PYEAWAGALY PFMDIPALVT AIVVANIYLN KKKRKATVMO ESSLQKPVAA GDYPSSRQEY VSQQQPEDNR VKIWPPIEES LRGPALSAM LGLALGILTQ PESVYKGYD PPFRGGLSIL MLVMGMMEAW SRIGELKVAQ WVVYVSAAP FIHGLLAFLG GMIAHYTMGF SMGGVILAV IASSSSDISG PPTLRAGIPS ANPSAYIGAS TAIGTPVAIG LCIPFFFVGLA QAIGGF		
<i>Arthrospha</i> <i>piatensis</i> str. Paraca	MDFLSGFLTR FLAQLQSPTL GFLIGGMVIA AVNSQLQIPD ZP_06383808.1 SEQ. ID. No. 88 AIYKFVVFML LMKVGLSGGI AIRGSNLTEM LLPAVFALVT GIVIVFIGRY TLAKLPNVKT VDAIATAGLF GAVSGSTMAA ALTLLEEQGM EYEAWAAALY PFMDIPALVS AIVLASIYVS		

TABLE D8-continued

Exemplary plasma membrane localized Bicarbonate transporters			
Organism	Sequence	Accession Number	SEQ. ID. NO
	KQKHSDMADE SLSKHESLSK QPVAAGDYP S KPEYPTTRQE YLSQQRGSAN QGVEIWIPIK ESLQGSALSA LLLGLALGLL TRPESVFSF YEPLFRGLLS IILMLVMGMEA TARLGELRKV AQWYAVYAFI APPLHGGLIAF GLGMIAHVVT GFSLGGVVIL AVIASSSSDI SGPTTLRAGI PSANPSAYIG SSTAVGTPVA IALGIPLYIG LAQALMGG		

TABLE D9

Exemplary chloroplast envelope localized Bicarbonate transporters			
Organism	Sequence	Accession Number	SEQ. ID. NO
<i>Chlamydomonas reinhardtii</i>	MQTTMTRPCL AQPVLRSRVL RSPMRVVAAS APTAVTTVVT SNNGNGHfq AATTVPVPTE AVPAVASAPVR AVSVLTPPOV YENAINVGAY KAGLTPLATF VQGIQAGAYI AFGAFLAISV GGNIPGVAAA NPGLAKLLFA LVFPVGLSMV TNCGAELFTG NTMMMLTCALI EKKATWGQLL KNWSVSYFGN FVGSIAMVAA VVATGCLTN TLPVQMATLW ANLGFTEVLS RSILCNWLVC CAVWSASAAAT SLPGRILALW PCITAFVAIG LEHSVANMFV IPLGMMLGAE VTWSQFFFNN LIPVTLGNTI AGVLMMAIAY SISFGSLGKS AKPATA	BAD16681.1	SEQ. ID. NO. 89
<i>Volvox carteri f. nagariensis</i>	MQTTMSVTRP CVGLRPLPVR NVRSLIRQAQ APQQVSTAVS TNNGNGVAA ASLSVPAPVA APAQAVSTPV RAVSVLTPPO VYENANVGA YKASLGVLAT FVQGIQAGAY IAFGAFLACS VGGNIPGITA SNPGLAKLLE ALVFPVGLSM VTNCGAELYT GNTMMMLTCAI FEKKATWAQL VKNWWVSYAG NFVGSIAMVA AVVATGLMAS NQLPVNMATA KSSLGFTEVL SRSILCNWLVC CCAVWSASAA TSLPGRILGK WPPITAFVAI GLEHSVANMF VIPLGMLGA DVTWSQFFFNN NLVPVTLGNT IAGVVMMAVA YSVSYGSLGK TPKPATA	XP_002951507.1	SEQ. ID. NO. 79

TABLE D10

Transit Peptides		
Organism	SEQ ID NO	Name
<i>Arabidopsis thaliana</i>	8	Rbcs-1a transit peptide
<i>Arabidopsis thaliana</i>	14	PGR5 transit peptide
<i>Arabidopsis thaliana</i>	15	psdA transit peptide
<i>Arabidopsis thaliana</i>	22	DNAJ transit peptide
<i>Cyanophora paradoxa</i>	102	psdA transit peptide
<i>Arabidopsis thaliana</i>	104	CAB transit peptide
<i>Arabidopsis thaliana</i>	105	PGR5 transit peptide

TABLE D11-continued

Cyclic Electron Transfer modulator proteins							
40	45	50	55	60			
Organism	SEQ ID NO	Name	Organism	SEQ ID NO	Name	No.	Function
<i>Arabidopsis thaliana</i>	95	Ferredoxin2	<i>Arabidopsis thaliana</i>	95	Ferredoxin2	AAG40057.1	cyclic electron transfer modulator protein
		(FD2)					
<i>Arabidopsis thaliana</i>	96	ferredoxin-	<i>Arabidopsis thaliana</i>	96	ferredoxin-	AT5G66190	cyclic electron transfer modulator protein
		NADP(+)			NADP(+)	partial	
						oxidoreductase	
			<i>Arabidopsis thaliana</i>	97	ferredoxin-	BAH19611.1	cyclic electron transfer modulator protein
					NADP(+)		
						oxidoreductase	
						(FNR1)	
						(FNR2)	

TABLE D11

Cyclic Electron Transfer modulator proteins			
Organism	SEQ ID NO	Name	Accession No.
<i>Arabidopsis thaliana</i>	93	Ferredoxin1 (FD1)	AEE28669.1

65 An exemplary optimized DNA sequence for the plasma membrane localized bicarbonate transporter is shown in SEQ ID NO. 91

US 11,001,853 B2

61

62

(SEQ ID NO: 91)

atgctgcccgtc	gcctggcgta	catactgtgt	gtgctgcccc	tgcagtacta	cttcggctac	60
aagatcgtgc	agatcaagct	gcagaacgcc	aagcacgtcg	ccctgegcctc	cgcacatcg	120
caggaggtgc	tgcccgccat	caagctggtc	aagtactacg	cctggagca	gttcttttag	180
aaccagatca	gcaaggctcg	ccgcgaggag	atccgcctca	acttctggaa	ctgcgtgtat	240
aaggtecatca	acgtggctcg	cgtgttctgc	gtgcccggcca	tgaccggctt	cgtcatcttc	300
accacctacg	agttccagecg	ccgcggccctg	gtgtccagcg	tgcgccttac	caccctgtcg	360
ctgttcaaca	ttcttgccgtt	ccccctggtc	gtgctgcccc	aggccctgctg	tgccgtgtcc	420
gaggccaacg	ogtctctcca	gcccgtggag	gcctacgtgc	tggaggaggt	gcgcctgggc	480
actgcccggcg	tcaagacccc	caagaacgt	ccccccggcg	cgtcatcga	gaacgggtgt	540
ttccaccacc	cctccaaccc	caactggcac	ctgcacgtgc	ccaagttcga	ggtcaagccc	600
ggccaggtcg	ttgctgtgg	ggggccgcatt	ggccggccgca	agtcgtccct	ggtgcaggcc	660
atccteggc	acatggtaa	ggagcacggc	agttcaacg	tggccggccg	catctccat	720
gtgcccggaga	acccctggct	gcagaacctg	tccctgcgtg	acaacgtgt	gtttggcgag	780
cagttcgatg	agaacaagta	cacccggcgtc	atcgagtc	gcgcctgtac	cctggacactg	840
cagatcctgt	ccaaacggta	ccagtcctaa	gccggcatcc	gcccgtgtcaa	cttctccgg	900
ggccagggcc	agcgcgtgaa	cctggccggc	tgcgcctacg	ccgacgcccga	cctgggtgt	960
ctcgacaacg	ccctgtccgc	cgtggaccac	cacaccgccc	accacatctt	cgacaagtgc	1020
atcaaggccc	ttttctccga	caaggccgtg	gtgctggta	cccaccat	cgagttcat	1080
ccccgtcg	acaacgtggc	catcatggac	gaggccgct	gcctgtactt	cgcaagtgg	1140
aacgaggagg	cccagcacct	gctcgccaa	ctgctgcccc	tcacccac	gctgcacgccc	1200
gccggctccc	aggaggctcc	ccccggcccc	aagaagaagg	ccgaggacaa	ggccggccccc	1260
cagaagtgc	agtcgtgca	gctgacccctg	gcgcctac	ccatcgccaa	gcccaccgag	1320
aagcccaagg	acgtccagaa	gctgactgtc	taccaggccg	ccctcatcta	cacccgttac	1380
ggcaacctgt	ttctgggtgg	cgtgtgttcc	ttttcttcc	tggccggctca	gtgctctcg	1440
cagatctccg	atttctgggt	gcccgtgg	gtgaacgacg	agtacaagaa	gttccccgt	1500
aagggccgagc	aggactccggc	cgccaccacc	ttctactgtc	tcatctact	gctgtgtgt	1560
ggcctgttct	acatcttcat	gatcttccgc	ggccactt	ttctgtgg	ggtgcgtcaag	1620
tcctcgaga	ccatccgcag	gaaggccctg	cacaacgtcc	tcaacgcgcc	catggccctc	1680
ttccctggta	ccggccgtcg	cgacccgtcg	ctcaactca	ccaaggacca	ggacattat	1740
gatgagaacc	tgcccgatgc	cgttacttc	atggccat	acggccgtat	tctgtggcg	1800
accaccatca	ccgtgtccgt	caccatcaac	ttttccggcc	ccttcaccgg	cgcgtgtatc	1860
atcatgaccc	tcatcatgt	ctccatctac	ctggccggcc	ccactgtcc	gaagaaggcg	1920
cgcgcgtgt	ctggccggat	gctgggtcg	ctgggttgg	aggttctgg	gggccttggc	1980
gtgggttccgg	ccttcaacaa	gcaggaggtac	ttcattgagg	aggccggccc	ccgcaccaac	2040
atcaccaact	ccgcgcgtctt	caacgcgcag	gctgtgaacc	tgtggctggc	tttctgggt	2100
gacttcatcg	gcgcctgcct	ggtggggcg	gtgtccgcct	tgcgcgtgg	catggccaa	2160
gacccctggcg	gcgcgcaccgt	ccgcctggcc	tttccaaaca	tcattcagat	gcttgggttc	2220
taacacccgg	ttgtccgcctt	catctccgag	tccatctcc	tcttcaactc	cgtcgagg	2280
atggccctacc	tcggccacta	cgtggcccccac	gatgggtgt	tctatgacca	gcccagaag	2340
gacggcgatcg	ccaaagcaaat	cgttctgccc	gacggcaaca	tcgtgcccc	ccgcctcaag	2400

-Continued

gtccaggatcg	tgggtgacga	cgcgcgcctc	gcccgtggc	ctgccaccgg	caacatccgc	2460
ttcgaggacg	tgtggatgca	gtaccgcctg	gacgctcctt	gggctctgaa	ggcggtcacc	2520
ttcaagatca	acgacggcga	gaaggctggc	gcccgtggcc	gcacccggctc	cggcaagtcc	2580
accacgcgtgc	tggcgctgt	ccgcgtgttc	gagctggca	agggccgcatt	cctgggtcgc	2640
ggcgtggaca	tcgcccaccc	gtcgctcaag	cgccctgcgc	ccggccgtgc	catcattccc	2700
caggagcccg	tcatgttac	cggcacccgt	cgctccaacc	tggaccacc	cgcgagatcc	2760
aaggacgtat	ccattctgt	ggagggtgct	aagaaggatcg	gcctcgagga	ccaggcgccag	2820
cacggccggcg	gcctggacgg	ccaggatcgat	ggcacccggc	gcaaggccctg	gtctctggc	2880
cagatgcgc	tgggtgtgcct	ggctcgcc	gcctcgccgc	ccgtgeccat	cctgtgcctg	2940
gacgaggctc	ccggcccat	ggacccgcac	actgaggcca	tcgtgcagca	gaccatcaag	3000
aagggttgc	acgacccgcac	caccatcacc	attgcccacc	gcctggacac	catcatcgag	3060
tccgacaaga	tcatcgat	ggagcagggc	tcgctgtatgg	agtacgagtc	gcctcgaa	3120
ctgctcgcca	accgcgactc	catgttctcc	aagctggatcg	acaagaccgg	ccccggcc	3180
gccgctgcgc	tgcgcaagat	ggccgaggac	ttctggtcca	ctcgctccgc	gcagggccgc	3240
aaccagtaa						

An exemplary optimized DNA sequence for Chloroplast envelope localized Bicarbonate transporter is shown in SEQ ID NO: 92

combination of heterologous nucleotide sequences encoding additionally a Rubisco (for example SEQ ID NO:107). Further still a transit peptide amino acid sequence at the

(SEQ ID NO: 92)

atgcagacca	ctatgactcg	cccttgcctt	gcccagcccg	tgctgegatc	tcgtgtgctc	60
cggtcgccta	tgcggttgtt	tgcagcgacg	gtccctaccc	cggtgacgac	agtcgtgacc	120
tcgaatggaa	atggcaacgg	tcatttccaa	gctgctacta	cgcccgtgcc	ccctactccc	180
gtccccgtcg	ctgtttccgc	gcctgtgcgc	gtctgtcg	tgctgactcc	tcctcaagt	240
tatgagaacg	ccattaatgt	tggcgctac	aaggccgggc	taacgcctt	ggcaacgttt	300
gtccaggcaca	tccaaggccgg	tgcctacatt	gcgttccggc	ccttcctcgc	catctccgt	360
ggaggcaaca	tccccggcgt	cgccgcccgc	aaccccgcc	tggccaagct	gtatattgt	420
ctgggtttcc	ccgtgggtct	gtccatggtg	accaactgcg	gcgcccagat	gttacggc	480
aacaccatga	tgctcacatg	cgcgcgtc	gagaagaagg	ccacttgggg	gcagcttctg	540
aagaacttgg	gcgtgttcta	tttggcaac	ttctgtggct	ccatcgocat	ggtcggcc	600
gtgggtggcca	ccggctgcct	gaccaccaac	accctgcctt	tgcagatggc	caccctcaag	660
gccaacctgg	gttccacca	ggtgctgtcg	cgctccatcc	tgtgcaactg	gtgggtgtc	720
tgcgcgcgtgt	ggtccgcctc	cgccgcacc	tgcgtgccc	gcccgcac	ggcgctgtgg	780
cctgcata	ccgccttcgt	ggccatcgcc	ctggagact	ccgtcgccaa	catgttcgt	840
atccctctgg	gcatgtatgt	ggggctgttag	gtcacgttga	gcacgttctt	tttcaacaac	900
ctgatecccc	tcacccctgg	caacaccatt	gtggcgatcc	tcatgtatgc	catcgccatc	960
tccatctcg	tccgtccct	cgcaagcccg	ccaccggcg			1008

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the disclosure specifically described herein. For example a transgenic plant or alga of an embodiment disclosed herein further comprising within its genome, and expressing or overexpressing, a

amine terminal portion of a protein sequence identified herein may be cleaved leaving the protein sequence alone. The percent homology applies to the protein sequence without the transit peptide sequence also. Such equivalents are intended to be encompassed within the scope of the following claims.

1. Hausler R E, Hirsch H J, Kreuzaler F, Peterhansel C (2002) Overexpression of C(4)-cycle enzymes in transgenic C(3) plants: a biotechnological approach to improve C(3)-photosynthesis. *J Exp Bot* 53: 591-607.
2. Goldschmidt E E, Huber S C (1992) Regulation of photosynthesis by end-product accumulation in leaves of plants storing starch, sucrose, and hexose sugars. *Plant Physiol* 99: 1443-1448.
3. Duanmu D, Miller A R, Horken K M, Weeks D P, Spalding M H (2009) Knockdown of limiting-CO₂-induced gene HLA3 decreases HCO₃-transport and photosynthetic Ci affinity in *Chlamydomonas reinhardtii*. *Proc Natl Acad Sci USA* 106: 5990-5995.
4. Moroney J V, Jungnick N, Dimario R J, Longstreth D J (2013) Photorespiration and carbon concentrating mechanisms: two adaptations to high O₂, low CO₂ conditions. *Photosynth Res* 117: 121-131.
5. Wang Y, Duanmu D, Spalding M H (2011) Carbon dioxide concentrating mechanism in *Chlamydomonas reinhardtii*: inorganic carbon transport and CO₂ recapture. *Photosynth Res* 109: 115-122.
6. Perrine Z, Negi S, Sayre R (2012) Optimization of photosynthetic light energy utilization by microalgae. *Algal Research* 1: 134-142.
7. Elleby B, Chirica L C, Tu C, Zeppezauer M, Lindskog S (2003) Characterization of carbonic anhydrase from *Neisseria gonorrhoeae*. *Eur J Biochem* 286: 1613-1619.
8. Subramanian S, Barry A N, Pieris S, Sayre R T (2013) Comparative energetics and kinetics of autotrophic lipid and starch metabolism in chlorophytic microalgae: implications for biomass and biofuel production. *Biotechnol Biofuels* 6: 150.
9. Nakamura N, Iwano M, Havaux M, Yokota A, Munekage Y N (2013) Promotion of cyclic electron transport around photosystem I during the evolution of NADP-malic enzyme-type C4 photosynthesis in the genus *Flaveria*. *New Phytol* 199: 832-842.
10. Kramer D M, Evans J R (2011) The importance of energy balance in improving photosynthetic productivity. *Plant Physiol* 155: 70-78.
11. Alric J (2010) Cyclic electron flow around photosystem I in unicellular green algae. *Photosynth Res* 106: 47-56.
12. Amunts A, Drory O, Nelson N (2007) The structure of a plant photosystem I supercomplex at 3.4 Å resolution. *Nature* 447: 58-63.
13. Breyton C, Nandha B, Johnson G N, Joliot P, Finazzi G (2006) Redox modulation of cyclic electron flow around photosystem I in C3 plants. *Biochemistry* 45: 13465-13475.
14. Cardol P, Forti G, Finazzi G (2011) Regulation of electron transport in microalgae. *Biochim Biophys Acta* 1807: 912-918.
15. Hanke G T, Okutani S, Satomi Y, Takao T, Suzuki A, et al. (2005) Multiple iso-proteins of FNR in *Arabidopsis*: evidence for different contributions to chloroplast function and nitrogen assimilation. *Plant Cell Environ* 28: 1146-1157.
16. Johnson G N (2011) Physiology of PSI cyclic electron transport in higher plants. *Biochim Biophys Acta* 1807: 384-389.
17. Okutani S, Hanke G T, Satomi Y, Takao T, Kurisu G, et al. (2005) Three maize leaf ferredoxin:NADPH oxidoreductases vary in subchloroplast location, expression, and interaction with ferredoxin. *Plant Physiol* 139: 1451-1459.
18. Slewinski T L, Braun D M (2010) Current perspectives on the regulation of whole-plant carbohydrate partitioning. *Plant Science* 178: 341-349.
19. Arrivault S, Guenther M, Ivakov A, Feil R, Vosloh D, et al. (2009) Use of reverse-phase liquid chromatography, linked to tandem mass spectrometry, to profile the Calvin cycle and other metabolic intermediates in *Arabidopsis* rosettes at different carbon dioxide concentrations. *Plant Journal* 59: 824-839.
20. Huege J, Sulpice R, Gibon Y, Lisec J, Koehl K, et al. (2007) GC-EL-TOF-MS analysis of in vivo carbon-partitioning into soluble metabolite pools of higher plants by monitoring isotope dilution after (CO₂)—C-13 labelling. *Phytochemistry* 68: 2258-2272.
21. Romisch-Margl W, Schramek N, Radykewicz T, Ettenhuber C, Eylert E, et al. (2007) (CO₂)—C-13 as a universal metabolic tracer in isotopologue perturbation experiments. *Phytochemistry* 68: 2273-2289.
22. Sekiyama Y, Kikuchi J (2007) Towards dynamic metabolic network measurements by multi-dimensional NMR-based fluxomics. *Phytochemistry* 68: 2320-2329.
23. Szecowka M, Heise R, Tohge T, Nunes-Nesi A, Vosloh D, et al. (2013) Metabolic fluxes in an illuminated *Arabidopsis* rosette. *Plant Cell* 25: 694-714.
24. Ma F, Jazmin L J, Young J D, Allen D K (Submitted) Isotopically nonstationary 13C flux analysis of *Arabidopsis thaliana* leaf metabolism at varying light intensities. *Proc Natl Acad Sci USA*.
25. Shastri A A, Morgan J A (2007) A transient isotopic labeling methodology for 13C metabolic flux analysis of photoautotrophic microorganisms. *Phytochemistry* 68: 2302-2312.
26. Young J D, Shastri A A, Stephanopoulos G, Morgan J A (2011) Mapping photoautotrophic metabolism with isotopically nonstationary (13)C flux analysis. *Metab Eng* 13: 656-665.
27. Young J D (Submitted) INCA: A computational platform for isotopically nonstationary metabolic flux analysis. *Bioinformatics*.
28. Young J D, Walther J L, Antoniewicz M R, Yoo H, Stephanopoulos G (2008) An elementary metabolite unit (EMU) based method of isotopically nonstationary flux analysis. *Biotechnol Bioeng* 99: 686-699.
29. Masclaux-Daubresse C, Chardon F (2011) Exploring nitrogen remobilization for seed filling using natural variation in *Arabidopsis thaliana*. *J Exp Bot* 62: 2131-2142.
30. Hay R K M, Gilbert R A (2001) Variation in the harvest index of tropical maize: Evaluation of recent evidence from Mexico and Malawi. *Annals of Applied Biology* 138: 103-109.
31. Russell W A (1985) Evaluation for plant, ear and grain traits of maize cultivars representing seven years of breeding. *Maydica* 30: 85-96.
32. Sinclair T R (1998) Historical changes in harvest index and crop nitrogen accumulation. *Crop Science* 38: 638-643.
33. Victorio R G, Moreno U, Black Jr C C (1986) Growth, partitioning, and harvest index of tuber-bearing *Solanum* genotypes grown in two contrasting Peruvian environments. *Plant Physiology* 82: 103-108.
34. Vos J (1997) The nitrogen response of potato (*Solanum tuberosum* L.) in the field: Nitrogen uptake and yield, harvest index and nitrogen concentration. *Potato Research* 40: 237-248.

35. Parry M A, Andralojc P J, Scales J C, Salvucci M E, Carmo-Silva A E, et al. (2013) Rubisco activity and regulation as targets for crop improvement. *J Exp Bot* 64: 717-730.
36. Sage R F (2002) Variation in the k(cat) of Rubisco in C(3) and C(4) plants and some implications for photosynthetic performance at high and low temperature. *J Exp Bot* 53: 609-620.
37. Henkes S, Sonnewald U, Badur R, Flachmann R, Stitt M (2001) A small decrease of plastid transketolase activity in antisense tobacco transformants has dramatic effects on photosynthesis and phenylpropanoid metabolism. *Plant Cell* 13: 535-551.
38. Miyagawa Y, Tamoi M, Shigeoka S (2001) Overexpression of a cyanobacterial fructose-1,6-/sedoheptulose-1,7-bisphosphatase in tobacco enhances photosynthesis and growth. *Nature Biotechnology* 19: 965-969.
39. Peterhansel C, Blume C, Offermann S (2013) Photorespiratory bypasses: how can they work? *J Exp Bot* 64: 709-715.
40. Blanco N E, Ceccoli R D, Via M V, Voss I, Segretin M E, et al. (2013) Expression of the minor isoform pea ferredoxin in tobacco alters photosynthetic electron partitioning and enhances cyclic electron flow. *Plant Physiol* 161: 866-879.
41. Busch K B, Deckers-Hebestreit G, Hanke G T, Mulkidianian A Y (2012) Dynamics of bioenergetic microcompartments. *Biol Chem* 394: 163-188.
42. Minagawa J (2011) State transitions—the molecular remodeling of photosynthetic supercomplexes that controls energy flow in the chloroplast. *Biochim Biophys Acta* 1807: 897-905.
43. Peltier G, Tolteeter D, Billon E, Cournac L (2010) Auxiliary electron transport pathways in chloroplasts of microalgae. *Photosynth Res* 106: 19-31.
44. Peng L, Shikanai T (2011) Supercomplex formation with photosystem I is required for the stabilization of the chloroplast NADH dehydrogenase-like complex in *Arabidopsis*. *Plant Physiol* 155: 1629-1639.
45. Takahashi H, Clowez S, Wollman F A, Vallon O, Rappaport F (2013) Cyclic electron flow is redox-controlled but independent of state transition. *Nat Commun* 4: 1954.
46. Neale A P, Blunder T, Wunder T, Pesaresi P, Pribil M, et al. (2013) PGRL1 is the elusive ferredoxin-plastoquinone reductase in photosynthetic cyclic electron flow. *Mol Cell* 49: 511-523.
47. DalCorso G, Pesaresi P, Masiero S, Aseeva E, Schunemann D, et al. (2008) A complex containing PGRL1 and PGR5 is involved in the switch between linear and cyclic electron flow in *Arabidopsis*. *Cell* 132: 273-285.
48. Shikanai T (2014) Central role of cyclic electron transport around photosystem I in the regulation of photosynthesis. *Current Opinion in Biotechnology* 26: 25-30.
49. Walter J M, Greenfield D, Liphardt J (2010) Potential of light-harvesting proton pumps for bioenergy applications. *Curr Opin Biotechnol* 21: 265-270.
50. Dioumaev A K, Brown L S, Shih J, Spudich E N, Spudich J L, et al. (2002) Proton transfers in the photochemical reaction cycle of proteorhodopsin. *Biochemistry* 41: 5348-5358.
51. Friedrich T, Geibel S, Kalmbach R, Chizhov I, Ataka K, et al. (2002) Proteorhodopsin is a light-driven proton pump with variable vectoriality. *J Mol Biol* 321: 821-838.
52. Govindjee R, Ebrey T G, Crofts A R (1980) The quantum efficiency of proton pumping by the purple membrane of *Halobacterium halobium*. *Biophys J* 30: 231-242.

53. Govindjee R, Imasheva E S, Misra S, Balashov S P, Ebrey T G, et al. (1997) Mutation of a surface residue, lysine-129, reverses the order of proton release and uptake in bacteriorhodopsin; guanidine hydrochloride restores it. *Biophys J* 72: 886-898.
54. Govindjee R, Misra S, Balashov S P, Ebrey T G, Crouch R K, et al. (1996) Arginine-82 regulates the pKa of the group responsible for the light-driven proton release in bacteriorhodopsin. *Biophys J* 71: 1011-1023.
55. Lakatos M, Lanyi J K, Szakacs J, Varo G (2003) The photochemical reaction cycle of proteorhodopsin at low pH. *Biophys J* 84: 3252-3256.
56. Walter J M, Greenfield D, Bustamante C, Liphardt J (2007) Light-powering *Escherichia coli* with proteorhodopsin. *Proc Natl Acad Sci USA* 104: 2408-2412.
57. Kim J Y, Jo B H, Jo Y, Cha H J (2012) Improved production of biohydrogen in light-powered *Escherichia coli* by co-expression of proteorhodopsin and heterologous hydrogenase. *Microb Cell Fact* 11:2.
58. Froehlich J E, Keegstra K (2011) The role of the transmembrane domain in determining the targeting of membrane proteins to either the inner envelope or thylakoid membrane. *Plant J* 68: 844-856.
59. Beja O, Aravind L, Koonin E V, Suzuki M T, Hadd A, et al. (2000) Bacterial rhodopsin: evidence for a new type of phototrophy in the sea. *Science* 289: 1902-1906.
60. Lindqvist A, Andersson S (2002) Biochemical properties of purified recombinant human beta-carotene 15,15'-monoxygenase. *J Biol Chem* 277: 23942-23948.
61. Roslan H A, Salter M G, Wood C D, White M R, Croft K P, et al. (2001) Characterization of the ethanol-inducible alc gene-expression system in *Arabidopsis thaliana*. *Plant J* 28: 225-235.
62. Cao Y, Brown L S, Sasaki J, Maeda A, Needleman R, et al. (1995) Relationship of proton release at the extracellular surface to deprotonation of the schiff base in the bacteriorhodopsin photocycle. *Biophys J* 68: 1518-1530.
63. Joliot P, Johnson G N (2011) Regulation of cyclic and linear electron flow in higher plants. *Proc Natl Acad Sci USA* 108: 13317-13322.
64. Fabre N, Reiter I M, Becuwe-Linka N, Genty B, Rumeau D (2007) Characterization and expression analysis of genes encoding alpha and beta carbonic anhydrases in *Arabidopsis*. *Plant Cell Environ* 30: 617-629.
65. Bihmidine S, Hunter C T, 3rd, Johns C E, Koch K E, Braun D M (2013) Regulation of assimilate import into sink organs: update on molecular drivers of sink strength. *Front Plant Sci* 4: 177.
66. Ihemere U, Arias-Garzon D, Lawrence S, Sayre R (2006) Genetic modification of cassava for enhanced starch production. *Plant Biotechnol J* 4: 453-465.
67. Wunsche J N, Greer D H, Laing W A, Palmer J W (2005) Physiological and biochemical leaf and tree responses to crop load in apple. *Tree Physiol* 25: 1253-1263.
68. Paul M J, Foyer C H (2001) Sink regulation of photosynthesis. *J Exp Bot* 52: 1383-1400.
69. Sonnewald U, Lerchi J, Zrenner R, Frommer W (1994) Manipulation of sink-source relations in transgenic plants. *Plant Cell Environ* 17: 649-658.
70. Sonnewald U, Willmitzer L (1992) Molecular approaches to sink-source interactions. *Plant Physiol* 99: 1267-1270.
71. Willson W J (1972) Control of crop processes In: Rees A R, Cockshull K E, Hand D W, Hurd R G, editors. *Crop Processes in Controlled Environments*: London Academic Press. pp. 7-30.

72. Jonik C, Sonnewald U, Hajirezaei M R, Flugge U I, Ludewig F (2012) Simultaneous boosting of source and sink capacities doubles tuber starch yield of potato plants. *Plant Biotechnol J* 10: 1088-1098.
73. Sweetlove L J, Hill S A (2000) Source metabolism dominates the control of source to sink carbon flux in tuberizing potato plants throughout the diurnal cycle and under a range of environmental conditions. *Plant, Cell and Environment* 23: 523-529.
74. Allen D K, Goldford J, Gierse J, Mandy D, Diepenbrock C, et al. (2013) (submitted) Quantification of peptide m/z distributions from ¹³C-labeled cultures with high resolution mass spectrometry. *Analytical Chemistry*.
75. Choi J, Antoniewicz M R (2011) Tandem mass spectrometry: a novel approach for metabolic flux analysis. *Metab Eng* 13: 225-233.
76. Allen D K, Libourel I G L, Shachar-Hill Y (2009) Metabolic flux analysis in plants: Coping with complexity. *Plant, Cell and Environment* 32: 1241-1257.
77. Allen D K, Laclair R W, Ohlrogge J B, Shachar-Hill Y (2012) Isotope labelling of Rubisco subunits provides in vivo information on subcellular biosynthesis and exchange of amino acids between compartments. *Plant, Cell and Environment* 35: 1232-1244.
78. Allen D K, Shachar-Hill Y, Ohlrogge J B (2007) Compartment-specific labeling information in ¹³C metabolic flux analysis of plants. *Phytochemistry* 68: 2197-2210.
79. Mandy D, Goldford J, Yang H, Allen D K, Libourel I G L (2013) (submitted) Metabolic flux analysis using ¹³C peptide label measurements. *The Plant Journal*.
80. Allen D K, Young J D (2013) Carbon and nitrogen provisions alter the metabolic flux in developing soybean embryos. *Plant Physiol* 161: 1458-1475.
81. Allen D K, Ohlrogge J B, Shachar-Hill Y (2009) The role of light in soybean seed filling metabolism. *Plant Journal* 58: 220-234.
82. Jazmin L J, Young J D (2013) Isotopically nonstationary ¹³C metabolic flux analysis. *Methods Mol Biol* 985: 367-390.
83. Blankenship R E, Tiede D M, Barber J, Brudvig G W, Fleming G, et al. (2011) Comparing photosynthetic and photovoltaic efficiencies and recognizing the potential for improvement. *Science* 332: 805-809.
84. Jazmin L J, O'Grady J, Ma F, Allen D K, Morgan J A, et al. (In press) Isotopically nonstationary MFA (INST-MFA) of autotrophic metabolism. *Methods Mol Biol*.
85. Egnatchik R A, Leamy A K, Noguchi Y, Shiota M, Young J D (In press) Palmitate-induced activation of mitochondrial metabolism promotes oxidative stress and apoptosis in H411EC3 rat hepatocytes. *Metabolism*.
86. Leamy A K, Egnatchik R A, Young J D (2013) Molecular mechanisms and the role of saturated fatty acids in the progression of non-alcoholic fatty liver disease. *Prog Lipid Res* 52: 165-174.
87. Srour O, Young J D, Eldar Y C (2011) Fluxomers: a new approach for ¹³C metabolic flux analysis. *BMC Syst Biol* 5: 129.
88. Young J D, Allen D K, Morgan J A (2014) Isotopomer measurement techniques in metabolic flux analysis II: Mass spectrometry. *Methods Mol Biol* 1083: 85-108.
89. Egnatchik R A, Leamy A K, Jacobson D A, Young J D (Submitted) E R calcium stimulates mitochondrial alterations in hepatic lipotoxicity. *J Biol Chem*.
90. Leamy A K, Egnatchik R A, Shiota M, Young J D (Submitted) Modulating lipid fate controls E R stress and lipotoxicity in palmitate-treated hepatic cells. *FEBS J*.
91. Young J D (In press) Metabolic flux rewiring in mammalian cell cultures. *Curr Opin Biotechnol*.
92. McAtee A G, Templeton N, Young J D (Submitted) Role of CHO central carbon metabolism in controlling the quality of secreted biotherapeutic proteins. *Pharmaceutical Bioprocessing*.
93. Duckwall C S, Murphy T A, Young J D (2013) Mapping cancer cell metabolism with (¹³)C flux analysis: Recent progress and future challenges. *J Carcinog* 12: 13.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 108

```

<210> SEQ ID NO 1
<211> LENGTH: 132
<212> TYPE: PRT
<213> ORGANISM: Arabidopsis thaliana
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<222> LOCATION: (1)..(44)
<223> OTHER INFORMATION: PGR5 Amino acid sequence with chloroplast
transit peptide

```

<400> SEQUENCE: 1

```

Met Ala Ala Ala Ser Ile Ser Ala Ile Gly Cys Asn Gln Thr Leu Ile
1           5          10          15

Gly Thr Ser Phe Tyr Gly Gly Trp Gly Ser Ser Ile Ser Gly Glu Asp
20          25          30

Tyr Gln Thr Met Leu Ser Lys Thr Val Ala Pro Pro Gln Gln Ala Arg
35          40          45

Val Ser Arg Lys Ala Ile Arg Ala Val Pro Met Met Lys Asn Val Asn
50          55          60

Glu Gly Lys Gly Leu Phe Ala Pro Leu Val Val Thr Arg Asn Leu
65          70          75          80

Val Gly Lys Lys Arg Phe Asn Gln Leu Arg Gly Lys Ala Ile Ala Leu

```

US 11,001,853 B2

71

72

-continued

85 90 95

His Ser Gln Val Ile Thr Glu Phe Cys Lys Ser Ile Gly Ala Asp Ala
 100 105 110

Lys Gln Arg Gln Gly Leu Ile Arg Leu Ala Lys Lys Asn Gly Glu Arg
 115 120 125

Leu Gly Phe Leu
 130

<210> SEQ ID NO 2

<211> LENGTH: 402

<212> TYPE: DNA

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 2

atggctgctg	tttcgatttc	tgcaatagga	tgtaatcaa	ctttgatagg	aacttccttc	60
tatggaggat	ggggaaagttc	catctccgga	gaagattacc	aaaccatgct	ctccaagaca	120
gttgcgccac	cgcaacaagc	cagagtctca	aggaaagcaa	tcaagacgt	tccaatgtg	180
aagaatgtca	atgaaggcaa	aggcttattt	gcacctctag	ttgttgtcac	acgcaaccta	240
gtaggcaaga	agaggttaa	tcagctcaga	ggaaaagcca	ttgccttaca	ctctcaggtg	300
atcaactgagt	tttgcaaatac	gattggagca	gatgcaaaac	agagacaagg	gcttatcagg	360
cttgctaaga	agaatggaga	gaggcttggt	ttcccttgctt	ag		402

<210> SEQ ID NO 3

<211> LENGTH: 324

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 3

Met	Gly	Ser	Lys	Met	Leu	Phe	Ser	Leu	Thr	Ser	Pro	Arg	Leu	Phe	Ser
1				5			10		15						

Ala	Val	Ser	Arg	Lys	Pro	Ser	Ser	Ser	Phe	Ser	Pro	Ser	Pro	Pro	Ser
	20				25				30						

Pro	Ser	Ser	Arg	Thr	Gln	Trp	Thr	Gln	Leu	Ser	Pro	Gly	Lys	Ser	Ile
	35				40				45						

Ser	Leu	Arg	Arg	Val	Phe	Leu	Leu	Pro	Ala	Lys	Ala	Thr	Thr	Glu
	50			55			60							

Gln	Ser	Gly	Pro	Val	Gly	Gly	Asp	Asn	Val	Asp	Ser	Asn	Val	Leu	Pro
	65			70			75			80					

Tyr	Cys	Ser	Ile	Asn	Lys	Ala	Glu	Lys	Lys	Thr	Ile	Gly	Glu	Met	Glu
			85		90					95					

Gln	Glu	Phe	Leu	Gln	Ala	Leu	Gln	Ser	Phe	Tyr	Tyr	Asp	Gly	Lys	Ala
	100			105			110								

Ile	Met	Ser	Asn	Glu	Glu	Phe	Asp	Asn	Leu	Lys	Glu	Glu	Leu	Met	Trp
	115			120			125								

Glu	Gly	Ser	Ser	Val	Val	Met	Leu	Ser	Ser	Asp	Glu	Gln	Arg	Phe	Leu
	130			135			140								

Glu	Ala	Ser	Met	Ala	Tyr	Val	Ser	Gly	Asn	Pro	Ile	Leu	Asn	Asp	Glu
	145			150			155			160					

Glu	Tyr	Asp	Lys	Leu	Lys	Leu	Lys	Ile	Asp	Gly	Ser	Asp	Ile	
	165			170			175							

Val	Ser	Glu	Gly	Pro	Arg	Cys	Ser	Leu	Arg	Ser	Lys	Lys	Val	Tyr	Ser
	180			185			190								

Asp	Leu	Ala	Val	Asp	Tyr	Phe	Lys	Met	Leu	Leu	Asn	Val	Pro	Ala
	195			200			205							

-continued

Thr Val Val Ala Leu Gly Leu Phe Phe Leu Asp Asp Ile Thr Gly
 210 215 220

Phe Glu Ile Thr Tyr Ile Met Glu Leu Pro Glu Pro Tyr Ser Phe Ile
 225 230 235 240

Phe Thr Trp Phe Ala Ala Val Pro Val Ile Val Tyr Leu Ala Leu Ser
 245 250 255

Ile Thr Lys Leu Ile Ile Lys Asp Phe Leu Ile Leu Lys Gly Pro Cys
 260 265 270

Pro Asn Cys Gly Thr Glu Asn Thr Ser Phe Phe Gly Thr Ile Leu Ser
 275 280 285

Ile Ser Ser Gly Gly Lys Thr Asn Thr Val Lys Cys Thr Asn Cys Gly
 290 295 300

Thr Ala Met Val Tyr Asp Ser Gly Ser Arg Leu Ile Thr Leu Pro Glu
 305 310 315 320

Gly Ser Gln Ala

<210> SEQ ID NO 4
<211> LENGTH: 975
<212> TYPE: DNA
<213> ORGANISM: Neisseria gonorrhoeae
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: codon optimized for Arabidopsis thaliana

<400> SEQUENCE: 4

atgggttagca	agatgttgtt	tagtttgaca	agtccctcgac	ttttctccgc	cgttctcgc	60
aaacaccttc	tttctttctc	tccttctct	ccgtcgccgt	cttcgaggac	tcaatggact	120
cagctcagcc	ctggaaaatc	gatttctttg	agaagaagag	tcttcttgg	gcctgctaaa	180
gccacaacag	agcaatcagg	tccagtagga	ggagacaacg	tcgatagcaa	tgtttgccc	240
tattgtaca	tcaacaaggc	tgagaagaaa	acaatttggt	aatggaaaca	agagtttctc	300
caagcgttgc	aatctttcta	ttatgtatggc	aaagcgatca	tgtctaatga	agagtttgat	360
aaccttaaag	aagagttaat	gtgggaagga	agcagtgtt	tgtatgtaag	ttccgatgaa	420
caaagattct	tggaaagcttc	catggcttat	gtttctggaa	atccaatctt	aatgtatgaa	480
gaatatgata	agctcaaact	caaactaaag	attgtatggta	gcgcacattgt	gagcgagggt	540
ccaagatgca	gtctccgtag	taaaaagggt	tatagtgtatc	tgcgtgtaga	ttatttcaaa	600
atgttattgt	tgaatgttcc	agcaaccgtt	gttgctctcg	gactctttt	cttcctggac	660
gacattacag	gttttgagat	cacatacatc	atggagcttc	cagaaccata	cagtttcata	720
ttcacttgg	tgcgtgtgt	gcctgtgatt	gtatatctgg	ctttatcaat	caccaaattg	780
atccatcaagg	acttcttgat	cttgaagggt	ccttgtccga	attgtggAAC	ggaaaacacc	840
tccttctttg	gaacaattct	gtcaatctcc	agcggcggca	aaaccaacac	tgtcaaatgc	900
accaactgcg	gaaccgcgat	ggtgtatgac	tcgggttcta	ggttgtatcac	attgccagaa	960
ggaagccaag	cttaa					975

<210> SEQ ID NO 5
<211> LENGTH: 280
<212> TYPE: PRT
<213> ORGANISM: Neisseria gonorrhoeae
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<222> LOCATION: (1) .. (54)
<223> OTHER INFORMATION: Bacterial carbonic anhydrase (BCA) amino acid sequence with rbcs-1a transit peptide

US 11,001,853 B2

75

76

-continued

<400> SEQUENCE: 5

Met Ala Ser Ser Met Leu Ser Ser Ala Thr Met Val Ala Ser Pro Ala			
1	5	10	15

Gln Ala Thr Met Val Ala Pro Phe Asn Gly Leu Lys Ser Ser Ala Ala		
20	25	30

Phe Pro Ala Thr Arg Lys Ala Asn Asn Asp Ile Thr Ser Ile Thr Ser		
35	40	45

Asn Gly Gly Arg Val Asn His Gly Asn His Thr His Trp Gly Tyr Thr		
50	55	60

Gly His Asp Ser Pro Glu Ser Trp Gly Asn Leu Ser Glu Glu Phe Arg			
65	70	75	80

Leu Cys Ser Thr Gly Lys Asn Gln Ser Pro Val Asn Ile Thr Glu Thr		
85	90	95

Val Ser Gly Lys Leu Pro Ala Ile Lys Val Asn Tyr Lys Pro Ser Met		
100	105	110

Val Asp Val Glu Asn Asn Gly His Thr Ile Gln Val Asn Tyr Pro Glu		
115	120	125

Gly Gly Asn Thr Leu Thr Val Asn Gly Arg Thr Tyr Thr Leu Lys Gln		
130	135	140

Phe His Phe His Val Pro Ser Glu Asn Gln Ile Lys Gly Arg Thr Phe			
145	150	155	160

Pro Met Glu Ala His Phe Val His Leu Asp Glu Asn Lys Gln Pro Leu		
165	170	175

Val Leu Ala Val Leu Tyr Glu Ala Gly Lys Thr Asn Gly Arg Leu Ser		
180	185	190

Ser Ile Trp Asn Val Met Pro Met Thr Ala Gly Lys Val Lys Leu Asn		
195	200	205

Gln Pro Phe Asp Ala Ser Thr Leu Leu Pro Lys Arg Leu Lys Tyr Tyr		
210	215	220

Arg Phe Ala Gly Ser Leu Thr Thr Pro Pro Cys Thr Glu Gly Val Ser			
225	230	235	240

Trp Leu Val Leu Lys Thr Tyr Asp His Ile Asp Gln Ala Gln Ala Glu		
245	250	255

Lys Phe Thr Arg Ala Val Gly Ser Glu Asn Asn Arg Pro Val Gln Pro		
260	265	270

Leu Asn Ala Arg Val Val Ile Glu	
275	280

<210> SEQ ID NO 6

<211> LENGTH: 684

<212> TYPE: DNA

<213> ORGANISM: Neisseria gonorrhoeae

<400> SEQUENCE: 6

aaccacggca atcacaccca ttggggctat accggacacg actctccga aagctgggc	60
---	----

aatctgtcag aagaattccg tttgtgctcc accggcaaaa accaatctcc ggtaaacatt	120
---	-----

accgaaacccg ttcccgcaa actgccgcg atcaaagtca attacaacc gactatggtt	180
---	-----

gacgtggaaa acaacggcca caccattcg gtcaattatc ccgaaggccg caataccctg	240
--	-----

accgtgaacgc gcagaaccta taccctgaaa cagttccact tccacgtgcc gagcgaaaac	300
--	-----

caaatcaaag gcagaacttt cccgatggaa gctcacttcg tccacttaga cgaaaacaaa	360
---	-----

cagcctttag tattagccgt gctgtatgaa gccggcaaaa ccaacgggag actgtctcc	420
--	-----

-continued

atctggAACG	tcatGCCGAT	gaccGCAGGA	aaagtGAAAC	tcaACCAACC	gttcGACGCA	480
tccaccCTAC	tgCCGAAAAG	attGAAATAc	tacAGATTG	ccggTTcgCT	gaccACGCCG	540
ccgtGCACAG	aggGeGTATC	atggTTGGTG	ttGAAAACtT	atGACCACAT	cgACCAAGCG	600
caAGCGGAAA	aattCACCAg	agCCGTCGGT	tcggAAAACA	acAGACCCGT	acageCCTCTG	660
aatgcACGTG	tagTTATTG	ataa				684
<210> SEQ ID NO 7						
<211> LENGTH: 356						
<212> TYPE: DNA						
<213> ORGANISM: Arabidopsis thaliana						
<400> SEQUENCE: 7						
gtttacatt	gatgtctca	ggattcata	aggatagaga	gatctattcg	tatacgTgTC	60
acgtcatgag	tgggtgttC	gccaatccat	gaaacgcacc	tagatatcta	aaacacatAT	120
caattgcgaa	tctgcgaagt	gcgagccatt	aaccacgtaa	gcaaacaAAAC	aatctaaACC	180
ccaaaaaaaaaa	tctatgacta	gccaatAGCA	acctcagaga	ttgatattc	aagataAGAC	240
agtatTTAGA	tttctgtatt	atataTAGCG	aaaatcgcAT	caataccaaa	ccacCCATT	300
cttggcttac	aacaacAAAT	cTTAACGTT	ttactttgtG	ctgcactact	caacCT	356
<210> SEQ ID NO 8						
<211> LENGTH: 162						
<212> TYPE: DNA						
<213> ORGANISM: Arabidopsis thaliana						
<220> FEATURE:						
<221> NAME/KEY: misc_feature						
<223> OTHER INFORMATION: Rbcs-1a transit peptide						
<400> SEQUENCE: 8						
atggcttcct	ctatgtctc	ttccgctact	atgggtgcct	ctccggctca	ggccactatG	60
gtcgctcTT	tcaacggact	taagtccTCC	gctgcctcc	cagccaccAG	aaaggctaAC	120
aacgacatta	cttccatcac	aagcaacggc	ggaagagttA	ac		162
<210> SEQ ID NO 9						
<211> LENGTH: 207						
<212> TYPE: DNA						
<213> ORGANISM: Artificial Sequence						
<220> FEATURE:						
<223> OTHER INFORMATION: NOS terminator from cloning vector.						
<220> FEATURE:						
<221> NAME/KEY: misc_feature						
<223> OTHER INFORMATION: Nos terminator						
<400> SEQUENCE: 9						
atcgTTAAA	cattggCAA	taaagtTTCT	taagattGAA	tcctgttgCC	ggTCTTGCgA	60
tgattatCAT	ataatttCTG	ttGAATTACG	ttaAGCATGT	aataattaAC	atgtAATGCA	120
tgacgttatt	tatGAGATGG	gttttatGA	ttagAGTccc	gcaattatac	atTTAATACG	180
cgatAGAAAAA	caaaatatAG	cgcGCAA				207
<210> SEQ ID NO 10						
<211> LENGTH: 251						
<212> TYPE: DNA						
<213> ORGANISM: Nicotiana sylvestris						
<400> SEQUENCE: 10						
tgtggTCACA	cctcaaACTA	aatcaACCAg	tttgcattt	tttccttCTC	aatgttaATT	60
tgctgacttG	gctaggGTGc	gaatcaaATC	acacgttCTA	attgggCAA	atCCGTATAT	120

-continued

caccttatcc tatatcctt ttctccacca cccatcatct cttctatgca aaaaaatag	180
cttcttcctt ttcattttc acttctctca atccaacttt tctatggcca tggcatccca	240
agcttccctt t	251
<210> SEQ ID NO 11	
<211> LENGTH: 255	
<212> TYPE: DNA	
<213> ORGANISM: Arabidopsis thaliana	
<400> SEQUENCE: 11	
tatacaaagg aaccgatcaa gtggagacta gtaaaccata cacaatca catttcctca	60
caaaagaaaag ataagataag ggtgtcaaca ctttcctta atcatgtggt agtgaacgag	120
ttatcatgaa tcccgaccc tttgtattt agggctttt gctcttacg gttctacta	180
tataaagatg acaaaaccaa tagaaaaaca attaagcaaa agaagaagaa gaagaagtaa	240
tggcttcctc tatgc	255
<210> SEQ ID NO 12	
<211> LENGTH: 3249	
<212> TYPE: DNA	
<213> ORGANISM: Chlamydomonas reinhardtii	
<400> SEQUENCE: 12	
atgcttcctg gtcttggtgt catccttctt gtgcttccta tgcaagtacta cttcggttac	60
aagatgtgc agatcaagct tcagaacgct aagcacgtcg ctcttgcgtc tgctatcatg	120
caggaggtgc ttccctgctat caagcttgc aagtactacg cttggagca gttctttag	180
aaccagatct ctaaggatccg tcgtgaggag atccgtctca acttctggaa ctgcgtgtat	240
aaggcatca acgtggctt cgtgttctgc gtgccgctt tgaccgttt cgtcatctc	300
accacctacg agtccacgcg tgctcgttgc gtgtcttgc tgcgttccac acccttct	360
ctttcaaca ttcttcgtt ccctcttgc tgcttccta aggctttcg tgctgtgtct	420
gaggctaacg cttcttcca gcgtcttgcg gttacccctt ttgaggaggt gcctctgg	480
actgctgtcg tcaagacccc taagaacgct cttcttgcgtt ctgtcatcga gaacgggttg	540
ttccaccacc cttctaaccct taactggac cttcacgtcg ctaagttcga ggtcaaggct	600
ggtcaggatcg ttgctgtggt gggtcgtatc gctgctggta agtcttctt tgcgtcaggct	660
atcctcggta acatggtaaa ggagcacggt tcttcaacg tgggtggcg tatctctac	720
gtgccgcaga acccttggct tcagaacctt tctcttcgtt acaacgtgtt ttttgtgag	780
cagttcgatg agaacaagta cacggacgtc atcgagtctt ggcgttccac ctttgaccc	840
cagatctttt ctaacggta ccagtctaa gctggatcc gttgttcaaa cttctctgg	900
ggtcagcgctc agcgtgtgaa ctttcgttgc tgccgttac ctgcgtgttgc ctttgttgc	960
ctcgacaacg ctcttctgc tttggaccac cacaccgttc accacatctt cgacaagtgc	1020
atcaagggttc tttctctga caaggctgtt gttgttgc cccaccatcg cgagttcatg	1080
cctcggttgc acaacgtggc tatcatggac gagggttgc gctttactt cggtaagtgg	1140
aacgaggagg ctcagcacct tctcggttgc cttcttccta tcacccaccc tcttcacgt	1200
gctgggttgc aggaggctcc tctcggttgc aagaagaagg ctgaggacaa gggtggcct	1260
cagaagtctc agtctttca gcttaccctt gctccttaccc tctatcgtaa gcctaccgag	1320
aaggcttaagg acgtccagaa gcttactgtt taccaggctg ctctcatcta cacctggat	1380

US 11,001,853 B2

81

82

-continued

ggtaacctt	tccttgttgg	tgtgtgcttc	ttcttcttcc	ttgctgctca	gtgctctcg	1440
cagatctctg	atttctgggt	gcgttgggtgg	gtgaacgacg	agtacaagaa	gttccctgtg	1500
aagggtgagc	aggactctgc	tgctaccacc	ttctactgcc	tcatctaccc	tcttcttgc	1560
ggtctttct	acatcttcat	gatcttccgt	ggtgctactt	tcctttgggt	ggtgetcaag	1620
tcttctgaga	ccatccgtag	gaaggcttcc	cacaacgtcc	tcaacgctcc	tatgggttc	1680
ttccttgc	cgccgggtgg	tgaccccttcc	ctcaactca	ccaaggacca	ggacattatg	1740
gatgagaacc	tccctgtatgc	tgttcaacttc	atgggtatct	acggcttat	tcttcttgc	1800
accaccatca	ccgtgtctgt	caccatcaac	ttcttcgtct	ctttcacccg	tgctcttata	1860
atcatgaccc	tcatcatgtct	ctctatctac	cttcctgtgt	ctactgtct	taagaaggct	1920
cgtgctgtgt	ctgggtgtat	gcttgcgtt	cttgcgtgt	agggttctga	gggtcttgg	1980
gtgggtcagg	ctttcaacaa	gcaggagtag	ttcattgagg	aggctgtcg	tcgtaccaac	2040
atcaccaact	ctgctgtctt	caacgctgag	gctcttaacc	tttgcgttgc	tttcttgc	2100
gacttcatcg	tggttgcct	tgtgggtgt	gtgtctgtct	tgcgtgtgg	tatggctaa	2160
gaccccttgg	gtgctaccgt	cggttgcgt	ttctctaaca	tcatcagat	gcttgcgttgc	2220
tacaccttgg	ttgttgcgtt	catctctgag	tctatctctc	tcttcaactc	tgtcgagggt	2280
atggcttacc	tcgctgacta	cgtgcctc	gatgggtgt	tctatgacca	gcgtcagaag	2340
gacgggtgtcg	ctaagcaaat	cgtccttct	gacggtaaca	tcgtgcctc	tgcttctaa	2400
gtccagggtcg	ttgttgcgt	cgtgcgttgc	gtcgatccgg	taacatccgt		2460
ttcgaggacg	tgtggatgca	gtaccgttct	gacgcttctt	gggctttaa	gggtgtcacc	2520
ttcaagatca	acgacggta	gaagggtcggt	gctgtgggtc	gtaccgggtc	tggtaagtct	2580
accacgcttc	ttgctcttta	ccgttatgttc	gagcttggta	agggtcgat	ccttgcgtac	2640
ggtgtggaca	tgcgtaccct	ttctctcaag	cgtcttcgt	ccggcttctt	tatcattct	2700
caggagcctg	tcatgttac	cggttaccgt	cgttctaaacc	ttgaccctt	cggtgagttc	2760
aaggacgatg	ctatttttgc	ggaggtgtt	aagaaggctcg	gtctcgagga	ccaggctcag	2820
cacgctgggt	gtcttgcgtt	tcaggctcgat	ggtaccgggt	gttacggctt	gtcttgcgtt	2880
cagatgcacg	ttgttgcgtt	tgcgtgtgt	gtcttcgtt	ctgtgcctat	ccttgcctt	2940
gacgaggctt	ccgctgttat	ggacccgcac	actgaggctt	tcgtgcacca	gaccatcaag	3000
aagggttgc	acgacggta	caccatcacc	attgttaccc	gtcttgacac	catcatcgag	3060
tctgacaaga	tcatcgat	ggaggcagggt	tctttatgg	agtacgagtc	tcttctaa	3120
cttctcgcta	accgtgactc	tatgttctt	aagcttgcgt	acaagaccgg	tcttgcgtct	3180
getgtgtctc	ttcgtaagat	ggctgaggac	ttctggtcta	tcgttgcgt	tcagggtcg	3240
aaccagtaa						3249

<210> SEQ ID NO 13

<211> LENGTH: 35

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 13

Met	Ala	Ala	Ser	Thr	Met	Ala	Leu	Ser	Ser	Pro	Ala	Phe	Ala	Gly	Lys
1					5			10			15				

Ala	Val	Asn	Leu	Ser	Pro	Ala	Ala	Ser	Glu	Val	Leu	Gly	Ser	Gly	Arg
						20		25			30				

Val Thr Met

-continued

35

<210> SEQ_ID NO 14
<211> LENGTH: 44
<212> TYPE: PRT
<213> ORGANISM: Arabidopsis thaliana
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: PGR5 transit peptide

<400> SEQUENCE: 14

Met	Ala	Ala	Ala	Ser	Ile	Ser	Ala	Ile	Gly	Cys	Asn	Gln	Thr	Leu	Ile	
1									5					10		15

Gly	Thr	Ser	Phe	Tyr	Gly	Gly	Trp	Gly	Ser	Ser	Ile	Ser	Gly	Glu	Asp	
									20					25		30

Tyr	Gln	Thr	Met	Leu	Ser	Lys	Thr	Val	Ala	Pro	Pro					
			35						40							

<210> SEQ_ID NO 15
<211> LENGTH: 45
<212> TYPE: PRT
<213> ORGANISM: Arabidopsis thaliana
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: psaD transit peptide

<400> SEQUENCE: 15

Met	Ala	Thr	Gln	Ala	Ala	Gly	Ile	Phe	Asn	Ser	Ala	Ile	Thr	Thr	Ala	
1									5					10		15

Ala	Thr	Ser	Gly	Val	Lys	Lys	Leu	His	Phe	Phe	Ser	Thr	Thr	His	Arg	
				20					25					30		

Pro	Lys	Ser	Leu	Ser	Phe	Thr	Lys	Thr	Ala	Ile	Arg	Ala				
			35						40					45		

<210> SEQ_ID NO 16
<211> LENGTH: 1011
<212> TYPE: DNA
<213> ORGANISM: Chlamydomonas reinhardtii

<400> SEQUENCE: 16

```

atgcagacca ctatgactcg cccttgcctt gcccagcccc tgctgcgatc tcgtgtgctc      60
cggttcgccta tgcgggtgg tgcagcgacg gctccctaccg cggtgacgac agtcgtgacc      120
tcgaatggaa atggcaacgg tcatttccaa gctgctacta cgcccggtgcc ccctactccc      180
gtccccgtcg ctgtttccgc gcctgtgcgc gctgtgtcg tgctgactcc tcctcaagtg      240
tatgagaacg ccattaatgt tggcgctac aaggccgggc taacgcctct ggcaacgttt      300
gtccaggggca tccaaggccgg tgcctacatt gcgttccggc cttccctcgc catctccgtg      360
ggaggcaaca tccccggcgt cgccggccgc aaccccgccc tggccaagct gctatttgct      420
ctgggtgttcc ccgtgggtct gtccatggtg accaactcgcc gggccggagct gttcacgggc      480
aacaccatga tgctcacatg cgcgctcatac gagaagaagg ccacttgggg gcagttctg      540
aagaactgga gcgtgtccata cttcggcaac ttctggggct ccatcgccat ggtcgccgccc      600
gtgggtggcca ccggctgcct gaccaccaac accctgcctg tgcagatggc caccctcaag      660
gcacaacctgg gtttacccga ggtgtgtcg cgctccatcc tggcaactg gctgggtgtc      720
tgcggcgtgt ggtccgcctc cgccggccacc tgcgtggccg ggcgcattcc ggcgcgtgtgg      780
ccctgcata cccgcattcggt ggccatcgcc ctggagcact ccgtcgccaa catgttcgtg      840
attccctctgg gcatgatgct gggcgctgag gtcacgtggg gccagtttt tttcaacaac      900

```

-continued

ctgatccccg tcaccctggg caacaccatt gctggcgttc tcatgatggc catcgctac 960
 tccatctcggtcggtccct cggcaagttcc gccaaggcccg ccaccgcgtaa 1011

<210> SEQ ID NO 17
 <211> LENGTH: 892
 <212> TYPE: DNA
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 17

atgatatacct tttagtgcgtgt gactacagtc agccgtgtttt ctacgggtgca atcggcccg 60
 gtggctccat tcggggccct caaatccatg actggattcc cagttaaagaa ggtcaacact 120
 gacattactt ccattacaag caatggtgga agagtaaagt gcatgcaggt ggagctctct 180
 catcattggg gttatggtaa acacaatggt cctgaacact ggcataaaga ctttccaatt 240
 gaaaaagggtg aacgtcaatc acctgttgcattt attgacactc atacagctaa atatgaccct 300
 tctttaaaac cattatctgt ttcatatgtt caagcaactt ctttacgtat tttaaacaat 360
 ggtcatgctt ttaatgtaga atttgatgac tctcaagata aagcagtatt aaaagggtgt 420
 ccatttagatg gtacttaccg tttaattcaa ttcaacttcc actgggggttc attagatgg 480
 caagggttcag aacatactgt agataaaaaaa aaatatgtc cagaattaca cttagttcac 540
 tggAACACAA aatatggtaa ttttggtaaa gctgtacaac aacctgtatgg ttttagcttt 600
 ttaggtatTTT tttaaaaagt tggtagtgct aaaccagggtc ttcaaaaagt tgtagtgc 660
 ttacttccctg aatctttaga ttactggaca tatccaggtt cattaacaac accttctt 720
 tttagatgtg taacatggat tggatggaa gaaccaatta gtgttagtgc tgaacaagta 780
 tttaaaattcc gtaaaacttaa tttcaatggt gaaggtgaac cagaagaatt aa 840
 892

<210> SEQ ID NO 18
 <211> LENGTH: 336
 <212> TYPE: PRT
 <213> ORGANISM: Chlamydomonas reinhardtii

<400> SEQUENCE: 18

Met Gln Thr Thr Met Thr Arg Pro Cys Leu Ala Gln Pro Val Leu Arg
 1 5 10 15

Ser Arg Val Leu Arg Ser Pro Met Arg Val Val Ala Ala Ser Ala Pro
 20 25 30

Thr Ala Val Thr Thr Val Val Thr Ser Asn Gly Asn Gly Asn Gly His
 35 40 45

Phe Gln Ala Ala Thr Thr Pro Val Pro Pro Thr Pro Ala Pro Val Ala
 50 55 60

Val Ser Ala Pro Val Arg Ala Val Ser Val Leu Thr Pro Pro Gln Val
 65 70 75 80

Tyr Glu Asn Ala Ile Asn Val Gly Ala Tyr Lys Ala Gly Leu Thr Pro
 85 90 95

Leu Ala Thr Phe Val Gln Gly Ile Gln Ala Gly Ala Tyr Ile Ala Phe
 100 105 110

Gly Ala Phe Leu Ala Ile Ser Val Gly Gly Asn Ile Pro Gly Val Ala
 115 120 125

Ala Ala Asn Pro Gly Leu Ala Lys Leu Leu Phe Ala Leu Val Phe Pro
 130 135 140

Val Gly Leu Ser Met Val Thr Asn Cys Gly Ala Glu Leu Phe Thr Gly

US 11,001,853 B2

87**88**

-continued

145	150	155	160
Asn Thr Met Met Leu Thr Cys Ala Leu Ile Glu Lys Lys Ala Thr Trp			
165	170	175	
Gly Gln Leu Leu Lys Asn Trp Ser Val Ser Tyr Phe Gly Asn Phe Val			
180	185	190	
Gly Ser Ile Ala Met Val Ala Ala Val Val Ala Thr Gly Cys Leu Thr			
195	200	205	
Thr Asn Thr Leu Pro Val Gln Met Ala Thr Leu Lys Ala Asn Leu Gly			
210	215	220	
Phe Thr Glu Val Leu Ser Arg Ser Ile Leu Cys Asn Trp Leu Val Cys			
225	230	235	240
Cys Ala Val Trp Ser Ala Ser Ala Ala Thr Ser Leu Pro Gly Arg Ile			
245	250	255	
Leu Ala Leu Trp Pro Cys Ile Thr Ala Phe Val Ala Ile Gly Leu Glu			
260	265	270	
His Ser Val Ala Asn Met Phe Val Ile Pro Leu Gly Met Met Leu Gly			
275	280	285	
Ala Glu Val Thr Trp Ser Gln Phe Phe Asn Asn Leu Ile Pro Val			
290	295	300	
Thr Leu Gly Asn Thr Ile Ala Gly Val Leu Met Met Ala Ile Ala Tyr			
305	310	315	320
Ser Ile Ser Phe Gly Ser Leu Gly Lys Ser Ala Lys Pro Ala Thr Ala			
325	330	335	

<210> SEQ ID NO 19

<211> LENGTH: 260

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 19

Met	Ser	His	His	Trp	Gly	Tyr	Gly	Lys	His	Asn	Gly	Pro	Glu	His	Trp
1				5			10			15					
His Lys Asp Phe Pro Ile Ala Lys Gly Glu Arg Gln Ser Pro Val Asp															
				20			25			30					
Ile Asp Thr His Thr Ala Lys Tyr Asp Pro Ser Leu Lys Pro Leu Ser															
				35			40			45					
Val Ser Tyr Asp Gln Ala Thr Ser Leu Arg Ile Leu Asn Asn Gly His															
				50			55			60					
Ala Phe Asn Val Glu Phe Asp Asp Ser Gln Asp Lys Ala Val Leu Lys															
				65			70			75			80		
Gly Gly Pro Leu Asp Gly Thr Tyr Arg Leu Ile Gln Phe His Phe His															
				85			90			95					
Trp Gly Ser Leu Asp Gly Gln Gly Ser Glu His Thr Val Asp Lys Lys															
				100			105			110					
Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly															
				115			120			125					
Asp Phe Gly Lys Ala Val Gln Gln Pro Asp Gly Leu Ala Val Leu Gly															
				130			135			140					
Ile Phe Leu Lys Val Gly Ser Ala Lys Pro Gly Leu Gln Lys Val Val															
				145			150			155			160		
Asp Val Leu Asp Ser Ile Lys Thr Lys Gly Lys Ser Ala Asp Phe Thr															
				165			170			175					
Asn Phe Asp Pro Arg Gly Leu Leu Pro Glu Ser Leu Asp Tyr Trp Thr															
				180			185			190					

US 11,001,853 B2

89**90**

-continued

Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp
 195 200 205

Ile Val Leu Lys Glu Pro Ile Ser Val Ser Ser Glu Gln Val Leu Lys
 210 215 220

Phe Arg Lys Leu Asn Phe Asn Gly Glu Gly Pro Glu Glu Leu Met
 225 230 235 240

Val Asp Asn Trp Arg Pro Ala Gln Pro Leu Lys Asn Arg Gln Ile Lys
 245 250 255

Ala Ser Phe Lys
 260

<210> SEQ ID NO 20

<211> LENGTH: 159

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 20

Met Val Met Leu Ser Thr Trp Ser Leu Met Thr Leu Arg Thr Lys Gln
 1 5 10 15

Leu His Leu Val His Trp Asn Thr Lys Tyr Gly Asp Phe Gly Lys Ala
 20 25 30

Val Gln Gln Pro Asp Gly Leu Ala Val Leu Gly Ile Phe Leu Lys Val
 35 40 45

Gly Ser Ala Lys Pro Gly Leu Gln Lys Val Val Asp Val Leu Asp Ser
 50 55 60

Ile Lys Thr Lys Gly Lys Ser Ala Asp Phe Thr Asn Phe Asp Pro Arg
 65 70 75 80

Gly Leu Leu Pro Glu Ser Leu Asp Tyr Trp Thr Tyr Pro Gly Ser Leu
 85 90 95

Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp Ile Val Leu Lys Glu
 100 105 110

Pro Ile Ser Val Ser Ser Glu Gln Val Leu Lys Phe Arg Lys Leu Asn
 115 120 125

Phe Asn Gly Glu Gly Glu Pro Glu Glu Leu Met Val Asp Asn Trp Arg
 130 135 140

Pro Ala Gln Pro Leu Lys Asn Arg Gln Ile Lys Ala Ser Phe Lys
 145 150 155

<210> SEQ ID NO 21

<211> LENGTH: 252

<212> TYPE: PRT

<213> ORGANISM: Neisseria gonorrhoeae

<400> SEQUENCE: 21

Met Pro Arg Phe Pro Arg Thr Leu Pro Arg Leu Thr Ala Val Leu Leu
 1 5 10 15

Leu Ala Cys Thr Ala Phe Ser Ala Ala Ala His Gly Asn His Thr His
 20 25 30

Trp Gly Tyr Thr Gly His Asp Ser Pro Glu Ser Trp Gly Asn Leu Ser
 35 40 45

Glu Glu Phe Arg Leu Cys Ser Thr Gly Lys Asn Gln Ser Pro Val Asn
 50 55 60

Ile Thr Glu Thr Val Ser Gly Lys Leu Pro Ala Ile Lys Val Asn Tyr
 65 70 75 80

Lys Pro Ser Met Val Asp Val Glu Asn Asn Gly His Thr Ile Gln Val
 85 90 95

-continued

Asn Tyr Pro Glu Gly Gly Asn Thr Leu Thr Val Asn Gly Arg Thr Tyr
 100 105 110
 Thr Leu Lys Gln Phe His Phe His Val Pro Ser Glu Asn Gln Ile Lys
 115 120 125
 Gly Arg Thr Phe Pro Met Glu Ala His Phe Val His Leu Asp Glu Asn
 130 135 140
 Lys Gln Pro Leu Val Leu Ala Val Leu Tyr Glu Ala Gly Lys Thr Asn
 145 150 155 160
 Gly Arg Leu Ser Ser Ile Trp Asn Val Met Pro Met Thr Ala Gly Lys
 165 170 175
 Val Lys Leu Asn Gln Pro Phe Asp Ala Ser Thr Leu Leu Pro Lys Arg
 180 185 190
 Leu Lys Tyr Tyr Arg Phe Ala Gly Ser Leu Thr Thr Pro Pro Cys Thr
 195 200 205
 Glu Gly Val Ser Trp Leu Val Leu Lys Thr Tyr Asp His Ile Asp Gln
 210 215 220
 Ala Gln Ala Glu Lys Phe Thr Arg Ala Val Gly Ser Glu Asn Asn Arg
 225 230 235 240
 Pro Val Gln Pro Leu Asn Ala Arg Val Val Ile Glu
 245 250

<210> SEQ_ID NO 22
 <211> LENGTH: 217
 <212> TYPE: PRT
 <213> ORGANISM: Arabidopsis thaliana
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: DNAJ transit peptide

 <400> SEQUENCE: 22

 Met Ala Ser Leu Ser Thr Ile Thr Gln Pro Ser Leu Val His Ile Pro
 1 5 10 15
 Gly Glu Ser Val Leu His His Val Pro Ser Thr Cys Ser Phe Pro Trp
 20 25 30
 Lys Pro Thr Ile Asn Thr Lys Arg Ile Ile Cys Ser Pro Ala Arg Asn
 35 40 45
 Ser Ser Glu Val Ser Ala Glu Ala Glu Thr Glu Gly Gly Ser Ser Thr
 50 55 60
 Ala Val Asp Glu Ala Pro Lys Glu Ser Pro Ser Leu Ile Ser Ala Leu
 65 70 75 80
 Asn Val Glu Arg Ala Leu Arg Gly Leu Pro Ile Thr Asp Val Asp His
 85 90 95
 Tyr Gly Arg Leu Gly Ile Phe Arg Asn Cys Ser Tyr Asp Gln Val Thr
 100 105 110
 Ile Gly Tyr Lys Glu Arg Val Lys Glu Leu Lys Glu Gln Gly Leu Asp
 115 120 125
 Glu Glu Gln Leu Lys Thr Lys Met Asp Leu Ile Lys Ser Tyr Thr Ile
 130 135 140
 Leu Ser Thr Val Glu Glu Arg Arg Met Tyr Asp Trp Ser Leu Ala Arg
 145 150 155 160
 Ser Glu Lys Ala Glu Arg Tyr Val Trp Pro Phe Glu Val Asp Ile Met
 165 170 175
 Glu Pro Ser Arg Glu Glu Pro Pro Pro Gln Glu Pro Glu Asp Val Gly
 180 185 190
 Pro Thr Arg Ile Leu Gly Tyr Phe Ile Gly Ala Trp Leu Val Leu Gly
 195 200 205

-continued

Val Ala Leu Ser Val Ala Phe Asn Arg
210 215

<210> SEQ ID NO 23
<211> LENGTH: 67
<212> TYPE: PRT
<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 23

Met Asp Lys Ala Leu Thr Gly Ile Ser Ala Ala Ala Leu Thr Ala Ser
1 5 10 15

Met Val Ile Pro Glu Ile Ala Glu Ala Ala Gly Ser Gly Ile Ser Pro
20 25 30

Ser Leu Lys Asn Phe Leu Leu Ser Ile Ala Ser Gly Gly Leu Val Leu
35 40 45

Thr Val Ile Ile Gly Val Val Val Gly Val Ser Asn Phe Asp Pro Val
50 55 60

Lys Arg Thr
65

<210> SEQ ID NO 24
<211> LENGTH: 260
<212> TYPE: PRT
<213> ORGANISM: Macaca fascicularis

<400> SEQUENCE: 24

Met Ser His His Trp Gly Tyr Gly Lys His Asn Gly Pro Glu His Trp
1 5 10 15

His Lys Asp Phe Pro Ile Ala Lys Gly Gln Arg Gln Ser Pro Val Asp
20 25 30

Ile Asp Thr His Thr Ala Lys Tyr Asp Pro Ser Leu Lys Pro Leu Ser
35 40 45

Val Ser Tyr Asp Gln Ala Thr Ser Leu Arg Ile Leu Asn Asn Gly His
50 55 60

Ser Phe Asn Val Glu Phe Asp Asp Ser Gln Asp Lys Ala Val Ile Lys
65 70 75 80

Gly Gly Pro Leu Asp Gly Thr Tyr Arg Leu Ile Gln Phe His Phe His
85 90 95

Trp Gly Ser Leu Asp Gly Gln Gly Ser Glu His Thr Val Asp Lys Lys
100 105 110

Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly
115 120 125

Asp Phe Gly Lys Ala Val Gln Gln Pro Asp Gly Leu Ala Val Leu Gly
130 135 140

Ile Phe Leu Lys Val Gly Ser Ala Lys Pro Gly Leu Gln Lys Val Val
145 150 155 160

Asp Val Leu Asp Ser Ile Lys Thr Lys Gly Lys Ser Ala Asp Phe Thr
165 170 175

Asn Phe Asp Pro Arg Gly Leu Leu Pro Glu Ser Leu Asp Tyr Trp Thr
180 185 190

Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp
195 200 205

Ile Val Leu Lys Glu Pro Ile Ser Val Ser Ser Glu Gln Met Ser Lys
210 215 220

Phe Arg Lys Leu Asn Phe Asn Gly Glu Gly Pro Glu Glu Leu Met
225 230 235 240

US 11,001,853 B2

95**96**

-continued

Val Asp Asn Trp Arg Pro Ala Gln Pro Leu Lys Asn Arg Gln Ile Lys
245 250 255

Ala Ser Phe Lys
260

<210> SEQ ID NO 25

<211> LENGTH: 260

<212> TYPE: PRT

<213> ORGANISM: Pan troglodytes

<400> SEQUENCE: 25

Met Ser His His Trp Gly Tyr Gly Lys His Asn Gly Pro Glu His Trp
1 5 10 15

His Lys Asp Phe Pro Ile Ala Lys Gly Glu Arg Gln Ser Pro Val Asp
20 25 30

Ile Asp Thr His Thr Ala Lys Tyr Asp Pro Ser Leu Lys Pro Leu Ser
35 40 45

Val Ser Tyr Gly Gln Ala Thr Ser Leu Arg Ile Leu Asn Asn Gly His
50 55 60

Ala Phe Asn Val Glu Phe Asp Asp Ser Gln Asp Lys Ala Val Leu Lys
65 70 75 80

Gly Gly Pro Leu Asp Gly Thr Tyr Arg Leu Ile Gln Phe His Phe His
85 90 95

Trp Gly Ser Leu Asp Gly Gln Gly Ser Glu His Thr Val Asp Lys Lys
100 105 110

Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly
115 120 125

Asp Phe Gly Lys Ala Val Gln Gln Pro Asp Gly Leu Ala Val Leu Gly
130 135 140

Ile Phe Leu Lys Val Gly Ser Ala Lys Pro Gly Leu Gln Lys Val Val
145 150 155 160

Asp Val Leu Asp Ser Ile Lys Thr Lys Gly Lys Ser Ala Asp Phe Thr
165 170 175

Asn Phe Asp Pro His Gly Leu Leu Pro Glu Ser Leu Asp Tyr Trp Thr
180 185 190

Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp
195 200 205

Ile Val Leu Lys Glu Pro Ile Ser Val Ser Ser Glu Gln Met Leu Lys
210 215 220

Phe Arg Lys Leu Asn Phe Asn Gly Glu Gly Glu Pro Glu Glu Leu Met
225 230 235 240

Val Asp Asn Trp Arg Pro Ala Gln Pro Leu Lys Asn Arg Gln Ile Lys
245 250 255

Ala Ser Phe Lys
260

<210> SEQ ID NO 26

<211> LENGTH: 260

<212> TYPE: PRT

<213> ORGANISM: Pongo abelii

<400> SEQUENCE: 26

Met Ser His His Trp Gly Tyr Gly Lys His Asn Gly Pro Glu His Trp
1 5 10 15

His Lys Asp Phe Pro Ile Ala Lys Gly Glu Arg Gln Ser Pro Val Asp
20 25 30

US 11,001,853 B2

97

-continued

Ile Asp Thr His Thr Ala Lys Tyr Asp Pro Ser Leu Lys Pro Leu Ser
 35 40 45

Val Cys Tyr Asp Gln Ala Thr Ser Leu Arg Ile Leu Asn Asn Gly His
 50 55 60

Ser Phe Asn Val Glu Phe Asp Asp Ser Gln Asp Lys Ala Val Leu Lys
 65 70 75 80

Gly Gly Pro Leu Asp Gly Thr Tyr Arg Leu Ile Gln Phe His Phe His
 85 90 95

Trp Gly Ser Leu Asp Gly Gln Gly Ser Glu His Thr Val Asp Lys Lys
 100 105 110

Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly
 115 120 125

Asp Phe Gly Lys Ala Val Gln Gln Pro Asp Gly Leu Ala Val Leu Gly
 130 135 140

Ile Phe Leu Lys Val Gly Ser Ala Lys Pro Gly Leu Gln Lys Val Val
 145 150 155 160

Asp Val Leu Asp Ser Ile Lys Thr Lys Gly Lys Cys Ala Asp Phe Thr
 165 170 175

Asn Phe Asp Pro Arg Gly Leu Leu Pro Ala Ser Leu Asp Tyr Trp Thr
 180 185 190

Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp
 195 200 205

Ile Val Leu Lys Glu Pro Ile Ser Val Ser Ser Glu Gln Met Leu Lys
 210 215 220

Phe Arg Lys Leu Asn Phe Asn Gly Glu Gly Glu Pro Glu Glu Leu Met
 225 230 235 240

Val Asp Asn Trp Arg Pro Ala Gln Pro Leu Lys Lys Arg Gln Ile Lys
 245 250 255

Ala Ser Phe Lys
 260

<210> SEQ_ID NO 27

<211> LENGTH: 260

<212> TYPE: PRT

<213> ORGANISM: Pongo abelii

<400> SEQUENCE: 27

Met Ser His His Trp Gly Tyr Gly Lys His Asn Gly Pro Glu His Trp
 1 5 10 15

His Lys Asp Phe Pro Ile Ala Lys Gly Glu Arg Gln Ser Pro Val Asp
 20 25 30

Ile Asp Thr His Thr Ala Lys Tyr Asp Pro Ser Leu Lys Pro Leu Ser
 35 40 45

Val Cys Tyr Asp Gln Ala Thr Ser Leu Arg Ile Leu Asn Asn Gly His
 50 55 60

Ser Phe Asn Val Glu Phe Asp Asp Ser Gln Asp Lys Ala Val Leu Lys
 65 70 75 80

Gly Gly Pro Leu Asp Gly Thr Tyr Arg Leu Ile Gln Phe His Phe His
 85 90 95

Trp Gly Ser Leu Asp Gly Gln Gly Ser Glu His Thr Val Asp Lys Lys
 100 105 110

Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly
 115 120 125

Asp Phe Gly Lys Ala Val Gln Gln Pro Asp Gly Leu Ala Val Leu Gly

98

US 11,001,853 B2

99**100**

-continued

130 135 140

Ile Phe Leu Lys Val Gly Ser Ala Lys Pro Gly Leu Gln Lys Val Val
 145 150 155 160

Asp Val Leu Asp Ser Ile Lys Thr Lys Gly Lys Cys Ala Asp Phe Thr
 165 170 175

Asn Phe Asp Pro Arg Gly Leu Leu Pro Ala Ser Leu Asp Tyr Trp Thr
 180 185 190

Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp
 195 200 205

Ile Val Leu Lys Glu Pro Ile Ser Val Ser Ser Glu Gln Met Leu Lys
 210 215 220

Phe Arg Lys Leu Asn Phe Asn Gly Glu Gly Glu Pro Glu Glu Leu Met
 225 230 235 240

Val Asp Asn Trp Arg Pro Ala Gln Pro Leu Lys Lys Arg Gln Ile Lys
 245 250 255

Ala Ser Phe Lys
 260

<210> SEQ ID NO: 28

<211> LENGTH: 260

<212> TYPE: PRT

<213> ORGANISM: Callithrix jacchus

<400> SEQUENCE: 28

Met Ser His His Trp Gly Tyr Gly Lys His Asn Gly Pro Glu His Trp
 1 5 10 15

His Lys Asp Phe Pro Ile Ala Lys Gly Glu Arg Gln Ser Pro Val Asp
 20 25 30

Ile Asp Thr His Thr Ala Lys Tyr Asp Pro Ser Leu Lys Pro Leu Ser
 35 40 45

Val Ser Tyr Asp Gln Ala Thr Ser Trp Arg Ile Leu Asn Asn Gly His
 50 55 60

Ser Phe Asn Val Glu Phe Asp Asp Ser Gln Asp Lys Ala Val Leu Lys
 65 70 75 80

Gly Gly Pro Leu Asp Gly Thr Tyr Arg Leu Ile Gln Phe His Phe His
 85 90 95

Trp Gly Ser Thr Asp Gly Gln Gly Ser Glu His Thr Val Asp Lys Lys
 100 105 110

Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly
 115 120 125

Asp Phe Gly Lys Ala Ala Gln Gln Pro Asp Gly Leu Ala Val Leu Gly
 130 135 140

Ile Phe Leu Lys Val Gly Ser Ala Lys Pro Gly Leu Gln Lys Val Val
 145 150 155 160

Asp Val Leu Asp Ser Ile Lys Thr Lys Gly Lys Ser Ala Asp Phe Thr
 165 170 175

Asn Phe Asp Pro Arg Gly Leu Leu Pro Glu Ser Leu Asp Tyr Trp Thr
 180 185 190

Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Ser Val Thr Trp
 195 200 205

Ile Val Leu Lys Glu Pro Ile Ser Val Ser Ser Glu Gln Ile Leu Lys
 210 215 220

Phe Arg Lys Leu Asn Phe Ser Gly Glu Gly Glu Pro Glu Glu Leu Met
 225 230 235 240

US 11,001,853 B2

101**102**

-continued

Val	Asp	Asn	Trp	Arg	Pro	Ala	Gln	Pro	Leu	Lys	Asn	Arg	Gln	Ile	Lys
245									250						255

Ala	Ser	Phe	Lys
260			

<210> SEQ ID NO 29

<211> LENGTH: 260

<212> TYPE: PRT

<213> ORGANISM: Lemur catta

<400> SEQUENCE: 29

Met	Ser	His	His	Trp	Gly	Tyr	Gly	Lys	His	Asn	Gly	Pro	Glu	His	Trp
1															
				5				10						15	

His	Lys	Asp	Phe	Pro	Ile	Ala	Lys	Gly	Glu	Arg	Gln	Ser	Pro	Val	Asp
20								25						30	

Ile	Asn	Thr	Gly	Ala	Ala	Lys	His	Asp	Pro	Ser	Leu	Lys	Pro	Leu	Ser
35								40				45			

Val	Tyr	Tyr	Glu	Gln	Ala	Thr	Ser	Arg	Arg	Ile	Leu	Asn	Asn	Gly	His
50								55				60			

Ser	Phe	Asn	Val	Glu	Phe	Asp	Asp	Ser	Gln	Asp	Lys	Ala	Val	Leu	Lys
65								70				75		80	

Gly	Gly	Pro	Leu	Asp	Gly	Thr	Tyr	Arg	Leu	Ile	Gln	Phe	His	Phe	His
85								90				95			

Trp	Gly	Ser	Leu	Asp	Gly	Gln	Gly	Ser	Glu	His	Thr	Val	Asp	Lys	Lys
100								105				110			

Lys	Tyr	Ala	Ala	Glu	Leu	His	Leu	Val	His	Trp	Asn	Thr	Lys	Tyr	Gly
115								120				125			

Asp	Phe	Gly	Lys	Ala	Val	Gln	Gln	Pro	Asp	Gly	Leu	Ala	Val	Leu	Gly
130								135				140			

Ile	Phe	Leu	Lys	Val	Gly	Ser	Ala	Lys	Pro	Gly	Leu	Gln	Lys	Val	Val
145								150				155		160	

Asp	Val	Leu	Asp	Ser	Ile	Lys	Thr	Lys	Gly	Lys	Ser	Ala	Asp	Phe	Thr
165								170				175			

Asn	Phe	Asp	Pro	Arg	Gly	Leu	Leu	Pro	Glu	Ser	Leu	Asp	Tyr	Trp	Thr
180								185				190			

Tyr	Leu	Gly	Ser	Leu	Thr	Thr	Pro	Pro	Leu	Leu	Glu	Cys	Val	Thr	Trp
195								200				205			

Ile	Val	Leu	Lys	Glu	Pro	Ile	Ser	Val	Ser	Ser	Glu	Gln	Met	Met	Lys
210								215				220			

Phe	Arg	Lys	Leu	Ser	Phe	Ser	Gly	Glu	Gly	Glu	Pro	Glu	Glu	Leu	Met
225								230				235		240	

Val	Asp	Asn	Trp	Arg	Pro	Ala	Gln	Pro	Leu	Lys	Asn	Arg	Gln	Ile	Lys
245								250				255			

Ala	Ser	Phe	Lys
260			

<210> SEQ ID NO 30

<211> LENGTH: 260

<212> TYPE: PRT

<213> ORGANISM: Ailuropoda melanoleuca

<400> SEQUENCE: 30

Met	Ala	His	His	Trp	Gly	Tyr	Gly	Lys	His	Asn	Gly	Pro	Glu	His	Trp
1															
				5				10				15			

Tyr	Lys	Asp	Phe	Pro	Ile	Ala	Lys	Gly	Gln	Arg	Gln	Ser	Pro	Val	Asp
20								25				30			

US 11,001,853 B2

103**104**

-continued

Ile Asp Thr Lys Ala Ala Ile His Asp Pro Ala Leu Lys Ala Leu Cys
 35 40 45

Pro Thr Tyr Glu Gln Ala Val Ser Gln Arg Val Ile Asn Asn Gly His
 50 55 60

Ser Phe Asn Val Glu Phe Asp Asp Ser Gln Asp Asn Ala Val Leu Lys
 65 70 75 80

Gly Gly Pro Leu Thr Gly Thr Tyr Arg Leu Ile Gln Phe His Phe His
 85 90 95

Trp Gly Ser Ser Asp Gly Gln Gly Ser Glu His Thr Val Asp Lys Lys
 100 105 110

Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly
 115 120 125

Asp Phe Gly Lys Ala Val Gln Gln Pro Asp Gly Leu Ala Val Leu Gly
 130 135 140

Ile Phe Leu Lys Ile Gly Asp Ala Arg Pro Gly Leu Gln Lys Val Leu
 145 150 155 160

Asp Ala Leu Asp Ser Ile Lys Thr Lys Gly Lys Ser Ala Asp Phe Thr
 165 170 175

Asn Phe Asp Pro Arg Gly Leu Leu Pro Glu Ser Leu Asp Tyr Trp Thr
 180 185 190

Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp
 195 200 205

Ile Val Leu Lys Glu Pro Ile Ser Val Ser Ser Glu Gln Met Leu Lys
 210 215 220

Phe Arg Arg Leu Asn Phe Asn Lys Glu Gly Glu Pro Glu Glu Leu Met
 225 230 235 240

Val Asp Asn Trp Arg Pro Ala Gln Pro Leu His Asn Arg Gln Ile Asn
 245 250 255

Ala Ser Phe Lys
 260

<210> SEQ ID NO 31
 <211> LENGTH: 260
 <212> TYPE: PRT
 <213> ORGANISM: Equus caballus

<400> SEQUENCE: 31

Met Ser His His Trp Gly Tyr Gly Gln His Asn Gly Pro Lys His Trp
 1 5 10 15

His Lys Asp Phe Pro Ile Ala Lys Gly Gln Arg Gln Ser Pro Val Asp
 20 25 30

Ile Asp Thr Lys Ala Ala Val His Asp Ala Ala Leu Lys Pro Leu Ala
 35 40 45

Val His Tyr Glu Gln Ala Thr Ser Arg Arg Ile Val Asn Asn Gly His
 50 55 60

Ser Phe Asn Val Glu Phe Asp Asp Ser Gln Asp Lys Ala Val Leu Gln
 65 70 75 80

Gly Gly Pro Leu Thr Gly Thr Tyr Arg Leu Ile Gln Phe His Phe His
 85 90 95

Trp Gly Ser Ser Asp Gly Gln Gly Ser Glu His Thr Val Asp Lys Lys
 100 105 110

Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly
 115 120 125

Asp Phe Gly Lys Ala Val Gln Gln Pro Asp Gly Leu Ala Val Val Gly
 130 135 140

US 11,001,853 B2

105**106**

-continued

Val Phe Leu Lys Val Gly Gly Ala Lys Pro Gly Leu Gln Lys Val Leu
 145 150 155 160

Asp Val Leu Asp Ser Ile Lys Thr Lys Gly Lys Ser Ala Asp Phe Thr
 165 170 175

Asn Phe Asp Pro Arg Gly Leu Leu Pro Glu Ser Leu Asp Tyr Trp Thr
 180 185 190

Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp
 195 200 205

Ile Val Leu Arg Glu Pro Ile Ser Val Ser Ser Glu Gln Leu Leu Lys
 210 215 220

Phe Arg Ser Leu Asn Phe Asn Ala Glu Gly Lys Pro Glu Asp Pro Met
 225 230 235 240

Val Asp Asn Trp Arg Pro Ala Gln Pro Leu Asn Ser Arg Gln Ile Arg
 245 250 255

Ala Ser Phe Lys
 260

<210> SEQ ID NO 32

<211> LENGTH: 260

<212> TYPE: PRT

<213> ORGANISM: Canis lupus

<400> SEQUENCE: 32

Met Ala His His Trp Gly Tyr Ala Lys His Asn Gly Pro Glu His Trp
 1 5 10 15

His Lys Asp Phe Pro Ile Ala Lys Gly Glu Arg Gln Ser Pro Val Asp
 20 25 30

Ile Asp Thr Lys Ala Ala Val His Asp Pro Ala Leu Lys Ser Leu Cys
 35 40 45

Pro Cys Tyr Asp Gln Ala Val Ser Gln Arg Ile Ile Asn Asn Gly His
 50 55 60

Ser Phe Asn Val Glu Phe Asp Asp Ser Gln Asp Lys Thr Val Leu Lys
 65 70 75 80

Gly Gly Pro Leu Thr Gly Thr Tyr Arg Leu Ile Gln Phe His Phe His
 85 90 95

Trp Gly Ser Ser Asp Gly Gln Gly Ser Glu His Thr Val Asp Lys Lys
 100 105 110

Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly
 115 120 125

Glu Phe Gly Lys Ala Val Gln Gln Pro Asp Gly Leu Ala Val Leu Gly
 130 135 140

Ile Phe Leu Lys Ile Gly Gly Ala Asn Pro Gly Leu Gln Lys Ile Leu
 145 150 155 160

Asp Ala Leu Asp Ser Ile Lys Thr Lys Gly Lys Ser Ala Asp Phe Thr
 165 170 175

Asn Phe Asp Pro Arg Gly Leu Leu Pro Glu Ser Leu Asp Tyr Trp Thr
 180 185 190

Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp
 195 200 205

Ile Val Leu Lys Glu Pro Ile Ser Val Ser Ser Glu Gln Met Leu Lys
 210 215 220

Phe Arg Lys Leu Asn Phe Asn Lys Glu Gly Glu Pro Glu Glu Leu Met
 225 230 235 240

Met Asp Asn Trp Arg Pro Ala Gln Pro Leu His Ser Arg Gln Ile Asn

US 11,001,853 B2

107

-continued

108

245	250	255
-----	-----	-----

Ala Ser Phe Lys
260

<210> SEQ ID NO 33
<211> LENGTH: 260
<212> TYPE: PRT
<213> ORGANISM: Oryctolagus cuniculus

<400> SEQUENCE: 33

Met Ser His His Trp Gly Tyr Gly Lys His Asn Gly Pro Glu His Trp	5 10 15	
His Lys Asp Phe Pro Ile Ala Asn Gly Glu Arg Gln Ser Pro Ile Asp	20 25 30	
Ile Asp Thr Asn Ala Ala Lys His Asp Pro Ser Leu Lys Pro Leu Arg	35 40 45	
Val Cys Tyr Glu His Pro Ile Ser Arg Arg Ile Ile Asn Asn Gly His	50 55 60	
Ser Phe Asn Val Glu Phe Asp Asp Ser His Asp Lys Thr Val Leu Lys	65 70 75 80	
Glu Gly Pro Leu Glu Gly Thr Tyr Arg Leu Ile Gln Phe His Phe His	85 90 95	
Trp Gly Ser Ser Asp Gly Gln Gly Ser Glu His Thr Val Asn Lys Lys	100 105 110	
Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly	115 120 125	
Asp Phe Gly Lys Ala Val Lys His Pro Asp Gly Leu Ala Val Leu Gly	130 135 140	
Ile Phe Leu Lys Ile Gly Ser Ala Thr Pro Gly Leu Gln Lys Val Val	145 150 155 160	
Asp Thr Leu Ser Ser Ile Lys Thr Lys Gly Lys Ser Val Asp Phe Thr	165 170 175	
Asp Phe Asp Pro Arg Gly Leu Leu Pro Glu Ser Leu Asp Tyr Trp Thr	180 185 190	
Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp	195 200 205	
Ile Val Leu Lys Glu Pro Ile Thr Val Ser Ser Glu Gln Met Leu Lys	210 215 220	
Phe Arg Asn Leu Asn Phe Asn Lys Glu Ala Glu Pro Glu Glu Pro Met	225 230 235 240	
Val Asp Asn Trp Arg Pro Thr Gln Pro Leu Lys Gly Arg Gln Val Lys	245 250 255	
Ala Ser Phe Val	260	

<210> SEQ ID NO 34
<211> LENGTH: 249
<212> TYPE: PRT
<213> ORGANISM: Ailuropoda melanoleuca

<400> SEQUENCE: 34

Gly Pro Glu His Trp Tyr Lys Asp Phe Pro Ile Ala Lys Gly Gln Arg	5 10 15	
Gln Ser Pro Val Asp Ile Asp Thr Lys Ala Ala Ile His Asp Pro Ala	20 25 30	
Leu Lys Ala Leu Cys Pro Thr Tyr Glu Gln Ala Val Ser Gln Arg Val		

US 11,001,853 B2

109**110**

-continued

35 40 45

Ile Asn Asn Gly His Ser Phe Asn Val Glu Phe Asp Asp Ser Gln Asp
 50 55 60

Asn Ala Val Leu Lys Gly Gly Pro Leu Thr Gly Thr Tyr Arg Leu Ile
 65 70 75 80

Gln Phe His Phe His Trp Gly Ser Ser Asp Gly Gln Gly Ser Glu His
 85 90 95

Thr Val Asp Lys Lys Tyr Ala Ala Glu Leu His Leu Val His Trp
 100 105 110

Asn Thr Lys Tyr Gly Asp Phe Gly Lys Ala Val Gln Gln Pro Asp Gly
 115 120 125

Leu Ala Val Leu Gly Ile Phe Leu Lys Ile Gly Asp Ala Arg Pro Gly
 130 135 140

Leu Gln Lys Val Leu Asp Ala Leu Asp Ser Ile Lys Thr Lys Gly Lys
 145 150 155 160

Ser Ala Asp Phe Thr Asn Phe Asp Pro Arg Gly Leu Leu Pro Glu Ser
 165 170 175

Leu Asp Tyr Trp Thr Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu
 180 185 190

Glu Cys Val Thr Trp Ile Val Leu Lys Glu Pro Ile Ser Val Ser Ser
 195 200 205

Glu Gln Met Leu Lys Phe Arg Arg Leu Asn Phe Asn Lys Glu Gly Glu
 210 215 220

Pro Glu Glu Leu Met Val Asp Asn Trp Arg Pro Ala Gln Pro Leu His
 225 230 235 240

Asn Arg Gln Ile Asn Ala Ser Phe Lys
 245

<210> SEQ ID NO 35

<211> LENGTH: 260

<212> TYPE: PRT

<213> ORGANISM: Sus scrofa

<400> SEQUENCE: 35

Met Ser His His Trp Gly Tyr Asp Lys His Asn Gly Pro Glu His Trp
 1 5 10 15

His Lys Asp Phe Pro Ile Ala Lys Gly Asp Arg Gln Ser Pro Val Asp
 20 25 30

Ile Asn Thr Ser Thr Ala Val His Asp Pro Ala Leu Lys Pro Leu Ser
 35 40 45

Leu Cys Tyr Glu Gln Ala Thr Ser Gln Arg Ile Val Asn Asn Gly His
 50 55 60

Ser Phe Asn Val Glu Phe Asp Ser Ser Gln Asp Lys Gly Val Leu Glu
 65 70 75 80

Gly Gly Pro Leu Ala Gly Thr Tyr Arg Leu Ile Gln Phe His Phe His
 85 90 95

Trp Gly Ser Ser Asp Gly Gln Gly Ser Glu His Thr Val Asp Lys Lys
 100 105 110

Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Lys
 115 120 125

Asp Phe Gly Glu Ala Ala Gln Gln Pro Asp Gly Leu Ala Val Leu Gly
 130 135 140

Val Phe Leu Lys Ile Gly Asn Ala Gln Pro Gly Leu Gln Lys Ile Val
 145 150 155 160

US 11,001,853 B2

111**112**

-continued

Asp	Val	Leu	Asp	Ser	Ile	Lys	Thr	Lys	Gly	Lys	Ser	Val	Glu	Phe	Thr
															165
															170
															175
Gly	Phe	Asp	Pro	Arg	Asp	Leu	Leu	Pro	Gly	Ser	Leu	Asp	Tyr	Trp	Thr
															180
															185
															190
Tyr	Pro	Gly	Ser	Leu	Thr	Thr	Pro	Pro	Leu	Leu	Glu	Ser	Val	Thr	Trp
															195
															200
															205
Ile	Val	Leu	Arg	Glu	Pro	Ile	Ser	Val	Ser	Ser	Gly	Gln	Met	Met	Lys
															210
															215
															220
Phe	Arg	Thr	Leu	Asn	Phe	Asn	Lys	Glu	Gly	Glu	Pro	Glu	His	Pro	Met
															225
															230
															235
															240
Val	Asp	Asn	Trp	Arg	Pro	Thr	Gln	Pro	Leu	Lys	Asn	Arg	Gln	Ile	Arg
															245
															250
															255
Ala	Ser	Phe	Gln												
															260

<210> SEQ ID NO 36

<211> LENGTH: 235

<212> TYPE: PRT

<213> ORGANISM: Callithrix jacchus

<400> SEQUENCE: 36

Met	Ser	His	His	Trp	Gly	Tyr	Gly	Lys	His	Asn	Gly	Pro	Glu	His	Trp
1															
															5
															10
															15
His	Lys	Asp	Phe	Pro	Ile	Ala	Lys	Gly	Glu	Arg	Gln	Ser	Pro	Val	Asp
															20
															25
															30
Ile	Asp	Thr	His	Thr	Ala	Lys	Tyr	Asp	Pro	Ser	Leu	Lys	Pro	Leu	Ser
															35
															40
															45
Val	Ser	Tyr	Asp	Gln	Ala	Thr	Ser	Trp	Arg	Ile	Leu	Asn	Asn	Gly	His
															50
															55
															60
Ser	Phe	Asn	Val	Glu	Phe	Asp	Asp	Ser	Gln	Asp	Lys	Ala	Val	Leu	Lys
															65
															70
															75
															80
Gly	Gly	Pro	Leu	Asp	Gly	Thr	Tyr	Arg	Leu	Ile	Gln	Leu	His	Leu	Val
															85
															90
															95
His	Trp	Asn	Thr	Lys	Tyr	Gly	Asp	Phe	Gly	Lys	Ala	Ala	Gln	Gln	Pro
															100
															105
															110
Asp	Gly	Leu	Ala	Val	Leu	Gly	Ile	Phe	Leu	Lys	Val	Gly	Ser	Ala	Lys
															115
															120
															125
Pro	Gly	Leu	Gln	Lys	Val	Val	Asp	Val	Leu	Asp	Ser	Ile	Lys	Thr	Lys
															130
															135
															140
Gly	Lys	Ser	Ala	Asp	Phe	Thr	Asn	Phe	Asp	Pro	Arg	Gly	Leu	Leu	Pro
															145
															150
															155
															160
Glu	Ser	Leu	Asp	Tyr	Trp	Thr	Tyr	Pro	Gly	Ser	Leu	Thr	Thr	Pro	Pro
															165
															170
															175
Leu	Leu	Glu	Ser	Val	Thr	Trp	Ile	Val	Leu	Lys	Glu	Pro	Ile	Ser	Val
															180
															185
															190
Ser	Ser	Glu	Gln	Ile	Leu	Lys	Phe	Arg	Lys	Leu	Asn	Phe	Ser	Gly	Glu
															195
															200
															205
Gly	Glu	Pro	Glu	Glu	Leu	Met	Val	Asp	Asn	Trp	Arg	Pro	Ala	Gln	Pro
															210
															215
															220
Leu	Lys	Asn	Arg	Gln	Ile	Lys	Ala	Ser	Phe	Lys					
															225
															230
															235

<210> SEQ ID NO 37

<211> LENGTH: 260

<212> TYPE: PRT

<213> ORGANISM: Mus musculus

-continued

<400> SEQUENCE: 37

Met Ser His His Trp Gly Tyr Ser Lys His Asn Gly Pro Glu Asn Trp
 1 5 10 15

 His Lys Asp Phe Pro Ile Ala Asn Gly Asp Arg Gln Ser Pro Val Asp
 20 25 30

 Ile Asp Thr Ala Thr Ala Gln His Asp Pro Ala Leu Gln Pro Leu Leu
 35 40 45

 Ile Ser Tyr Asp Lys Ala Ala Ser Lys Ser Ile Val Asn Asn Gly His
 50 55 60

 Ser Phe Asn Val Glu Phe Asp Asp Ser Gln Asp Asn Ala Val Leu Lys
 65 70 75 80

 Gly Gly Pro Leu Ser Asp Ser Tyr Arg Leu Ile Gln Phe His Phe His
 85 90 95

 Trp Gly Ser Ser Asp Gly Gln Gly Ser Glu His Thr Val Asn Lys Lys
 100 105 110

 Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly
 115 120 125

 Asp Phe Gly Lys Ala Val Gln Gln Pro Asp Gly Leu Ala Val Leu Gly
 130 135 140

 Ile Phe Leu Lys Ile Gly Pro Ala Ser Gln Gly Leu Gln Lys Val Leu
 145 150 155 160

 Glu Ala Leu His Ser Ile Lys Thr Lys Gly Lys Arg Ala Ala Phe Ala
 165 170 175

 Asn Phe Asp Pro Cys Ser Leu Leu Pro Gly Asn Leu Asp Tyr Trp Thr
 180 185 190

 Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp
 195 200 205

 Ile Val Leu Arg Glu Pro Ile Thr Val Ser Ser Glu Gln Met Ser His
 210 215 220

 Phe Arg Thr Leu Asn Phe Asn Glu Glu Gly Asp Ala Glu Glu Ala Met
 225 230 235 240

 Val Asp Asn Trp Arg Pro Ala Gln Pro Leu Lys Asn Arg Lys Ile Lys
 245 250 255

 Ala Ser Phe Lys
 260

<210> SEQ ID NO 38

<211> LENGTH: 260

<212> TYPE: PRT

<213> ORGANISM: Bos taurus

<400> SEQUENCE: 38

Met Ser His His Trp Gly Tyr Gly Lys His Asn Gly Pro Glu His Trp
 1 5 10 15

 His Lys Asp Phe Pro Ile Ala Asn Gly Glu Arg Gln Ser Pro Val Asp
 20 25 30

 Ile Asp Thr Lys Ala Val Val Gln Asp Pro Ala Leu Lys Pro Leu Ala
 35 40 45

 Leu Val Tyr Gly Glu Ala Thr Ser Arg Arg Met Val Asn Asn Gly His
 50 55 60

 Ser Phe Asn Val Glu Tyr Asp Asp Ser Gln Asp Lys Ala Val Leu Lys
 65 70 75 80

 Asp Gly Pro Leu Thr Gly Thr Tyr Arg Leu Val Gln Phe His Phe His
 85 90 95

US 11,001,853 B2

115**116**

-continued

Trp Gly Ser Ser Asp Asp Gln Gly Ser Glu His Thr Val Asp Arg Lys
 100 105 110

Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly
 115 120 125

Asp Phe Gly Thr Ala Ala Gln Gln Pro Asp Gly Leu Ala Val Val Gly
 130 135 140

Val Phe Leu Lys Val Gly Asp Ala Asn Pro Ala Leu Gln Lys Val Leu
 145 150 155 160

Asp Ala Leu Asp Ser Ile Lys Thr Lys Gly Lys Ser Thr Asp Phe Pro
 165 170 175

Asn Phe Asp Pro Gly Ser Leu Leu Pro Asn Val Leu Asp Tyr Trp Thr
 180 185 190

Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Ser Val Thr Trp
 195 200 205

Ile Val Leu Lys Glu Pro Ile Ser Val Ser Ser Gln Gln Met Leu Lys
 210 215 220

Phe Arg Thr Leu Asn Phe Asn Ala Glu Gly Glu Pro Glu Leu Leu Met
 225 230 235 240

Leu Ala Asn Trp Arg Pro Ala Gln Pro Leu Lys Asn Arg Gln Val Arg
 245 250 255

Gly Phe Pro Lys
 260

<210> SEQ_ID NO 39
 <211> LENGTH: 232
 <212> TYPE: PRT
 <213> ORGANISM: Oryctolagus cuniculus

<400> SEQUENCE: 39

Gly Lys His Asn Gly Pro Glu His Trp His Lys Asp Phe Pro Ile Ala
 1 5 10 15

Asn Gly Glu Arg Gln Ser Pro Ile Asp Ile Asp Thr Asn Ala Ala Lys
 20 25 30

His Asp Pro Ser Leu Lys Pro Leu Arg Val Cys Tyr Glu His Pro Ile
 35 40 45

Ser Arg Arg Ile Ile Asn Asn Gly His Ser Phe Asn Val Glu Phe Asp
 50 55 60

Asp Ser His Asp Lys Thr Val Leu Lys Glu Gly Pro Leu Glu Gly Thr
 65 70 75 80

Tyr Arg Leu Ile Gln Phe His Phe His Trp Gly Ser Ser Asp Gly Gln
 85 90 95

Gly Ser Glu His Thr Val Asn Lys Lys Tyr Ala Ala Glu Leu His
 100 105 110

Leu Val His Trp Asn Thr Lys Tyr Gly Asp Phe Gly Lys Ala Val Lys
 115 120 125

His Pro Asp Gly Leu Ala Val Leu Gly Ile Phe Leu Lys Ile Gly Ser
 130 135 140

Ala Thr Pro Gly Leu Gln Lys Val Val Asp Thr Leu Ser Ser Ile Lys
 145 150 155 160

Thr Lys Gly Lys Ser Val Asp Phe Thr Asp Phe Asp Pro Arg Gly Leu
 165 170 175

Leu Pro Glu Ser Leu Asp Tyr Trp Thr Tyr Pro Gly Ser Leu Thr Thr
 180 185 190

Pro Pro Leu Leu Glu Cys Val Thr Trp Ile Val Leu Lys Glu Pro Ile

US 11,001,853 B2

117

-continued

118

195	200	205
-----	-----	-----

Thr Val Ser Ser Glu Gln Met Leu Lys Phe Arg Asn Leu Asn Phe Asn
210 215 220

Lys Glu Ala Glu Pro Glu Glu Pro
225 230

<210> SEQ ID NO 40
<211> LENGTH: 260
<212> TYPE: PRT
<213> ORGANISM: Rattus norvegicus

<400> SEQUENCE: 40

Met Ser His His Trp Gly Tyr Ser Lys Ser Asn Gly Pro Glu Asn Trp
1 5 10 15

His Lys Glu Phe Pro Ile Ala Asn Gly Asp Arg Gln Ser Pro Val Asp
20 25 30

Ile Asp Thr Gly Thr Ala Gln His Asp Pro Ser Leu Gln Pro Leu Leu
35 40 45

Ile Cys Tyr Asp Lys Val Ala Ser Lys Ser Ile Val Asn Asn Gly His
50 55 60

Ser Phe Asn Val Glu Phe Asp Asp Ser Gln Asp Phe Ala Val Leu Lys
65 70 75 80

Glu Gly Pro Leu Ser Gly Ser Tyr Arg Leu Ile Gln Phe His Phe His
85 90 95

Trp Gly Ser Ser Asp Gly Gln Gly Ser Glu His Thr Val Asn Lys Lys
100 105 110

Lys Tyr Ala Ala Glu Leu His Leu Val His Trp Asn Thr Lys Tyr Gly
115 120 125

Asp Phe Gly Lys Ala Val Gln His Pro Asp Gly Leu Ala Val Leu Gly
130 135 140

Ile Phe Leu Lys Ile Gly Pro Ala Ser Gln Gly Leu Gln Lys Ile Thr
145 150 155 160

Glu Ala Leu His Ser Ile Lys Thr Lys Gly Lys Arg Ala Ala Phe Ala
165 170 175

Asn Phe Asp Pro Cys Ser Leu Leu Pro Gly Asn Leu Asp Tyr Trp Thr
180 185 190

Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Leu Glu Cys Val Thr Trp
195 200 205

Ile Val Leu Lys Glu Pro Ile Thr Val Ser Ser Glu Gln Met Ser His
210 215 220

Phe Arg Lys Leu Asn Phe Asn Ser Glu Gly Glu Ala Glu Glu Leu Met
225 230 235 240

Val Asp Asn Trp Arg Pro Ala Gln Pro Leu Lys Asn Arg Lys Ile Lys
245 250 255

Ala Ser Phe Lys
260

<210> SEQ ID NO 41
<211> LENGTH: 208
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 41

Met Ser Leu Ser Ile Thr Asn Asn Gly His Ser Val Gln Val Asp Phe
1 5 10 15

Asn Asp Ser Asp Asp Arg Thr Val Val Thr Gly Gly Pro Leu Glu Gly

US 11,001,853 B2

119

-continued

120

20	25	30
Pro	Tyr	Arg
Leu	Lys	Gln
Phe	His	His
Trp	Gly	Lys
Asp		His
35	40	45
Val	Gly	Ser
Glu	His	Thr
Asp	Val	Asp
Gly	Lys	Ser
Pro	Ser	Glu
Leu		
50	55	60
His	Leu	Val
Val	His	Trp
Asn	Ala	Lys
Tyr		
70	75	80
Ala	Ser	Ala
Pro	Asp	Gly
Leu	Ala	Val
Val	Gly	Val
Phe		
85	90	95
Gly	Asp	Glu
His	Pro	Ser
Met	Asn	Arg
Leu	Thr	Asp
100	105	110
Val	Arg	Phe
Phe	Lys	Gly
Asp	Thr	lys
115	120	125
Cys	Leu	Leu
Pro	Ala	Ser
Arg	His	Tyr
130	135	140
Thr	Thr	Pro
Pro	Leu	Ser
Glu	Ser	Val
145	150	155
Pro	Ile	Cys
Ile	Ser	Glu
Arg	Gln	Met
Gly	Lys	Phe
165	170	175
Phe	Thr	Ser
Glu	Asp	Asp
Asp	Glu	Arg
180	185	190
Pro	Pro	Gln
Gln	Pro	Leu
Leu	Lys	Gly
Arg	Val	Val
195	200	205

<210> SEQ ID NO 42

<211> LENGTH: 264

<212> TYPE: PRT

<213> ORGANISM: Pongo abelii

<400> SEQUENCE: 42

Met	Thr	Gly	His	His	Gly	Trp	Gly	Tyr	Gly	Gln	Asp	Asp	Gly	Pro	Ser	
1						5		10		15						
His	Trp	His	Trp	Lys	Leu	Tyr	Pro	Ile	Ala	Gln	Gly	Asp	Arg	Gln	Ser	Pro
					20			25		30						
Ile	Asn	Ile	Ile	Ser	Ser	Gln	Ala	Val	Tyr	Ser	Pro	Ser	Leu	Gln	Pro	
					35			40		45						
Leu	Glu	Leu	Ser	Tyr	Glu	Ala	Cys	Met	Ser	Leu	Ser	Ile	Thr	Asn	Asn	
					50			55		60						
Gly	His	Ser	Val	Gln	Val	Asp	Phe	Asn	Asp	Ser	Asp	Asp	Arg	Thr	Val	
					65			70		75			80			
Val	Thr	Gly	Gly	Pro	Leu	Glu	Gly	Pro	Tyr	Arg	Leu	Lys	Gln	Phe	His	
					85			90		95						
Phe	His	Trp	Gly	Lys	His	Asp	Val	Gly	Ser	Glu	His	Thr	Val	Asp		
					100			105		110						
Gly	Lys	Ser	Phe	Pro	Ser	Glu	Leu	His	Leu	Val	His	Trp	Asn	Ala	Lys	
					115			120		125						
Lys	Tyr	Ser	Thr	Phe	Gly	Glu	Ala	Ala	Ser	Ala	Pro	Asp	Gly	Leu	Ala	
					130			135		140						
Val	Val	Gly	Val	Phe	Leu	Glu	Thr	Gly	Asp	Glu	His	Pro	Ser	Met	Asn	
					145			150		155			160			
Arg	Leu	Thr	Asp	Ala	Leu	Tyr	Met	Val	Arg	Phe	Lys	Gly	Thr	Lys	Ala	
					165			170		175						
Gln	Phe	Ser	Cys	Phe	Asn	Pro	Lys	Ser	Leu	Leu	Pro	Ala	Ser	Arg	His	
					180			185		190						

US 11,001,853 B2

121

122

-continued

Tyr Trp Thr Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Ser Glu Ser
195 200 205

Val Thr Trp Ile Val Leu Arg Glu Pro Ile Cys Ile Ser Glu Arg Gln
210 215 220

Met Gly Lys Phe Arg Ser Leu Leu Phe Thr Ser Glu Asp Asp Glu Arg
225 230 235 240

Ile His Met Val Asn Asn Phe Arg Pro Pro Gln Pro Leu Lys Gly Arg
245 250 255

Val Val Lys Ala Ser Phe Arg Ala
260

<210> SEQ ID NO 43

<211> LENGTH: 264

<212> TYPE: PRT

<213> ORGANISM: Pan troglodytes

<400> SEQUENCE: 43

Met Thr Gly His His Gly Trp Gly Tyr Gln Asp Asp Gly Pro Ser
1 5 10 15

His Trp His Lys Leu Tyr Pro Ile Ala Gln Gly Asp Arg Gln Ser Pro
20 25 30

Ile Asn Ile Ile Ser Ser Gln Ala Val Tyr Ser Pro Ser Leu Gln Pro
35 40 45

Leu Glu Leu Ser Tyr Glu Ala Cys Met Ser Leu Ser Ile Thr Asn Asn
50 55 60

Gly His Ser Val Gln Val Asp Phe Asn Asp Ser Asp Asp Arg Thr Val
65 70 75 80

Val Thr Gly Gly Pro Leu Glu Gly Pro Tyr Arg Leu Lys Gln Phe His
85 90 95

Phe His Trp Gly Lys His Asp Val Gly Ser Glu His Thr Val Asp
100 105 110

Gly Lys Ser Phe Pro Ser Glu Leu His Leu Val His Trp Asn Ala Lys
115 120 125

Lys Tyr Ser Thr Phe Gly Glu Ala Ala Ser Ala Pro Asp Gly Leu Ala
130 135 140

Val Val Gly Val Phe Leu Glu Thr Gly Asp Glu His Pro Ser Met Asn
145 150 155 160

Arg Leu Thr Asp Ala Leu Tyr Met Val Arg Phe Lys Gly Thr Lys Ala
165 170 175

Gln Phe Ser Cys Phe Asn Pro Lys Cys Leu Leu Pro Ala Ser Arg His
180 185 190

Tyr Trp Thr Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Ser Glu Ser
195 200 205

Val Thr Trp Ile Val Leu Arg Glu Pro Ile Cys Ile Ser Glu Arg Gln
210 215 220

Met Arg Lys Phe Arg Ser Leu Leu Phe Thr Ser Glu Asp Asp Glu Arg
225 230 235 240

Ile His Met Val Asn Asn Phe Arg Pro Pro Gln Pro Leu Lys Gly Arg
245 250 255

Val Val Lys Ala Ser Phe Arg Ala
260

<210> SEQ ID NO 44

<211> LENGTH: 264

<212> TYPE: PRT

<213> ORGANISM: Callithrix jacchus

-continued

<400> SEQUENCE: 44

Met	Thr	Gly	His	His	Gly	Trp	Gly	Tyr	Gly	Gln	Asp	Asp	Gly	Pro	Ser
1						5		10						15	

His	Trp	His	Lys	Leu	Tyr	Pro	Ile	Ala	Gln	Gly	Asp	Arg	Gln	Ser	Pro
						20		25					30		

Ile	Asn	Ile	Ile	Ser	Ser	Gln	Ala	Val	Tyr	Ser	Pro	Ser	Leu	Gln	Pro
						35		40				45			

Leu	Glu	Leu	Ser	Tyr	Glu	Ala	Cys	Met	Ser	Leu	Ser	Ile	Thr	Asn	Asn
						50		55			60				

Gly	His	Ser	Val	Gln	Val	Asp	Phe	Asn	Asp	Ser	Asp	Asp	Arg	Thr	Val
65						70		75					80		

Val	Thr	Gly	Gly	Pro	Leu	Glu	Gly	Pro	Tyr	Arg	Leu	Lys	Gln	Phe	His
						85		90			95				

Phe	His	Trp	Gly	Lys	Lys	His	Asp	Val	Gly	Ser	Glu	His	Thr	Val	Asp
						100		105			110				

Gly	Lys	Ser	Phe	Pro	Ser	Glu	Leu	His	Leu	Val	His	Trp	Asn	Ala	Lys
						115		120			125				

Lys	Tyr	Ser	Thr	Phe	Gly	Glu	Ala	Ala	Ser	Ala	Pro	Asp	Gly	Leu	Ala
						130		135			140				

Val	Val	Gly	Val	Phe	Leu	Glu	Thr	Gly	Asp	Glu	His	Pro	Ser	Met	Asn
145						150		155			160				

Arg	Leu	Thr	Asp	Ala	Leu	Tyr	Met	Val	Arg	Phe	Lys	Gly	Thr	Lys	Ala
						165		170			175				

Gln	Phe	Ser	Cys	Phe	Asn	Pro	Lys	Cys	Leu	Leu	Pro	Ala	Ser	Trp	His
						180		185			190				

Tyr	Trp	Thr	Tyr	Pro	Gly	Ser	Leu	Thr	Thr	Pro	Pro	Leu	Ser	Glu	Ser
						195		200			205				

Val	Thr	Trp	Ile	Val	Leu	Arg	Glu	Pro	Ile	Cys	Ile	Ser	Glu	Arg	Gln
						210		215			220				

Met	Gly	Lys	Phe	Arg	Ser	Leu	Leu	Phe	Thr	Ser	Glu	Asp	Asp	Glu	Arg
225						230		235			240				

Val	His	Met	Val	Asn	Asn	Phe	Arg	Pro	Pro	Gln	Pro	Leu	Lys	Gly	Arg
						245		250			255				

Val	Val	Lys	Ala	Ser	Phe	Arg	Ala								
						260									

<210> SEQ ID NO 45

<211> LENGTH: 251

<212> TYPE: PRT

<213> ORGANISM: Ailuropoda melanoleuca

<400> SEQUENCE: 45

Gly	Pro	Ser	Gln	Trp	His	Lys	Leu	Tyr	Pro	Ile	Ala	Gln	Gly	Asp	Arg
1						5		10			15				

Gln	Ser	Pro	Ile	Asn	Ile	Val	Ser	Ser	Gln	Ala	Val	Tyr	Ser	Pro	Ser
						20		25			30				

Leu	Lys	Pro	Leu	Glu	Leu	Ser	Tyr	Glu	Ala	Cys	Ile	Ser	Leu	Ser	Ile
						35		40			45				

Ala	Asn	Asn	Gly	His	Ser	Val	Gln	Val	Asp	Phe	Asn	Asp	Ser	Asp	Asp
						50		55			60				

Arg	Thr	Val	Val	Thr	Gly	Gly	Pro	Leu	Asp	Gly	Pro	Tyr	Arg	Leu	Lys
65						70		75			80				

Gln	Phe	His	Phe	His	Trp	Gly	Lys	Lys	His	Ser	Val	Gly	Ser	Glu	His
						85		90			95				

US 11,001,853 B2

125**126**

-continued

Thr Val Asp Gly Lys Ser Phe Pro Ser Glu Leu His Leu Val His Trp
 100 105 110
 Asn Ala Lys Lys Tyr Ser Thr Phe Gly Glu Ala Ala Ser Ala Pro Asp
 115 120 125
 Gly Leu Ala Val Val Gly Val Phe Leu Glu Thr Gly Asp Glu His Pro
 130 135 140
 Ser Met Asn Arg Leu Thr Asp Ala Leu Tyr Met Val Arg Phe Lys Gly
 145 150 155 160
 Thr Lys Ala Gln Phe Ser Cys Phe Asn Pro Lys Cys Leu Leu Pro Ala
 165 170 175
 Ser Arg His Tyr Trp Thr Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu
 180 185 190
 Ser Glu Ser Val Thr Trp Ile Val Leu Arg Glu Pro Ile Ser Ile Ser
 195 200 205
 Glu Arg Gln Met Glu Lys Phe Arg Ser Leu Leu Phe Thr Ser Glu Asp
 210 215 220
 Asp Glu Arg Ile His Met Val Asn Asn Phe Arg Pro Pro Gln Pro Leu
 225 230 235 240
 Lys Gly Arg Val Val Lys Ala Ser Phe Arg Ala
 245 250

<210> SEQ ID NO 46
 <211> LENGTH: 264
 <212> TYPE: PRT
 <213> ORGANISM: Canis familiaris
 <400> SEQUENCE: 46

Met Thr Gly His His Cys Trp Gly Tyr Gly Gln Asn Asp Gly Pro Ser
 1 5 10 15
 Gln Trp His Lys Leu Tyr Pro Ile Ala Gln Gly Asp Arg Gln Ser Pro
 20 25 30
 Ile Asn Ile Val Ser Ser Gln Ala Val Tyr Ser Pro Ser Leu Lys Pro
 35 40 45
 Leu Glu Leu Ser Tyr Glu Ala Cys Ile Ser Leu Ser Ile Thr Asn Asn
 50 55 60
 Gly His Ser Val Gln Val Asp Phe Asn Asp Ser Asp Asp Arg Thr Ala
 65 70 75 80
 Val Thr Gly Gly Pro Leu Asp Gly Pro Tyr Arg Leu Lys Gln Leu His
 85 90 95
 Phe His Trp Gly Lys Lys His Ser Val Gly Ser Glu His Thr Val Asp
 100 105 110
 Gly Lys Ser Phe Pro Ser Glu Leu His Leu Val His Trp Asn Ala Lys
 115 120 125
 Lys Tyr Ser Thr Phe Gly Glu Ala Ala Ser Ala Pro Asp Gly Leu Ala
 130 135 140
 Val Val Gly Ile Phe Leu Glu Thr Gly Asp Glu His Pro Ser Met Asn
 145 150 155 160
 Arg Leu Thr Asp Ala Leu Tyr Met Val Arg Phe Lys Gly Thr Lys Ala
 165 170 175
 Gln Phe Ser Cys Phe Asn Pro Lys Cys Leu Leu Pro Ala Ser Arg His
 180 185 190
 Tyr Trp Thr Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Ser Glu Ser
 195 200 205
 Val Thr Trp Ile Val Leu Arg Glu Pro Ile Ser Ile Ser Glu Arg Gln

US 11,001,853 B2

127

-continued

210	215	220
Met Glu Lys Phe Arg Ser Leu Leu Phe Thr Ser Glu Glu Asp Glu Arg		
225	230	235
Ile His Met Val Asn Asn Phe Arg Pro Pro Gln Pro Leu Lys Gly Arg		
245	250	255
Val Val Lys Ala Ser Phe Arg Ala		
260		

<210> SEQ ID NO 47

<211> LENGTH: 264

<212> TYPE: PRT

<213> ORGANISM: Bos taurus

<400> SEQUENCE: 47

Met Thr Gly His His Gly Trp Gly Tyr Gly Gln Asn Asp Gly Pro Ser			
1	5	10	15
His Trp His Lys Leu Tyr Pro Ile Ala Gln Gly Asp Arg Gln Ser Pro			
20	25	30	
Ile Asn Ile Val Ser Ser Gln Ala Val Tyr Ser Pro Ser Leu Lys Pro			
35	40	45	
Leu Glu Ile Ser Tyr Glu Ser Cys Thr Ser Leu Ser Ile Ala Asn Asn			
50	55	60	
Gly His Ser Val Gln Val Asp Phe Asn Asp Ser Asp Asp Arg Thr Val			
65	70	75	80
Val Ser Gly Gly Pro Leu Asp Gly Pro Tyr Arg Leu Lys Gln Phe His			
85	90	95	
Phe His Trp Gly Lys His Gly Val Gly Ser Glu His Thr Val Asp			
100	105	110	
Gly Lys Ser Phe Pro Ser Glu Leu His Leu Val His Trp Asn Ala Lys			
115	120	125	
Lys Tyr Ser Thr Phe Gly Glu Ala Ala Ser Ala Pro Asp Gly Leu Ala			
130	135	140	
Val Val Gly Val Phe Leu Glu Thr Gly Asp Glu His Pro Ser Met Asn			
145	150	155	160
Arg Leu Thr Asp Ala Leu Tyr Met Val Arg Phe Lys Gly Thr Lys Ala			
165	170	175	
Gln Phe Ser Cys Phe Asn Pro Lys Cys Leu Leu Pro Ala Ser Arg His			
180	185	190	
Tyr Trp Thr Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Ser Glu Ser			
195	200	205	
Val Thr Trp Ile Val Leu Arg Glu Pro Ile Arg Ile Ser Glu Arg Gln			
210	215	220	
Met Glu Lys Phe Arg Ser Leu Leu Phe Thr Ser Glu Glu Asp Glu Arg			
225	230	235	240
Ile His Met Val Asn Asn Phe Arg Pro Pro Gln Pro Leu Lys Gly Arg			
245	250	255	
Val Val Lys Ala Ser Phe Arg Ala			
260			

<210> SEQ ID NO 48

<211> LENGTH: 271

<212> TYPE: PRT

<213> ORGANISM: Rattus norvegicus

<400> SEQUENCE: 48

Met Thr Val Leu Trp Trp Pro Met Leu Arg Glu Glu Leu Met Ser Lys

128

US 11,001,853 B2

129**130**

-continued

1	5	10	15
Leu Arg Thr Gly Gly Pro Ser Asn Trp His Lys Leu Tyr Pro Ile Ala			
20	25	30	
Gln Gly Asp Arg Gln Ser Pro Ile Asn Ile Ile Ser Ser Gln Ala Val			
35	40	45	
Tyr Ser Pro Ser Leu Gln Pro Leu Glu Leu Phe Tyr Glu Ala Cys Met			
50	55	60	
Ser Leu Ser Ile Thr Asn Asn Gly His Ser Val Gln Val Asp Phe Asn			
65	70	75	80
Asp Ser Asp Asp Arg Thr Val Val Ala Gly Gly Pro Leu Glu Gly Pro			
85	90	95	
Tyr Arg Leu Lys Gln Leu His Phe His Trp Gly Lys Lys Arg Asp Val			
100	105	110	
Gly Ser Glu His Thr Val Asp Gly Lys Ser Phe Pro Ser Glu Leu His			
115	120	125	
Leu Val His Trp Asn Ala Lys Lys Tyr Ser Thr Phe Gly Glu Ala Ala			
130	135	140	
Ala Ala Pro Asp Gly Leu Ala Val Val Gly Ile Phe Leu Glu Thr Gly			
145	150	155	160
Asp Glu His Pro Ser Met Asn Arg Leu Thr Asp Ala Leu Tyr Met Val			
165	170	175	
Arg Phe Lys Asp Thr Lys Ala Gln Phe Ser Cys Phe Asn Pro Lys Cys			
180	185	190	
Leu Leu Pro Thr Ser Arg His Tyr Trp Thr Tyr Pro Gly Ser Leu Thr			
195	200	205	
Thr Pro Pro Leu Ser Glu Ser Val Thr Trp Ile Val Leu Arg Glu Pro			
210	215	220	
Ile Arg Ile Ser Glu Arg Gln Met Glu Lys Phe Arg Ser Leu Leu Phe			
225	230	235	240
Thr Ser Glu Asp Asp Glu Arg Ile His Met Val Asn Asn Phe Arg Pro			
245	250	255	
Pro Gln Pro Leu Lys Gly Arg Val Val Lys Ala Ser Phe Gln Ser			
260	265	270	

<210> SEQ ID NO 49

<211> LENGTH: 266

<212> TYPE: PRT

<213> ORGANISM: Oryctolagus cuniculus

<400> SEQUENCE: 49

Met Thr Gly His His Gly Trp Gly Tyr Gly Gln Asp Asp Gly Gly Arg			
1	5	10	15
Pro Ser His Trp His Lys Leu Tyr Pro Ile Ala Gln Gly Asp Arg Gln			
20	25	30	
Ser Pro Ile Asn Ile Val Ser Ser Gln Ala Val Tyr Ser Pro Gly Leu			
35	40	45	
Gln Pro Leu Glu Leu Ser Tyr Glu Ala Cys Thr Ser Leu Ser Ile Ala			
50	55	60	
Asn Asn Gly His Ser Val Gln Val Asp Phe Asn Asp Ser Asp Asp Arg			
65	70	75	80
Thr Val Val Thr Gly Gly Pro Leu Glu Gly Pro Tyr Arg Leu Lys Gln			
85	90	95	
Phe His Phe His Trp Gly Lys Arg Arg Asp Ala Gly Ser Glu His Thr			
100	105	110	

US 11,001,853 B2

131

132

-continued

Val Asp Gly Lys Ser Phe Pro Ser Glu Leu His Leu Val His Trp Asn
115 120 125

Ala Arg Lys Tyr Ser Thr Phe Gly Glu Ala Ala Ser Ala Pro Asp Gly
130 135 140

Leu Ala Val Val Gly Val Phe Leu Glu Thr Gly Asn Glu His Pro Ser
145 150 155 160

Met Asn Arg Leu Thr Asp Ala Leu Tyr Met Val Arg Phe Lys Gly Thr
165 170 175

Lys Ala Gln Phe Ser Cys Phe Asn Pro Lys Cys Leu Leu Pro Ser Ser
180 185 190

Arg His Tyr Trp Thr Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Ser
195 200 205

Glu Ser Val Thr Trp Ile Val Leu Arg Glu Pro Ile Ser Ile Ser Glu
210 215 220

Arg Gln Met Glu Lys Phe Arg Ser Leu Leu Phe Thr Ser Glu Asp Asp
225 230 235 240

Glu Arg Val His Met Val Asn Asn Phe Arg Pro Pro Gln Pro Leu Arg
245 250 255

Gly Arg Val Val Lys Ala Ser Phe Arg Ala
260 265

<210> SEQ ID NO 50
<211> LENGTH: 255
<212> TYPE: PRT
<213> ORGANISM: Mus musculus

<400> SEQUENCE: 50

Gly Gln Asp Asp Gly Pro Ser Asn Trp His Lys Leu Tyr Pro Ile Ala
1 5 10 15

Gln Gly Asp Arg Gln Ser Pro Ile Asn Ile Ile Ser Ser Gln Ala Val
20 25 30

Tyr Ser Pro Ser Leu Gln Pro Leu Glu Leu Phe Tyr Glu Ala Cys Met
35 40 45

Ser Leu Ser Ile Thr Asn Asn Gly His Ser Val Gln Val Asp Phe Asn
50 55 60

Asp Ser Asp Asp Arg Thr Val Val Ser Gly Gly Pro Leu Glu Gly Pro
65 70 75 80

Tyr Arg Leu Lys Gln Leu His Phe His Trp Gly Lys Lys Arg Asp Met
85 90 95

Gly Ser Glu His Thr Val Asp Gly Lys Ser Phe Pro Ser Glu Leu His
100 105 110

Leu Val His Trp Asn Ala Lys Lys Tyr Ser Thr Phe Gly Glu Ala Ala
115 120 125

Ala Ala Pro Asp Gly Leu Ala Val Val Gly Val Phe Leu Glu Thr Gly
130 135 140

Asp Glu His Pro Ser Met Asn Arg Leu Thr Asp Ala Leu Tyr Met Val
145 150 155 160

Arg Phe Lys Asp Thr Lys Ala Gln Phe Ser Cys Phe Asn Pro Lys Cys
165 170 175

Leu Leu Pro Thr Ser Arg His Tyr Trp Thr Tyr Pro Gly Ser Leu Thr
180 185 190

Thr Pro Pro Leu Ser Glu Ser Val Thr Trp Ile Val Leu Arg Glu Pro
195 200 205

Ile Arg Ile Ser Glu Arg Gln Met Glu Lys Phe Arg Ser Leu Leu Phe
210 215 220

-continued

Thr Ser Glu Asp Asp Glu Arg Ile His Met Val Asp Asn Phe Arg Pro
 225 230 235 240

Pro Gln Pro Leu Lys Gly Arg Val Val Lys Ala Ser Phe Gln Ala
 245 250 255

<210> SEQ ID NO 51

<211> LENGTH: 264

<212> TYPE: PRT

<213> ORGANISM: Monodelphis domestica

<400> SEQUENCE: 51

Met Thr Gly His His Gly Trp Gly Tyr Gly Gln Glu Asp Gly Pro Ser
 1 5 10 15

Glu Trp His Lys Leu Tyr Pro Ile Ala Gln Gly Asp Arg Gln Ser Pro
 20 25 30

Ile Asp Ile Val Ser Ser Gln Ala Val Tyr Asp Pro Thr Leu Lys Pro
 35 40 45

Leu Val Leu Ala Tyr Glu Ser Cys Met Ser Leu Ser Ile Ala Asn Asn
 50 55 60

Gly His Ser Val Met Val Glu Phe Asp Asp Val Asp Asp Arg Thr Val
 65 70 75 80

Val Asn Gly Gly Pro Leu Asp Gly Pro Tyr Arg Leu Lys Gln Phe His
 85 90 95

Phe His Trp Gly Lys Lys His Ser Leu Gly Ser Glu His Thr Val Asp
 100 105 110

Gly Lys Ser Phe Ser Ser Glu Leu His Leu Val His Trp Asn Gly Lys
 115 120 125

Lys Tyr Lys Thr Phe Ala Glu Ala Ala Ala Pro Asp Gly Leu Ala
 130 135 140

Val Val Gly Ile Phe Leu Glu Thr Gly Asp Glu His Ala Ser Met Asn
 145 150 155 160

Arg Leu Thr Asp Ala Leu Tyr Met Val Arg Phe Lys Gly Thr Lys Ala
 165 170 175

Gln Phe Asn Ser Phe Asn Pro Lys Cys Leu Leu Pro Met Asn Leu Ser
 180 185 190

Tyr Trp Thr Tyr Pro Gly Ser Leu Thr Thr Pro Pro Leu Ser Glu Ser
 195 200 205

Val Thr Trp Ile Val Leu Lys Glu Pro Ile Thr Ile Ser Glu Lys Gln
 210 215 220

Met Glu Lys Phe Arg Ser Leu Leu Phe Thr Ala Glu Glu Asp Glu Lys
 225 230 235 240

Val Arg Met Val Asn Asn Phe Arg Pro Pro Gln Pro Leu Lys Gly Arg
 245 250 255

Val Val Gln Ala Ser Phe Arg Ser
 260

<210> SEQ ID NO 52

<211> LENGTH: 264

<212> TYPE: PRT

<213> ORGANISM: Gallus gallus

<400> SEQUENCE: 52

Met Thr Gly His His Ser Trp Gly Tyr Gly Gln Asp Asp Gly Pro Ser
 1 5 10 15

Glu Trp His Lys Ser Tyr Pro Ile Ala Gln Gly Asn Arg Gln Ser Pro
 20 25 30

US 11,001,853 B2

135

136

-continued

Ile Asp Ile Ile Ser Ala Lys Ala Val Tyr Asp Pro Lys Leu Met Pro
35 40 45

Leu Val Ile Ser Tyr Glu Ser Cys Thr Ser Leu Asn Ile Ser Asn Asn
50 55 60

Gly His Ser Val Met Val Glu Phe Glu Asp Ile Asp Asp Lys Thr Val
65 70 75 80

Ile Ser Gly Gly Pro Phe Glu Ser Pro Phe Arg Leu Lys Gln Phe His
85 90 95

Phe His Trp Gly Ala Lys His Ser Glu Gly Ser Glu His Thr Ile Asp
100 105 110

Gly Lys Pro Phe Pro Cys Glu Leu His Leu Val His Trp Asn Ala Lys
115 120 125

Lys Tyr Ala Thr Phe Gly Glu Ala Ala Ala Pro Asp Gly Leu Ala
130 135 140

Val Val Gly Val Phe Leu Glu Ile Gly Lys Glu His Ala Asn Met Asn
145 150 155 160

Arg Leu Thr Asp Ala Leu Tyr Met Val Lys Phe Lys Gly Thr Lys Ala
165 170 175

Gln Phe Arg Ser Phe Asn Pro Lys Cys Leu Leu Pro Leu Ser Leu Asp
180 185 190

Tyr Trp Thr Tyr Leu Gly Ser Leu Thr Thr Pro Pro Leu Asn Glu Ser
195 200 205

Val Ile Trp Val Val Leu Lys Glu Pro Ile Ser Ile Ser Glu Lys Gln
210 215 220

Leu Glu Lys Phe Arg Met Leu Leu Phe Thr Ser Glu Glu Asp Gln Lys
225 230 235 240

Val Gln Met Val Asn Asn Phe Arg Pro Pro Gln Pro Leu Lys Gly Arg
245 250 255

Thr Val Arg Ala Ser Phe Lys Ala
260

<210> SEQ_ID NO 53
<211> LENGTH: 264
<212> TYPE: PRT
<213> ORGANISM: Taeniopygia guttata

<400> SEQUENCE: 53

Met Thr Gly Gln His Ser Trp Gly Tyr Gly Gln Ala Asp Gly Pro Ser
1 5 10 15

Glu Trp His Lys Ala Tyr Pro Ile Ala Gln Gly Asn Arg Gln Ser Pro
20 25 30

Ile Asp Ile Asp Ser Ala Arg Ala Val Tyr Asp Pro Ser Leu Gln Pro
35 40 45

Leu Leu Ile Ser Tyr Glu Ser Cys Ser Ser Leu Ser Ile Ser Asn Thr
50 55 60

Gly His Ser Val Met Val Glu Phe Glu Asp Thr Asp Asp Arg Thr Ala
65 70 75 80

Ile Ser Gly Gly Pro Phe Gln Asn Pro Phe Arg Leu Lys Gln Phe His
85 90 95

Phe His Trp Gly Thr Thr His Ser Gln Gly Ser Glu His Thr Ile Asp
100 105 110

Gly Lys Pro Phe Pro Cys Glu Leu His Leu Val His Trp Asn Ala Arg
115 120 125

Lys Tyr Thr Thr Phe Gly Glu Ala Ala Ala Pro Asp Gly Leu Ala

US 11,001,853 B2

137

138

-continued

130 135 140

Val Val Gly Val Phe Leu Glu Ile Gly Lys Glu His Ala Ser Met Asn
 145 150 155 160

Arg Leu Thr Asp Ala Leu Tyr Met Val Lys Phe Lys Gly Thr Lys Ala
 165 170 175

Gln Phe Arg Gly Phe Asn Pro Lys Cys Leu Leu Pro Leu Ser Leu Asp
 180 185 190

Tyr Trp Thr Tyr Leu Gly Ser Leu Thr Thr Pro Pro Leu Asn Glu Ser
 195 200 205

Val Thr Trp Ile Val Leu Lys Glu Pro Ile Arg Ile Ser Val Lys Gln
 210 215 220

Leu Glu Lys Phe Arg Met Leu Leu Phe Thr Gly Glu Glu Asp Gln Arg
 225 230 235 240

Ile Gln Met Ala Asn Asn Phe Arg Pro Pro Gln Pro Leu Lys Gly Arg
 245 250 255

Ile Val Arg Ala Ser Phe Lys Ala
 260

<210> SEQ ID NO 54

<211> LENGTH: 262

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 54

Met Ser Arg Leu Ser Trp Gly Tyr Arg Glu His Asn Gly Pro Ile His
 1 5 10 15

Trp Lys Glu Phe Phe Pro Ile Ala Asp Gly Asp Gln Gln Ser Pro Ile
 20 25 30

Glu Ile Lys Thr Lys Glu Val Lys Tyr Asp Ser Ser Leu Arg Pro Leu
 35 40 45

Ser Ile Lys Tyr Asp Pro Ser Ser Ala Lys Ile Ile Ser Asn Ser Gly
 50 55 60

His Ser Phe Asn Val Asp Phe Asp Asp Thr Glu Asn Lys Ser Val Leu
 65 70 75 80

Arg Gly Gly Pro Leu Thr Gly Ser Tyr Arg Leu Arg Gln Val His Leu
 85 90 95

His Trp Gly Ser Ala Asp Asp His Gly Ser Glu His Ile Val Asp Gly
 100 105 110

Val Ser Tyr Ala Ala Glu Leu His Val Val His Trp Asn Ser Asp Lys
 115 120 125

Tyr Pro Ser Phe Val Glu Ala Ala His Glu Pro Asp Gly Leu Ala Val
 130 135 140

Leu Gly Val Phe Leu Gln Ile Gly Glu Pro Asn Ser Gln Leu Gln Lys
 145 150 155 160

Ile Thr Asp Thr Leu Asp Ser Ile Lys Glu Lys Gly Lys Gln Thr Arg
 165 170 175

Phe Thr Asn Phe Asp Leu Leu Ser Leu Leu Pro Pro Ser Trp Asp Tyr
 180 185 190

Trp Thr Tyr Pro Gly Ser Leu Thr Val Pro Pro Leu Leu Glu Ser Val
 195 200 205

Thr Trp Ile Val Leu Lys Gln Pro Ile Asn Ile Ser Ser Gln Gln Leu
 210 215 220

Ala Lys Phe Arg Ser Leu Leu Cys Thr Ala Glu Gly Glu Ala Ala Ala
 225 230 235 240

US 11,001,853 B2

139**140**

-continued

Phe	Leu	Val	Ser	Asn	His	Arg	Pro	Pro	Gln	Pro	Leu	Lys	Gly	Arg	Lys
245									250			255			

Val	Arg	Ala	Ser	Phe	His										
		260													

<210> SEQ ID NO 55

<211> LENGTH: 262

<212> TYPE: PRT

<213> ORGANISM: Pan troglodytes

<400> SEQUENCE: 55

Met	Ser	Arg	Leu	Ser	Trp	Gly	Tyr	Arg	Glu	His	Asn	Gly	Pro	Ile	His
1			5				10						15		

Trp	Lys	Glu	Phe	Phe	Pro	Ile	Ala	Asp	Gly	Asp	Gln	Gln	Ser	Pro	Ile
	20					25					30				

Glu	Ile	Lys	Thr	Lys	Glu	Val	Lys	Tyr	Asp	Ser	Ser	Leu	Arg	Pro	Leu
	35			40				45							

Ser	Ile	Lys	Tyr	Asp	Pro	Ser	Ser	Ala	Lys	Ile	Ile	Ser	Asn	Ser	Gly
	50					55				60					

His	Ser	Phe	Asn	Val	Asp	Phe	Asp	Asp	Thr	Glu	Asn	Lys	Ser	Val	Leu
	65				70				75			80			

Arg	Gly	Gly	Pro	Leu	Thr	Gly	Ser	Tyr	Arg	Leu	Arg	Gln	Phe	His	Leu
	85					90			95						

His	Trp	Gly	Ser	Ala	Asp	Asp	His	Gly	Ser	Glu	His	Ile	Val	Asp	Gly
	100					105				110					

Val	Ser	Tyr	Ala	Ala	Glu	Leu	His	Val	Val	His	Trp	Asn	Ser	Asp	Lys
	115				120				125						

Tyr	Pro	Ser	Phe	Val	Glu	Ala	Ala	His	Glu	Pro	Asp	Gly	Leu	Ala	Val
	130				135				140						

Leu	Gly	Val	Phe	Leu	Gln	Ile	Gly	Glu	Pro	Asn	Ser	Gln	Leu	Gln	Lys
	145				150			155			160				

Ile	Thr	Asp	Thr	Leu	Asp	Ser	Ile	Lys	Glu	Lys	Gly	Lys	Gln	Thr	Arg
	165					170				175					

Phe	Thr	Asn	Phe	Asp	Pro	Leu	Ser	Leu	Leu	Pro	Pro	Ser	Trp	Asp	Tyr
	180					185			190						

Trp	Thr	Tyr	Pro	Gly	Ser	Leu	Thr	Val	Pro	Pro	Leu	Leu	Glu	Ser	Val
	195					200			205						

Thr	Trp	Ile	Val	Leu	Lys	Gln	Pro	Ile	Asn	Ile	Ser	Ser	Gln	Gln	Leu
	210				215			220							

Ala	Lys	Phe	Arg	Ser	Leu	Leu	Cys	Thr	Ala	Glu	Gly	Glu	Ala	Ala	Ala
	225				230			235			240				

Phe	Leu	Val	Ser	Asn	His	Arg	Pro	Pro	Gln	Pro	Leu	Lys	Gly	Arg	Lys
	245					245			250			255			

Val	Arg	Ala	Ser	Phe	His										
	260														

<210> SEQ ID NO 56

<211> LENGTH: 262

<212> TYPE: PRT

<213> ORGANISM: Macaca mulatta

<400> SEQUENCE: 56

Met	Ser	Arg	Leu	Ser	Trp	Gly	Tyr	Arg	Glu	His	Asn	Gly	Pro	Ile	His
1			5				10			15					

Trp	Lys	Glu	Phe	Phe	Pro	Ile	Ala	Asp	Gly	Asp	Gln	Gln	Ser	Pro	Ile
	20				25				30						

US 11,001,853 B2

141

142

-continued

Glu Ile Lys Thr Gln Glu Val Lys Tyr Asp Ser Ser Leu Arg Pro Leu
 35 40 45

Ser Ile Lys Tyr Asp Pro Ser Ser Ala Lys Ile Ile Ser Asn Ser Gly
 50 55 60

His Ser Phe Asn Val Asp Phe Asp Asp Thr Glu Asp Lys Ser Val Leu
 65 70 75 80

Arg Gly Gly Pro Leu Ala Gly Ser Tyr Arg Leu Arg Gln Phe His Leu
 85 90 95

His Trp Gly Ser Ala Asp Asp His Gly Ser Glu His Ile Val Asp Gly
 100 105 110

Val Ser Tyr Ala Ala Glu Leu His Val Val His Trp Asn Ser Asp Lys
 115 120 125

Tyr Pro Ser Phe Val Glu Ala Ala His Glu Pro Asp Gly Leu Ala Val
 130 135 140

Leu Gly Val Phe Leu Gln Ile Gly Glu Pro Asn Ser Gln Leu Gln Lys
 145 150 155 160

Ile Thr Asp Ile Leu Asp Ser Ile Lys Glu Lys Gly Lys Gln Thr Arg
 165 170 175

Phe Thr Asn Phe Asp Pro Leu Ser Leu Leu Pro Pro Ser Trp Asp Tyr
 180 185 190

Trp Thr Tyr Pro Gly Ser Leu Thr Val Pro Pro Leu Leu Glu Ser Val
 195 200 205

Ile Trp Ile Val Leu Lys Gln Pro Ile Asn Val Ser Ser Gln Gln Leu
 210 215 220

Ala Lys Phe Arg Ser Leu Leu Cys Thr Ala Glu Gly Glu Ala Ala Ala
 225 230 235 240

Phe Leu Leu Ser Asn His Arg Pro Pro Gln Pro Leu Lys Gly Arg Lys
 245 250 255

Val Arg Ala Ser Phe Arg
 260

<210> SEQ ID NO 57
 <211> LENGTH: 262
 <212> TYPE: PRT
 <213> ORGANISM: Oryctolagus cuniculus

<400> SEQUENCE: 57

Met Ser Arg Ile Ser Trp Gly Tyr Gly Glu His Asn Gly Pro Ile His
 1 5 10 15

Trp Asn Gln Phe Phe Pro Ile Ala Asp Gly Asp Gln Gln Ser Pro Ile
 20 25 30

Glu Ile Lys Thr Lys Glu Val Lys Tyr Asp Ser Ser Leu Arg Pro Leu
 35 40 45

Ser Ile Lys Tyr Asp Pro Ser Ser Ala Lys Ile Ile Ser Asn Ser Gly
 50 55 60

His Ser Phe Asn Val Asp Phe Asp Asp Thr Glu Asp Lys Ser Val Leu
 65 70 75 80

Arg Gly Gly Pro Leu Thr Gly Asn Tyr Arg Leu Arg Gln Phe His Leu
 85 90 95

His Trp Gly Ser Ala Asp Asp His Gly Ser Glu His Val Val Asp Gly
 100 105 110

Val Arg Tyr Ala Ala Glu Leu His Val Val His Trp Asn Ser Asp Lys
 115 120 125

Tyr Pro Ser Phe Val Glu Ala Ala His Glu Pro Asp Gly Leu Ala Val
 130 135 140

-continued

Leu Gly Val Phe Leu Gln Ile Gly Glu Tyr Asn Ser Gln Leu Gln Lys
 145 150 155 160
 Ile Thr Asp Ile Leu Asp Ser Ile Lys Glu Lys Gly Lys Gln Thr Arg
 165 170 175
 Phe Thr Asn Phe Asp Pro Leu Ser Leu Leu Pro Ser Ser Trp Asp Tyr
 180 185 190
 Trp Thr Tyr Pro Gly Ser Leu Thr Val Pro Pro Leu Leu Glu Ser Val
 195 200 205
 Thr Trp Ile Val Leu Lys Gln Pro Ile Asn Ile Ser Ser Gln Gln Leu
 210 215 220
 Ala Lys Phe Arg Ser Leu Leu Cys Ser Ala Glu Gly Glu Ser Ala Ala
 225 230 235 240
 Phe Leu Leu Ser Asn His Arg Pro Pro Gln Pro Leu Lys Gly Arg Lys
 245 250 255
 Val Arg Ala Ser Phe His
 260

<210> SEQ ID NO 58
 <211> LENGTH: 262
 <212> TYPE: PRT
 <213> ORGANISM: Ailuropoda melanoleuca
 <400> SEQUENCE: 58

Met Ser Arg Leu Ser Trp Gly Tyr Gly Glu His Asn Gly Pro Ile His
 1 5 10 15
 Trp Asn Lys Phe Phe Pro Ile Ala Asp Gly Asp Gln Gln Ser Pro Ile
 20 25 30
 Glu Ile Lys Thr Lys Glu Val Lys Tyr Asp Ser Ser Leu Arg Pro Leu
 35 40 45
 Ser Ile Lys Tyr Asp Ala Asn Ser Ala Lys Ile Ile Ser Asn Ser Gly
 50 55 60
 His Ser Phe Ser Val Asp Phe Asp Asp Thr Glu Asp Lys Ser Val Leu
 65 70 75 80
 Arg Gly Gly Pro Leu Thr Gly Ser Tyr Arg Leu Arg Gln Phe His Leu
 85 90 95
 His Trp Gly Ser Ala Asp Asp His Gly Ser Glu His Val Val Asp Gly
 100 105 110
 Val Arg Tyr Ala Ala Glu Leu His Val Val His Trp Asn Ser Asp Lys
 115 120 125
 Tyr Pro Ser Phe Val Glu Ala Ala His Glu Pro Asp Gly Leu Ala Val
 130 135 140
 Leu Gly Val Phe Leu Gln Ile Gly Glu His Asn Ser Gln Leu Gln Lys
 145 150 155 160
 Ile Thr Asp Ile Leu Asp Ser Ile Lys Glu Lys Gly Lys Gln Thr Arg
 165 170 175
 Phe Thr Asn Phe Asp Pro Leu Ser Leu Leu Pro Pro Ser Trp Asp Tyr
 180 185 190
 Trp Thr Tyr Pro Gly Ser Leu Thr Val Pro Pro Leu Leu Glu Ser Val
 195 200 205
 Thr Trp Ile Val Leu Lys Gln Pro Ile Asn Ile Ser Ser Glu Gln Leu
 210 215 220
 Ala Thr Phe Arg Thr Leu Leu Cys Thr Ala Glu Gly Glu Ala Ala Ala
 225 230 235 240
 Phe Leu Leu Ser Asn His Arg Pro Pro Gln Pro Leu Lys Gly Arg Lys

US 11,001,853 B2

145**146**

-continued

245

250

255

Val Arg Ala Ser Phe His
260

<210> SEQ ID NO 59

<211> LENGTH: 262

<212> TYPE: PRT

<213> ORGANISM: Sus scrofa

<400> SEQUENCE: 59

Met	Ser	Arg	Phe	Ser	Trp	Gly	Tyr	Gly	Glu	His	Asn	Gly	Pro	Val	His
1					5			10			15				

Trp	Asn	Glu	Phe	Phe	Pro	Ile	Ala	Asp	Gly	Asp	Gln	Gln	Ser	Pro	Ile
	20				25						30				

Glu	Ile	Lys	Thr	Lys	Glu	Val	Lys	Tyr	Asp	Ser	Ser	Leu	Arg	Pro	Leu
	35				40						45				

Ser	Ile	Lys	Tyr	Asp	Pro	Ser	Ser	Ala	Lys	Ile	Ile	Ser	Asn	Ser	Gly
	50				55					60					

His	Ser	Phe	Ser	Val	Asp	Phe	Asp	Asp	Thr	Glu	Asp	Lys	Ser	Val	Leu
	65				70			75		80					

Arg	Gly	Gly	Pro	Leu	Thr	Gly	Ser	Tyr	Arg	Leu	Arg	Gln	Phe	His	Leu
	85				90					95					

His	Trp	Gly	Ser	Ala	Asp	Asp	His	Gly	Ser	Glu	His	Val	Val	Asp	Gly
	100				105					110					

Val	Lys	Tyr	Ala	Ala	Glu	Leu	His	Val	Val	His	Trp	Asn	Ser	Asp	Lys
	115				120					125					

Tyr	Pro	Ser	Phe	Val	Glu	Ala	Ala	His	Glu	Pro	Asp	Gly	Leu	Ala	Val
	130			135				140							

Leu	Gly	Val	Phe	Leu	Gln	Ile	Gly	Glu	His	Asn	Ser	Gln	Leu	Gln	Lys
	145			150			155		160						

Ile	Thr	Asp	Ile	Leu	Asp	Ser	Ile	Lys	Glu	Lys	Gly	Lys	Gln	Thr	Arg
	165			170				175							

Phe	Thr	Asn	Phe	Asp	Pro	Leu	Ser	Leu	Leu	Pro	Pro	Ser	Trp	Asp	Tyr
	180			185				190							

Trp	Thr	Thr	Pro	Gly	Ser	Leu	Thr	Val	Pro	Pro	Leu	Leu	Glu	Ser	Val
	195			200				205							

Thr	Trp	Ile	Ile	Leu	Lys	Gln	Pro	Ile	Asn	Ile	Ser	Ser	Gln	Gln	Leu
	210			215				220							

Ala	Thr	Phe	Arg	Thr	Leu	Leu	Cys	Thr	Lys	Glu	Gly	Glu	Glu	Ala	Ala
	225			230			235		240						

Phe	Leu	Leu	Ser	Asn	His	Arg	Pro	Leu	Gln	Pro	Leu	Lys	Gly	Arg	Lys
	245			250			255		260						

Val Arg Ala Ser Phe His
260

<210> SEQ ID NO 60

<211> LENGTH: 262

<212> TYPE: PRT

<213> ORGANISM: Callithrix jacchus

<400> SEQUENCE: 60

Met	Ser	Arg	Leu	Ser	Trp	Gly	Tyr	Gly	Glu	His	Asn	Gly	Pro	Ile	His
1					5			10		15					

Trp	Asn	Glu	Phe	Phe	Pro	Ile	Ala	Asp	Gly	Asp	Arg	Gln	Ser	Pro	Ile
	20				25					30					

Glu Ile Lys Ala Lys Glu Val Lys Tyr Asp Ser Ser Leu Arg Pro Leu

US 11,001,853 B2

147

-continued

35	40	45
Ser Ile Lys Tyr Asp Pro Ser Ser Ala Lys Ile Ile Ser Asn Ser Gly		
50	55	60
His Ser Phe Asn Val Asp Phe Asp Asp Thr Glu Asp Lys Ser Val Leu		
65	70	75
80		
His Gly Gly Pro Leu Thr Gly Ser Tyr Arg Leu Arg Gln Phe His Leu		
85	90	95
His Trp Gly Ser Ala Asp Asp His Gly Ser Glu His Val Val Asp Gly		
100	105	110
Val Arg Tyr Ala Ala Glu Leu His Val Val His Trp Asn Ser Glu Lys		
115	120	125
Tyr Pro Ser Phe Val Glu Ala Ala His Glu Pro Asp Gly Leu Ala Val		
130	135	140
Leu Gly Val Phe Leu Gln Ile Gly Glu Pro Asn Ser Gln Leu Gln Lys		
145	150	155
160		
Ile Ile Asp Ile Leu Asp Ser Ile Lys Glu Lys Gly Lys Gln Ile Arg		
165	170	175
Phe Thr Asn Phe Asp Pro Leu Ser Leu Phe Pro Pro Ser Trp Asp Tyr		
180	185	190
Trp Thr Tyr Ser Gly Ser Leu Thr Val Pro Pro Leu Leu Glu Ser Val		
195	200	205
Thr Trp Ile Leu Leu Lys Gln Pro Ile Asn Ile Ser Ser Gln Gln Leu		
210	215	220
Ala Lys Phe Arg Ser Leu Leu Cys Thr Ala Glu Gly Glu Ala Ala Ala		
225	230	235
240		
Phe Leu Leu Ser Asn Tyr Arg Pro Pro Gln Pro Leu Lys Gly Arg Lys		
245	250	255
Val Arg Ala Ser Phe Arg		
260		

<210> SEQ_ID NO 61

<211> LENGTH: 262

<212> TYPE: PRT

<213> ORGANISM: Rattus norvegicus

<400> SEQUENCE: 61

Met Ala Arg Leu Ser Trp Gly Tyr Asp Glu His Asn Gly Pro Ile His		
1	5	10
15		
Trp Asn Glu Leu Phe Pro Ile Ala Asp Gly Asp Gln Gln Ser Pro Ile		
20	25	30
Glu Ile Lys Thr Lys Glu Val Lys Tyr Asp Ser Ser Leu Arg Pro Leu		
35	40	45
Ser Ile Lys Tyr Asp Pro Ala Ser Ala Lys Ile Ile Ser Asn Ser Gly		
50	55	60
His Ser Phe Asn Val Asp Phe Asp Asp Thr Glu Asp Lys Ser Val Leu		
65	70	75
80		
Arg Gly Gly Pro Leu Thr Gly Ser Tyr Arg Leu Arg Gln Phe His Leu		
85	90	95
His Trp Gly Ser Ala Asp Asp His Gly Ser Glu His Val Val Asp Gly		
100	105	110
Val Arg Tyr Ala Ala Glu Leu His Val Val His Trp Asn Ser Asp Lys		
115	120	125
Tyr Pro Ser Phe Val Glu Ala Ala His Glu Ser Asp Gly Leu Ala Val		
130	135	140

US 11,001,853 B2

149**150**

-continued

Leu Gly Val Phe Leu Gln Ile Gly Glu His Asn Pro Gln Leu Gln Lys
 145 150 155 160
 Ile Thr Asp Ile Leu Asp Ser Ile Lys Glu Lys Gly Lys Gln Thr Arg
 165 170 175
 Phe Thr Asn Phe Asp Pro Leu Cys Leu Leu Pro Ser Ser Trp Asp Tyr
 180 185 190
 Trp Thr Tyr Pro Gly Ser Leu Thr Val Pro Pro Leu Leu Glu Ser Val
 195 200 205
 Thr Trp Ile Val Leu Lys Gln Pro Ile Ser Ile Ser Gln Gln Leu
 210 215 220
 Ala Arg Phe Arg Ser Leu Leu Cys Thr Ala Glu Gly Glu Ser Ala Ala
 225 230 235 240
 Phe Leu Leu Ser Asn His Arg Pro Pro Gln Pro Leu Lys Gly Arg Arg
 245 250 255
 Val Arg Ala Ser Phe Tyr
 260

<210> SEQ_ID NO 62
 <211> LENGTH: 262
 <212> TYPE: PRT
 <213> ORGANISM: Mus musculus

<400> SEQUENCE: 62

Met Ala Arg Leu Ser Trp Gly Tyr Gly Glu His Asn Gly Pro Ile His
 1 5 10 15
 Trp Asn Glu Leu Phe Pro Ile Ala Asp Gly Asp Gln Gln Ser Pro Ile
 20 25 30
 Glu Ile Lys Thr Lys Glu Val Lys Tyr Asp Ser Ser Leu Arg Pro Leu
 35 40 45
 Ser Ile Lys Tyr Asp Pro Ala Ser Ala Lys Ile Ile Ser Asn Ser Gly
 50 55 60
 His Ser Phe Asn Val Asp Phe Asp Asp Thr Glu Asp Lys Ser Val Leu
 65 70 75 80
 Arg Gly Gly Pro Leu Thr Gly Asn Tyr Arg Leu Arg Gln Phe His Leu
 85 90 95
 His Trp Gly Ser Ala Asp Asp His Gly Ser Glu His Val Val Asp Gly
 100 105 110
 Val Arg Tyr Ala Ala Glu Leu His Val Val His Trp Asn Ser Asp Lys
 115 120 125
 Tyr Pro Ser Phe Val Glu Ala Ala His Glu Ser Asp Gly Leu Ala Val
 130 135 140
 Leu Gly Val Phe Leu Gln Ile Gly Glu His Asn Pro Gln Leu Gln Lys
 145 150 155 160
 Ile Thr Asp Ile Leu Asp Ser Ile Lys Glu Lys Gly Lys Gln Thr Arg
 165 170 175
 Phe Thr Asn Phe Asp Pro Leu Cys Leu Leu Pro Ser Ser Trp Asp Tyr
 180 185 190
 Trp Thr Tyr Pro Gly Ser Leu Thr Val Pro Pro Leu Leu Glu Ser Val
 195 200 205
 Thr Trp Ile Val Leu Lys Gln Pro Ile Ser Ile Ser Gln Gln Leu
 210 215 220
 Ala Arg Phe Arg Ser Leu Leu Cys Thr Ala Glu Gly Glu Ser Ala Ala
 225 230 235 240
 Phe Leu Leu Ser Asn His Arg Pro Pro Gln Pro Leu Lys Gly Arg Arg
 245 250 255

US 11,001,853 B2

151

-continued

Val Arg Ala Ser Phe Tyr
260

<210> SEQ ID NO 63
<211> LENGTH: 262
<212> TYPE: PRT
<213> ORGANISM: Canis familiaris

<400> SEQUENCE: 63

Met Ser Arg Leu Ser Trp Gly Tyr Gly Glu His Asn Gly Pro Ile His
1 5 10 15

Trp Asn Lys Phe Phe Pro Ile Ala Asp Gly Asp Gln Gln Ser Pro Ile
20 25 30

Glu Ile Lys Thr Lys Glu Val Lys Tyr Asp Ser Ser Leu Arg Pro Leu
35 40 45

Ser Ile Lys Tyr Asp Ala Asn Ser Ala Lys Ile Ile Ser Asn Ser Gly
50 55 60

His Ser Phe Ser Val Asp Phe Asp Asp Thr Glu Asp Lys Ser Val Leu
65 70 75 80

Arg Gly Gly Pro Leu Thr Gly Ser Tyr Arg Leu Arg Gln Phe His Leu
85 90 95

His Trp Gly Ser Ala Asp Asp His Gly Ser Glu His Val Val Asp Gly
100 105 110

Val Arg Tyr Ala Ala Glu Leu His Val Val His Trp Asn Ser Asp Lys
115 120 125

Tyr Pro Ser Phe Val Glu Ala Ala His Glu Pro Asp Gly Leu Ala Val
130 135 140

Leu Gly Val Phe Leu Gln Ile Gly Glu His Asn Ser Gln Leu Gln Lys
145 150 155 160

Ile Thr Asp Ile Leu Asp Ser Ile Lys Glu Lys Gly Lys Gln Thr Arg
165 170 175

Phe Thr Asn Phe Asp Pro Leu Ser Leu Leu Pro Pro Ser Trp Asp Tyr
180 185 190

Trp Thr Tyr Pro Gly Ser Leu Thr Val Pro Pro Leu Leu Glu Ser Val
195 200 205

Thr Trp Ile Val Leu Lys Gln Pro Ile Asn Ile Ser Ser Gln Gln Leu
210 215 220

Ala Thr Phe Arg Thr Leu Leu Cys Thr Ala Glu Gly Glu Ala Ala Ala
225 230 235 240

Phe Leu Leu Ser Asn His Arg Pro Pro Gln Pro Leu Lys Gly Arg Lys
245 250 255

Val Arg Ala Ser Phe His
260

<210> SEQ ID NO 64
<211> LENGTH: 252
<212> TYPE: PRT
<213> ORGANISM: Equus caballus

<400> SEQUENCE: 64

Met Ser Gly Pro Val His Trp Asn Glu Phe Phe Pro Ile Ala Asp Gly
1 5 10 15

Asp Gln Gln Ser Pro Ile Glu Ile Lys Thr Lys Glu Val Lys Tyr Asp
20 25 30

Ser Ser Leu Arg Pro Leu Thr Ile Lys Tyr Asp Pro Ser Ser Ala Lys
35 40 45

152

US 11,001,853 B2

153

154

-continued

Ile Ile Ser Asn Ser Gly His Ser Phe Ser Val Gly Phe Asp Asp Thr
 50 55 60

Glu Asn Lys Ser Val Leu Arg Gly Gly Pro Leu Thr Gly Ser Tyr Arg
 65 70 75 80

Leu Arg Gln Phe His Leu His Trp Gly Ser Ala Asp Asp His Gly Ser
 85 90 95

Glu His Val Val Asp Gly Val Arg Tyr Ala Ala Glu Leu His Ile Val
 100 105 110

His Trp Asn Ser Asp Lys Tyr Pro Ser Phe Val Glu Ala Ala His Glu
 115 120 125

Pro Asp Gly Leu Ala Val Leu Gly Val Phe Leu Gln Val Gly Glu His
 130 135 140

Asn Ser Gln Leu Gln Lys Ile Thr Asp Thr Leu Asp Ser Ile Lys Glu
 145 150 155 160

Lys Gly Lys Gln Thr Leu Phe Thr Asn Phe Asp Pro Leu Ser Leu Leu
 165 170 175

Pro Pro Ser Trp Asp Tyr Trp Thr Tyr Pro Gly Ser Leu Thr Val Pro
 180 185 190

Pro Leu Leu Glu Ser Val Thr Trp Ile Ile Leu Lys Gln Pro Ile Asn
 195 200 205

Ile Ser Ser Gln Gln Leu Val Lys Phe Arg Thr Leu Leu Cys Thr Ala
 210 215 220

Glu Gly Glu Thr Ala Ala Phe Leu Leu Ser Asn His Arg Pro Pro Gln
 225 230 235 240

Pro Leu Lys Gly Arg Lys Val Arg Ala Ser Phe Arg
 245 250

<210> SEQ ID NO 65

<211> LENGTH: 262

<212> TYPE: PRT

<213> ORGANISM: Bos taurus

<400> SEQUENCE: 65

Met Ser Gly Phe Ser Trp Gly Tyr Gly Glu Arg Asp Gly Pro Val His
 1 5 10 15

Trp Asn Glu Phe Phe Pro Ile Ala Asp Gly Asp Gln Gln Ser Pro Ile
 20 25 30

Glu Ile Lys Thr Lys Glu Val Arg Tyr Asp Ser Ser Leu Arg Pro Leu
 35 40 45

Gly Ile Lys Tyr Asp Ala Ser Ser Ala Lys Ile Ile Ser Asn Ser Gly
 50 55 60

His Ser Phe Asn Val Asp Phe Asp Asp Thr Asp Asp Lys Ser Val Leu
 65 70 75 80

Arg Gly Gly Pro Leu Thr Gly Ser Tyr Arg Leu Arg Gln Phe His Leu
 85 90 95

His Trp Gly Ser Thr Asp Asp His Gly Ser Glu His Val Val Asp Gly
 100 105 110

Val Arg Tyr Ala Ala Glu Leu His Val Val His Trp Asn Ser Asp Lys
 115 120 125

Tyr Pro Ser Phe Val Glu Ala Ala His Glu Pro Asp Gly Leu Ala Val
 130 135 140

Leu Gly Ile Phe Leu Gln Ile Gly Glu His Asn Pro Gln Leu Gln Lys
 145 150 155 160

Ile Thr Asp Ile Leu Asp Ser Ile Lys Glu Lys Gly Lys Gln Thr Arg

US 11,001,853 B2

155**156**

-continued

165	170	175
-----	-----	-----

Phe Thr Asn Phe Asp Pro Val Cys Leu Leu Pro Pro Cys Arg Asp Tyr		
180	185	190

Trp Thr Tyr Pro Gly Ser Leu Thr Val Pro Pro Leu Leu Glu Ser Val		
195	200	205

Thr Trp Ile Ile Leu Lys Gln Pro Ile Asn Ile Ser Ser Gln Gln Leu		
210	215	220

Ala Ala Phe Arg Thr Leu Leu Cys Ser Arg Glu Gly Glu Thr Ala Ala		
225	230	235

Phe Leu Leu Ser Asn His Arg Pro Pro Gln Pro Leu Lys Gly Arg Lys		
245	250	255

Val Arg Ala Ser Phe Arg		
260		

<210> SEQ_ID NO 66

<211> LENGTH: 419

<212> TYPE: PRT

<213> ORGANISM: Monodelphis domestica

<400> SEQUENCE: 66

Met Ala Ser Val Phe Ala Gly Trp Gly Pro Gly Arg Thr His Leu Phe		
1	5	10

Phe Arg Phe Phe Pro Gly Pro Phe Ser Ala Leu Pro Ala Gln Thr Ser		
20	25	30

Arg Gly Val Leu Val Phe Thr Ala Pro Gly Pro Ser Pro Arg Arg Val		
35	40	45

Pro Asp Pro Val His Pro Gly Arg Asp Val Val Arg Pro Ser Gly Ser		
50	55	60

Leu Phe Ser Cys Arg Leu Pro Pro Pro Arg Pro Ser Ala Pro Ala Arg		
65	70	75

Glu Arg Arg Pro Leu Ala Glu Lys Val Gly Arg Ser Ser Ala Pro His		
85	90	95

Leu Pro Leu Asp Asn Phe Glu Phe Ile Ala Lys Arg Leu Arg Arg Arg		
100	105	110

Val Leu Ser Gly Leu Ala Ala Glu Ser Ala Gly Ala Leu Ala Pro Ser		
115	120	125

Leu Pro Arg Ser Leu His Ser Ser Leu Gly Leu Arg Ser Ser Leu Lys		
130	135	140

Ser Gln Arg Val Phe Pro Ser Pro His Ser Glu Glu Thr Met Ser Arg		
145	150	155

Leu Ser Trp Gly Tyr Cys Glu His Asn Gly Pro Val His Trp Ser Glu		
165	170	175

Leu Phe Pro Ile Ala Asp Gly Asp Tyr Gln Ser Pro Ile Glu Ile Asn		
180	185	190

Thr Lys Glu Val Lys Tyr Asp Ser Ser Leu Arg Pro Leu Ser Ile Lys		
195	200	205

Tyr Asp Pro Ala Ser Ala Lys Ile Ile Ser Asn Ser Gly His Ser Phe		
210	215	220

Ser Val Asp Phe Asp Asp Ser Glu Asp Lys Ser Val Leu Arg Gly Gly		
225	230	240

Pro Leu Ile Gly Thr Tyr Arg Leu Arg Gln Phe His Leu His Trp Gly		
245	250	255

Ser Thr Asp Asp Gln Gly Ser Glu His Thr Val Asp Gly Met Lys Tyr		
260	265	270

US 11,001,853 B2

157

-continued

Ala Ala Glu Leu His Val Val His Trp Asn Ser Asp Lys Tyr Pro Ser
 275 280 285

Phe Val Glu Ala Ala His Glu Pro Asp Gly Leu Ala Val Leu Gly Ile
 290 295 300

Phe Leu Gln Thr Gly Glu His Asn Leu Gln Met Gln Lys Ile Thr Asp
 305 310 315 320

Ile Leu Asp Ser Ile Lys Glu Lys Gly Lys Gln Ile Arg Phe Thr Asn
 325 330 335

Phe Asp Pro Ala Thr Leu Leu Pro Gln Ser Trp Asp Tyr Trp Thr Tyr
 340 345 350

Pro Gly Ser Leu Thr Val Pro Pro Leu Leu Glu Ser Val Thr Trp Ile
 355 360 365

Val Leu Lys Gln Pro Ile Thr Ile Ser Ser Gln Gln Leu Ala Lys Phe
 370 375 380

Arg Ser Leu Leu Tyr Thr Gly Glu Gly Glu Ala Ala Ala Phe Leu Leu
 385 390 395 400

Ser Asn Tyr Arg Pro Pro Gln Pro Leu Lys Gly Arg Lys Val Arg Ala
 405 410 415

Ser Phe Arg

<210> SEQ ID NO 67

<211> LENGTH: 428

<212> TYPE: PRT

<213> ORGANISM: Ornithorhynchus anatinus

<400> SEQUENCE: 67

Met Lys Lys Gly Val Gly Ser Phe Tyr Glu Leu Ala Val Asn Arg Trp
 1 5 10 15

Ser Val Val Asn Arg Val Gln Ile Met Ile Val Glu Ser Ile Thr Glu
 20 25 30

Pro Leu Leu Cys Gly Ser Ala Leu Ala Val Ala Pro Ala Leu Ala Leu
 35 40 45

Ala Val Val Gln Ala Leu Ala Leu Thr Val Val Gln Ala Leu Ala Leu
 50 55 60

Ala Val Ser Pro Ala Leu Ala Leu Ser Val Ala Pro Ala Leu Ala Leu
 65 70 75 80

Ala Val Val Gln Ala Leu Ala Leu Ala Val Val Gln Ala Leu Ala Leu
 85 90 95

Ala Val Ala Gln Ala Leu Ala Leu Ala Val Ala Gln Ala Leu Ala Leu
 100 105 110

Ala Val Ala Gln Ala Leu Ala Leu Ala Pro Gln Ala Leu Ala Leu
 115 120 125

Thr Leu Pro Gln Ala Leu Ala Leu Thr Leu Ser Pro Thr Leu Ala Leu
 130 135 140

Ser Val Ala Pro Ala Leu Ala Leu Ala Val Ala Pro Ala Leu Ala Leu
 145 150 155 160

Ala Asp Ser Pro Ala Leu Ala Leu Ala Arg Pro His Pro Ser
 165 170 175

Ser Gly Pro Ile His Trp Asn Glu Leu Phe Pro Ile Ala Asp Gly Asp
 180 185 190

Arg Gln Ser Pro Ile Glu Ile Lys Thr Lys Glu Val Lys Tyr Asp Ser
 195 200 205

Ser Leu Arg Pro Leu Ser Ile Lys Tyr Asp Pro Thr Ser Ala Lys Ile
 210 215 220

158

US 11,001,853 B2

159**160**

-continued

Ile Ser Asn Ser Gly His Ser Phe Ser Val Asp Phe Asp Asp Thr Glu
225 230 235 240

Asp Lys Ser Val Leu Arg Gly Gly Pro Leu Ser Gly Thr Tyr Arg Leu
245 250 255

Arg Gln Phe His His Trp Gly Ser Ala Asp Asp His Gly Ser Glu
260 265 270

His Thr Val Asp Gly Met Glu Tyr Ser Ala Glu Leu His Val Val His
275 280 285

Trp Asn Ser Asp Lys Tyr Ser Ser Phe Val Glu Ala Ala His Glu Pro
290 295 300

Asp Gly Leu Ala Val Leu Gly Ile Phe Leu Lys Arg Gly Glu His Asn
305 310 315 320

Leu Gln Leu Gln Lys Ile Thr Asp Ile Leu Asp Ala Ile Lys Glu Lys
325 330 335

Gly Lys Gln Met Arg Phe Thr Asn Phe Asp Pro Leu Ser Leu Leu Pro
340 345 350

Leu Thr Arg Asp Tyr Trp Thr Tyr Pro Gly Ser Leu Thr Val Pro Pro
355 360 365

Leu Leu Glu Ser Val Ile Trp Ile Ile Phe Lys Gln Pro Ile Ser Ile
370 375 380

Ser Ser Gln Gln Leu Ala Lys Phe Arg Asn Leu Leu Tyr Thr Ala Glu
385 390 395 400

Gly Glu Ala Ala Asp Phe Met Leu Ser Asn His Arg Pro Pro Gln Pro
405 410 415

Leu Lys Gly Arg Lys Val Arg Ala Ser Phe Arg Ser
420 425

<210> SEQ ID NO: 68

<211> LENGTH: 1082

<212> TYPE: PRT

<213> ORGANISM: Chlamydomonas reinhardtii

<400> SEQUENCE: 68

Met Leu Pro Gly Leu Gly Val Ile Leu Leu Val Leu Pro Met Gln Tyr
1 5 10 15

Tyr Phe Gly Tyr Lys Ile Val Gln Ile Lys Leu Gln Asn Ala Lys His
20 25 30

Val Ala Leu Arg Ser Ala Ile Met Gln Glu Val Leu Pro Ala Ile Lys
35 40 45

Leu Val Lys Tyr Tyr Ala Trp Glu Gln Phe Phe Glu Asn Gln Ile Ser
50 55 60

Lys Val Arg Arg Glu Glu Ile Arg Leu Asn Phe Trp Asn Cys Val Met
65 70 75 80

Lys Val Ile Asn Val Ala Cys Val Phe Cys Val Pro Pro Met Thr Ala
85 90 95

Phe Val Ile Phe Thr Thr Tyr Glu Phe Gln Arg Ala Arg Leu Val Ser
100 105 110

Ser Val Ala Phe Thr Thr Leu Ser Leu Phe Asn Ile Leu Arg Phe Pro
115 120 125

Leu Val Val Leu Pro Lys Ala Leu Arg Ala Val Ser Glu Ala Asn Ala
130 135 140

Ser Leu Gln Arg Leu Glu Ala Tyr Leu Leu Glu Glu Val Pro Ser Gly
145 150 155 160

Thr Ala Ala Val Lys Thr Pro Lys Asn Ala Pro Pro Gly Ala Val Ile
165 170 175

US 11,001,853 B2

161

-continued

Glu Asn Gly Val Phe His His Pro Ser Asn Pro Asn Trp His Leu His
 180 185 190
 Val Pro Lys Phe Glu Val Lys Pro Gly Gln Val Val Ala Val Val Gly
 195 200 205
 Arg Ile Ala Ala Gly Lys Ser Ser Leu Val Gln Ala Ile Leu Gly Asn
 210 215 220
 Met Val Lys Glu His Gly Ser Phe Asn Val Gly Gly Arg Ile Ser Tyr
 225 230 235 240
 Val Pro Gln Asn Pro Trp Leu Gln Asn Leu Ser Leu Arg Asp Asn Val
 245 250 255
 Leu Phe Gly Glu Gln Phe Asp Glu Asn Lys Tyr Thr Asp Val Ile Glu
 260 265 270
 Ser Cys Ala Leu Thr Leu Asp Leu Gln Ile Leu Ser Asn Gly Asp Gln
 275 280 285
 Ser Lys Ala Gly Ile Arg Gly Val Asn Phe Ser Gly Gly Gln Arg Gln
 290 295 300
 Arg Val Asn Leu Ala Arg Cys Ala Tyr Ala Asp Ala Asp Leu Val Leu
 305 310 315 320
 Leu Asp Asn Ala Leu Ser Ala Val Asp His His Thr Ala His His Ile
 325 330 335
 Phe Asp Lys Cys Ile Lys Gly Leu Phe Ser Asp Lys Ala Val Val Leu
 340 345 350
 Val Thr His Gln Ile Glu Phe Met Pro Arg Cys Asp Asn Val Ala Ile
 355 360 365
 Met Asp Glu Gly Arg Cys Leu Tyr Phe Gly Lys Trp Asn Glu Glu Ala
 370 375 380
 Gln His Leu Leu Gly Lys Leu Leu Pro Ile Thr His Leu Leu His Ala
 385 390 395 400
 Ala Gly Ser Gln Glu Ala Pro Pro Ala Pro Lys Lys Lys Ala Glu Asp
 405 410 415
 Lys Ala Gly Pro Gln Lys Ser Gln Ser Leu Gln Leu Thr Leu Ala Pro
 420 425 430
 Thr Ser Ile Gly Lys Pro Thr Glu Lys Pro Lys Asp Val Gln Lys Leu
 435 440 445
 Thr Ala Tyr Gln Ala Ala Leu Ile Tyr Thr Trp Tyr Gly Asn Leu Phe
 450 455 460
 Leu Val Gly Val Cys Phe Phe Phe Leu Ala Ala Gln Cys Ser Arg
 465 470 475 480
 Gln Ile Ser Asp Phe Trp Val Arg Trp Trp Val Asn Asp Glu Tyr Lys
 485 490 495
 Lys Phe Pro Val Lys Gly Glu Gln Asp Ser Ala Ala Thr Thr Phe Tyr
 500 505 510
 Cys Leu Ile Tyr Leu Leu Leu Val Gly Leu Phe Tyr Ile Phe Met Ile
 515 520 525
 Phe Arg Gly Ala Thr Phe Leu Trp Trp Val Leu Lys Ser Ser Glu Thr
 530 535 540
 Ile Arg Arg Lys Ala Leu His Asn Val Leu Asn Ala Pro Met Gly Phe
 545 550 555 560
 Phe Leu Val Thr Pro Val Gly Asp Leu Leu Leu Asn Phe Thr Lys Asp
 565 570 575
 Gln Asp Ile Met Asp Glu Asn Leu Pro Asp Ala Val His Phe Met Gly
 580 585 590

162

US 11,001,853 B2

163**164**

-continued

Ile Tyr Gly Leu Ile Leu Leu Ala Thr Thr Ile Thr Val Ser Val Thr
595 600 605

Ile Asn Phe Phe Ala Ala Phe Thr Gly Ala Leu Ile Ile Met Thr Leu
610 615 620

Ile Met Leu Ser Ile Tyr Leu Pro Ala Ala Thr Ala Leu Lys Lys Ala
625 630 635 640

Arg Ala Val Ser Gly Gly Met Leu Val Gly Leu Val Ala Glu Val Leu
645 650 655

Glu Gly Leu Gly Val Val Gln Ala Phe Asn Lys Gln Glu Tyr Phe Ile
660 665 670

Glu Glu Ala Ala Arg Arg Thr Asn Ile Thr Asn Ser Ala Val Phe Asn
675 680 685

Ala Glu Ala Leu Asn Leu Trp Leu Ala Phe Trp Cys Asp Phe Ile Gly
690 695 700

Ala Cys Leu Val Gly Val Val Ser Ala Phe Ala Val Gly Met Ala Lys
705 710 715 720

Asp Leu Gly Gly Ala Thr Val Gly Leu Ala Phe Ser Asn Ile Ile Gln
725 730 735

Met Leu Val Phe Tyr Thr Trp Val Val Arg Phe Ile Ser Glu Ser Ile
740 745 750

Ser Leu Phe Asn Ser Val Glu Gly Met Ala Tyr Leu Ala Asp Tyr Val
755 760 765

Pro His Asp Gly Val Phe Tyr Asp Gln Arg Gln Lys Asp Gly Val Ala
770 775 780

Lys Gln Ile Val Leu Pro Asp Gly Asn Ile Val Pro Ala Ala Ser Lys
785 790 795 800

Val Gln Val Val Asp Asp Ala Ala Leu Ala Arg Trp Pro Ala Thr
805 810 815

Gly Asn Ile Arg Phe Glu Asp Val Trp Met Gln Tyr Arg Leu Asp Ala
820 825 830

Pro Trp Ala Leu Lys Gly Val Thr Phe Lys Ile Asn Asp Gly Glu Lys
835 840 845

Val Gly Ala Val Gly Arg Thr Gly Ser Gly Lys Ser Thr Thr Leu Leu
850 855 860

Ala Leu Tyr Arg Met Phe Glu Leu Gly Lys Gly Arg Ile Leu Val Asp
865 870 875 880

Gly Val Asp Ile Ala Thr Leu Ser Leu Lys Arg Leu Arg Thr Gly Leu
885 890 895

Ser Ile Ile Pro Gln Glu Pro Val Met Phe Thr Gly Thr Val Arg Ser
900 905 910

Asn Leu Asp Pro Phe Gly Glu Phe Lys Asp Asp Ala Ile Leu Trp Glu
915 920 925

Val Leu Lys Lys Val Gly Leu Glu Asp Gln Ala Gln His Ala Gly Gly
930 935 940

Leu Asp Gly Gln Val Asp Gly Thr Gly Gly Lys Ala Trp Ser Leu Gly
945 950 955 960

Gln Met Gln Leu Val Cys Leu Ala Arg Ala Ala Leu Arg Ala Val Pro
965 970 975

Ile Leu Cys Leu Asp Glu Ala Thr Ala Ala Met Asp Pro His Thr Glu
980 985 990

Ala Ile Val Gln Gln Thr Ile Lys Lys Val Phe Asp Asp Arg Thr Thr
995 1000 1005

Ile Thr Ile Ala His Arg Leu Asp Thr Ile Ile Glu Ser Asp Lys

US 11,001,853 B2

165

-continued

1010	1015	1020
Ile Ile Val Met Glu Gln Gly Ser Leu Met Glu Tyr Glu Ser Pro		
1025	1030	1035
Ser Lys Leu Leu Ala Asn Arg Asp Ser Met Phe Ser Lys Leu Val		
1040	1045	1050
Asp Lys Thr Gly Pro Ala Ala Ala Ala Leu Arg Lys Met Ala		
1055	1060	1065
Glu Asp Phe Trp Ser Thr Arg Ser Ala Gln Gly Arg Asn Gln		
1070	1075	1080

<210> SEQ_ID NO 69
<211> LENGTH: 1321
<212> TYPE: PRT
<213> ORGANISM: Volvox carteri
<400> SEQUENCE: 69

Met Gly Thr Ile Ser His Pro Ala Arg Gly Asn Asp Pro Thr Ala Gly			
1	5	10	15
Phe Phe Asn Lys Phe Ala Phe Gly Trp Met Phe Lys His Val Ser Glu			
20	25	30	
Ala Arg Lys Asn Gly Asp Ile Asp Leu Asp Lys Met Gly Met Pro Pro			
35	40	45	
Glu Asn His Ala His Glu Ala Tyr Asp Met Phe Ala Ser Asn Trp Ala			
50	55	60	
Ala Glu Met Lys Leu Lys Asp Ser Gly Ala Lys Pro Ser Leu Val Arg			
65	70	75	80
Ala Leu Arg Lys Ser Phe Gly Leu Val Tyr Leu Leu Gly Gly Val Phe			
85	90	95	
Lys Cys Phe Trp Ser Thr Phe Val Ile Thr Gly Ala Phe Tyr Phe Val			
100	105	110	
Arg Ser Leu Leu Ala His Val Asn Gly Ile Lys Asp Gly Arg Leu Tyr			
115	120	125	
Ser Lys Thr Val Ser Gly Trp Cys Leu Met Ala Gly Phe Thr Leu Asp			
130	135	140	
Ala Trp Leu Leu Gly Leu Ser Leu Gln Arg Met Gly Tyr Ile Cys Met			
145	150	155	160
Ser Val Gly Ile Arg Ala Arg Ala Ala Leu Val Gln Ala Val Thr His			
165	170	175	
Lys Ala Phe Arg Leu Ser Ser Val Arg Ala Asp Gln Ser Ala Ala Ile			
180	185	190	
Val Asn Phe Val Ser Ser Asp Ile Gln Lys Ile Tyr Asp Gly Ala Leu			
195	200	205	
Glu Phe His Tyr Leu Trp Thr Ala Pro Phe Glu Ala Ala Ala Ile Leu			
210	215	220	
Ala Leu Leu Gly Tyr Leu Thr Asn Asp Ser Met Leu Pro Gly Leu Gly			
225	230	235	240
Val Ile Leu Leu Val Leu Pro Leu Gln Tyr Phe Phe Gly Tyr Lys Ile			
245	250	255	
Ile Gln Ile Lys Leu Gln Asn Ala Lys His Val Ala Leu Arg Ser Ser			
260	265	270	
Ile Leu Gln Glu Val Leu Pro Ala Ile Lys Leu Val Lys Tyr Tyr Ala			
275	280	285	
Trp Glu Gln Phe Phe Glu Asp Glu Ile Ser Lys Ile Arg Arg Glu Glu			
290	295	300	

166

US 11,001,853 B2

167

-continued

Met Arg Leu Ser Phe Trp Asn Ala Met Met Lys Val Ile Asn Val Ala
 305 310 315 320
 Cys Val Phe Cys Val Pro Pro Met Thr Ala Phe Val Ile Phe Thr Thr
 325 330 335
 Tyr Glu Phe Gln Lys Ala Arg Leu Val Ser Gly Val Ala Phe Thr Thr
 340 345 350
 Leu Ser Leu Phe Asn Ile Leu Arg Phe Pro Leu Val Val Leu Pro Lys
 355 360 365
 Ala Leu Arg Ala Val Ser Glu Ala His Ala Ser Leu Gln Arg Leu Glu
 370 375 380
 Ser Tyr Leu Leu Glu Asp Val Pro Gln Gly Thr Ala Ser Gly Gly Lys
 385 390 395 400
 Ser Ser Lys Ser Ser Ala Pro Gly Val His Ile Asp Asn Ala Val Tyr
 405 410 415
 His His Pro Ser Asn Pro Asn Trp His Leu His Val Pro Arg Phe Asp
 420 425 430
 Val Arg Pro Gly Gln Val Val Ala Val Val Gly Arg Ile Gly Ala Gly
 435 440 445
 Lys Ser Ser Leu Val Gln Ala Ile Leu Gly Asn Met Val Lys Glu His
 450 455 460
 Gly Ser Gln Gln Val Gly Gly Arg Ile Ser Tyr Val Pro Gln Asn Pro
 465 470 475 480
 Trp Leu Gln Asn Leu Ser Ile Arg Asp Asn Val Thr Phe Gly Glu Gly
 485 490 495
 Trp Asp Glu Asn Lys Tyr Glu Ala Val Ile Asp Ala Cys Ala Leu Thr
 500 505 510
 Met Asp Leu Gln Ile Leu Pro Gln Gly Asp Gln Ser Lys Ala Gly Ile
 515 520 525
 Arg Gly Val Asn Phe Ser Gly Gly Gln Arg Gln Arg Val Asn Leu Ala
 530 535 540
 Arg Cys Ala Tyr Ala Asp Ala Asp Leu Val Leu Leu Asp Asn Ala Leu
 545 550 555 560
 Ser Ala Val Asp His His Thr Ala His His Ile Phe Asp Lys Cys Ile
 565 570 575
 Lys Gly Leu Phe Ser Asp Lys Ala Val Val Leu Ile Thr His Gln Ile
 580 585 590
 Glu Phe Met Pro Arg Cys Asp Ala Val Ala Ile Met Asp Glu Gly Arg
 595 600 605
 Cys Leu Tyr Phe Gly Lys Trp Asn Glu Glu Ser Gln His Leu Leu Gly
 610 615 620
 Lys Leu Leu Pro Ile Thr His Leu Leu His Ala Ala Gly Ser Gln Glu
 625 630 635 640
 Ala Pro Pro Ala Ala Pro Lys Lys Asp Asp Lys Ala Thr Pro Gln
 645 650 655
 Lys Ser Gln Ser Leu Gln Leu Thr Leu Ala Pro Thr Ser Ile Gly Lys
 660 665 670
 Pro Thr Gln Lys Asp Thr Lys Ala Ala Pro Lys Leu Thr Ala Phe Lys
 675 680 685
 Ala Ala Leu Ile Tyr Thr Tyr Gly Asn Ile Leu Leu Val Phe Val
 690 695 700
 Cys Phe Ile Thr Phe Leu Ala Ala Gln Thr Cys Arg Gln Met Ser Asp
 705 710 715 720
 Phe Trp Val Arg Trp Trp Val Asn Asp Glu Tyr Lys His Phe Pro Lys

168

US 11,001,853 B2

169**170**

-continued

725	730	735
Arg Thr Gly Val Arg Glu Glu Ser Ala Thr Lys Phe Tyr Ala Leu Ile		
740	745	750
Tyr Leu Leu Leu Val Gly Leu Phe Tyr Phe Thr Met Val Ala Arg Gly		
755	760	765
Ser Thr Phe Leu Trp Trp Val Leu Arg Ser Ser Glu Asn Ile Arg Lys		
770	775	780
Lys Ala Leu Asn Asn Val Leu Asn Ala Pro Met Gly Phe Phe Leu Val		
785	790	795
800		
Thr Pro Val Gly Asp Leu Leu Leu Asn Phe Thr Lys Asp Gln Asp Ile		
805	810	815
Met Asp Glu Asn Leu Pro Asp Ala Ile His Phe Met Gly Ile Tyr Gly		
820	825	830
Leu Ile Leu Ala Thr Thr Ile Thr Val Ser Val Thr Ile Asn Phe		
835	840	845
Phe Gly Ala Phe Thr Gly Phe Leu Ile Ile Met Thr Leu Ile Met Leu		
850	855	860
Ala Ile Tyr Leu Pro Ala Ala Thr Ala Leu Lys Lys Ala Arg Ala Val		
865	870	875
880		
Ser Gly Gly Gln Leu Val Gly Leu Val Ala Glu Val Leu Glu Gly Leu		
885	890	895
Asn Val Val Gln Ala Phe Ser Lys Gln Glu Tyr Phe Ile Glu Glu Ala		
900	905	910
Ala Arg Arg Thr Asp Val Thr Asn Ala Ala Val Phe Asn Ala Glu Ser		
915	920	925
Leu Asn Leu Trp Leu Ala Phe Trp Cys Asp Leu Ile Gly Ala Ser Leu		
930	935	940
Val Gly Val Val Ser Ala Phe Ala Val Gly Leu Lys Asp Gln Leu Gly		
945	950	955
960		
Ala Ala Thr Val Gly Leu Ala Phe Ser Asn Ile Ile Gln Met Leu Val		
965	970	975
Phe Tyr Thr Trp Val Val Arg Phe Ile Ala Glu Ser Ile Ser Leu Phe		
980	985	990
Asn Ser Val Glu Ala Met Ala Trp Leu Ala Asp Tyr Val Pro Lys Asp		
995	1000	1005
Gly Ile Phe Tyr Asp Gln Lys Gln Leu Asp Gly Val Ala Lys Ser		
1010	1015	1020
Ile Thr Leu Pro Asp Gly Gln Ile Val Pro Ala Thr Ser Lys Val		
1025	1030	1035
Gln Val Val Val Asp Asp Ala Ala Leu Ala Arg Trp Pro Ala Thr		
1040	1045	1050
Gly Asn Ile Arg Phe Glu Asp Val Trp Met Gln Tyr Arg Leu Asp		
1055	1060	1065
Ala Ala Trp Ala Leu Lys Gly Val Thr Phe Lys Ile Asn Asp Gly		
1070	1075	1080
Glu Lys Val Gly Ala Val Gly Arg Thr Gly Ser Gly Lys Ser Thr		
1085	1090	1095
Thr Leu Leu Ala Leu Tyr Arg Met Phe Glu Leu Gly Lys Gly Arg		
1100	1105	1110
Ile Leu Ile Asp Gly Val Asp Ile Ala Thr Leu Ser Leu Lys Arg		
1115	1120	1125
Leu Arg Thr Gly Leu Ser Ile Ile Pro Gln Glu Pro Val Met Phe		
1130	1135	1140

US 11,001,853 B2

171

172

-continued

Thr Gly Thr Val Arg Ser Asn Leu Asp Pro Phe Gly Glu Phe Lys
 1145 1150 1155
 Asp Asp Ser Val Leu Trp Glu Val Leu Gln Lys Val Gly Leu Glu
 1160 1165 1170
 Ala Gln Ala Gln His Ala Gly Gly Leu Asp Gly Arg Val Asp Gly
 1175 1180 1185
 Thr Gly Gly Lys Ala Trp Ser Leu Gly Gln Met Gln Leu Val Cys
 1190 1195 1200
 Leu Ala Arg Ala Ala Leu Arg Ala Val Pro Ile Leu Cys Leu Asp
 1205 1210 1215
 Glu Ala Thr Ala Ala Met Asp Pro His Thr Glu Gln Val Val Gln
 1220 1225 1230
 Glu Thr Ile Lys Lys Val Phe Asp Asp Arg Thr Thr Ile Thr Ile
 1235 1240 1245
 Ala His Arg Leu Asp Thr Ile Ile Glu Ser Asp Lys Val Leu Val
 1250 1255 1260
 Met Glu Ala Gly Glu Leu Lys Glu Phe Ala Pro Pro Ala Gln Leu
 1265 1270 1275
 Leu Ala Asn Arg Glu Thr Met Phe Ser Lys Leu Val Asp Lys Thr
 1280 1285 1290
 Gly Pro Ala Ala Ala Ala Leu Arg Lys Met Ala Asp Glu His
 1295 1300 1305
 Phe Ser Lys Ser Gln Ala Arg Ala Ala Ala Gln Arg His
 1310 1315 1320

<210> SEQ ID NO 70

<211> LENGTH: 2297

<212> TYPE: PRT

<213> ORGANISM: Chlorella variabilis

<400> SEQUENCE: 70

Met Val Pro Leu Leu Ala Gln Arg Gly Arg Ile Arg Ser Gln Ala Pro
 1 5 10 15
 Arg Thr Trp His Pro Asp Pro Gln Pro Leu His Ala Glu Arg Ser Arg
 20 25 30
 Gln Cys Pro Gly Arg Gly Val Arg Ala Ala Ala Lys Arg Gly Gly Gly
 35 40 45
 Ser Gly Gly Ala Thr His Lys Ser Lys Ser Lys Glu Leu Asp Glu
 50 55 60
 Val Ala Ala Phe Glu Gln Leu Met Cys Asp Trp Asp Asp Ala Phe Ala
 65 70 75 80
 Ala Asp Cys Tyr Asp Asn Glu Arg Ala Ala Arg Met Ala Arg Leu Ala
 85 90 95
 Glu Glu Gly Tyr Gln His His Gly Arg Gly Phe Val Phe Val Arg Ser
 100 105 110
 Arg Leu Asp Lys Arg Ser Arg Lys Ala Arg Asn Asp Ser Gly Ala Ser
 115 120 125
 Lys Gly Phe Gly Ala Ala Ala Lys Ala Leu Ser Val Glu Gln Gly Thr
 130 135 140
 Pro Leu Glu Asn Asn Pro Gln Leu His Leu Leu Ser Trp Thr Ala Cys
 145 150 155 160
 Tyr Ile Ala Ser Ser Gln Leu Asp Ser Leu Gly Gly Leu Phe Ser Thr
 165 170 175
 Gln Glu Gly Val Leu Leu Pro Asp Ser Gly Ser Leu Leu Thr Asp Gly

US 11,001,853 B2

173**174**

-continued

180	185	190
Gly Ser Gly Ala Ser Gly Ser Asn Ala Ala Asp Ala Val	Gly Glu Leu	
195	200	205
Gln Arg Val Leu Arg Gly Gln Asp Leu Ser Gln Leu Arg	Gly Tyr Val	
210	215	220
Gly Ala Pro Pro Gln Ala Arg Pro Ala Ser Gly Ser Asp	Asp Asp Gly	
225	230	235
Ser Ser Thr Thr Gly Ser Asn Asn Gly Ala Ala Gly	Glu Gly Ser Glu	
245	250	255
Val Glu Glu Gly Thr Ala Met Gly Gly Ile Arg Arg	Tyr Glu Pro Glu	
260	265	270
Ser Gly Glu Leu Val Val Leu Leu Ser Cys Lys Ile Gly	Gly Lys Pro	
275	280	285
Ala Val Gly Ala Glu Leu Leu Ala Val Ala Gln Ala	Glu Asp Gly Lys	
290	295	300
His Ala Pro Gly Ala Ser Pro Asp Thr Arg Leu Cys	Lys Glu Pro Ser	
305	310	315
Gln Ser Ala Phe Asp Leu Trp Ser Phe Gly Trp Met Asn	Lys Ile Val	
325	330	335
Pro Ala Ala Arg Arg Gly Glu Val Glu Val Ala Asp	Leu Pro Leu Pro	
340	345	350
Glu Ala Gln Gln Ala Glu Pro Cys Tyr Glu Glu Leu	Asn Thr Asn Trp	
355	360	365
Glu Ala Ala Val Gln Glu Ala Lys Lys Ala Gly	Lys Glu Pro Lys Leu	
370	375	380
Met Lys Val Leu Trp Lys Thr Tyr Gly Lys Asp Ile	Val Leu Ala Gly	
385	390	395
Ile Phe Lys Leu Met Trp Ser Val Phe Val Ile Leu	Gly Ala Tyr Tyr	
405	410	415
Phe Thr Arg Ser Ile Leu Met Cys Ile Arg Thr Leu	Glu Gly Lys Asp	
420	425	430
Asp Ser Ile Tyr Asp Thr Glu Trp Lys Gly Trp Val	Leu Thr Gly Phe	
435	440	445
Phe Phe Leu Asp Ala Trp Leu Leu Gly Met Met	Leu Gln Arg Met Ala	
450	455	460
Phe Asn Cys Leu Lys Val Gly Ile Lys Ala Arg Ala	Ala Leu Thr Thr	
465	470	475
Met Ile Ala Arg Lys Cys Tyr Asn Met Ala His Leu	Thr Lys Asp Thr	
485	490	495
Ala Ala Glu Ala Val Gly Phe Val Ala Ser Asp Ile	Asn Lys Val Phe	
500	505	510
Glu Gly Ile Gln Glu Val His Tyr Leu Trp Gly Ala	Pro Val Glu Ala	
515	520	525
Gly Ala Ile Leu Ala Leu Leu Gly Thr Leu Val	Gly Val Tyr Cys Ile	
530	535	540
Gly Gly Val Ile Ile Val Cys Met Val Val Pro	Leu Gln Tyr Tyr Phe	
545	550	555
Gly Tyr Lys Ile Ile Lys Asn Lys Ile Lys Asn Ala	Pro Asn Val Thr	
565	570	575
Glu Arg Trp Ser Ile Ile Gln Glu Ile Leu Pro Ala	Met Lys Leu Val	
580	585	590
Lys Tyr Tyr Ala Trp Glu Arg Phe Phe Glu Lys His	Val Ala Asp Met	
595	600	605

US 11,001,853 B2

175**176**

-continued

Arg Thr Arg Glu Arg His Tyr Met Phe Trp Asn Ala Val Val Lys Thr
 610 615 620
 Val Asn Val Thr Met Val Phe Gly Val Pro Pro Met Val Thr Phe Ala
 625 630 635 640
 Val Leu Val Pro Tyr Glu Leu Trp His Val Asp Ser Ser Thr Ser Glu
 645 650 655
 Pro Tyr Ile Lys Pro Gln Thr Ala Phe Thr Met Leu Ser Leu Phe Asn
 660 665 670
 Val Leu Arg Phe Pro Leu Val Val Leu Pro Lys Ala Met Arg Cys Val
 675 680 685
 Ser Glu Ala Leu Arg Ser Val Gly Asn Leu Glu Lys Phe Leu Ala Glu
 690 695 700
 Pro Val Ala Pro Arg Gln Asp Leu Glu Gly Lys Pro Gly Ala Gln Leu
 705 710 715 720
 Ser Lys Ala Val Leu Arg His Glu Met Asp Thr Ser Gly Phe Thr Leu
 725 730 735
 Arg Val Pro Glu Phe Ser Val Lys Ala Gly Glu Leu Val Ala Val Val
 740 745 750
 Gly Arg Val Gly Ala Gly Lys Ser Ser Ile Leu Gln Ala Met Leu Gly
 755 760 765
 Asn Met Gln Thr Ala Ser Gly Leu Ala Lys Cys Gln His Ser Ala Ser
 770 775 780
 Ser Cys Leu Pro Phe Leu Val Glu Gly Thr Ala His Ser Gly Gly Arg
 785 790 795 800
 Ile Ala Tyr Val Pro Gln Thr Ala Trp Cys Gln Asn Leu Ser Leu Arg
 805 810 815
 Asp Asn Ile Thr Phe Gly Gln Pro Trp Asp Glu Ala Lys Tyr Lys Gln
 820 825 830
 Val Ile His Ala Cys Ala Leu Glu Leu Asp Leu Ala Ile Leu Ala Ala
 835 840 845
 Gly Asp Gln Ser Lys Ala Gly Leu Arg Gly Ile Asn Leu Ser Gly Gly
 850 855 860
 Gln Arg Gln Arg Leu Asn Leu Ala Arg Cys Ala Tyr Phe Asp Gly Asp
 865 870 875 880
 Leu Val Leu Leu Asp Asn Ala Leu Ser Ala Val Asp His His Thr Ala
 885 890 895
 His His Ile Phe Glu His Cys Val Arg Gly Met Phe Arg Asp Lys Ala
 900 905 910
 Thr Val Leu Val Thr His Gln Val Glu Phe Leu Pro Gln Cys Asp Lys
 915 920 925
 Val Ala Ile Met Asp Asp Gly Thr Cys Val Tyr Phe Gly Pro Trp Asn
 930 935 940
 Ala Ala Ala Gln Gln Leu Leu Ser Lys Tyr Leu Pro Ala Ser His Leu
 945 950 955 960
 Leu Ala Ala Gly Gly Asn Ala Glu Gln Pro Arg Asp Thr Lys Lys Lys
 965 970 975
 Val Val Lys Lys Glu Glu Thr Lys Lys Thr Glu Asp Ala Gly Lys Ala
 980 985 990
 Lys Arg Val His Ser Ala Ser Leu Thr Leu Lys Ser Ala Leu Trp Glu
 995 1000 1005
 Tyr Cys Trp Asp Ala Arg Trp Ile Ile Phe Cys Leu Ser Leu Phe
 1010 1015 1020

US 11,001,853 B2

177

178

-continued

Phe Phe Leu Thr Ala Gln Ala Ser Arg Gln Leu Ala Asp Tyr Phe
1025 1030 1035

Ile Arg Trp Trp Thr Arg Asp His Tyr Asn Lys Tyr Gly Val Leu
1040 1045 1050

Cys Ile Asp Glu Gly Asp Asn Pro Cys Gly Pro Leu Phe Tyr Val
1055 1060 1065

Gln Tyr Tyr Gly Ile Leu Gly Leu Leu Cys Phe Ile Val Leu Met
1070 1075 1080

Ala Phe Arg Gly Ala Phe Leu Tyr Thr Trp Ser Leu Gly Ala Ser
1085 1090 1095

Tyr Arg Gln His Glu Lys Ser Ile His Arg Val Leu Tyr Ala Pro
1100 1105 1110

Leu Gly Phe Phe Leu Thr Thr Pro Val Gly Asp Leu Leu Val Ser
1115 1120 1125

Phe Thr Lys Asp Gln Asp Val Met Asp Asp Ala Leu Pro Asp Ala
1130 1135 1140

Leu Tyr Tyr Ala Gly Ile Tyr Gly Leu Ile Leu Leu Ala Thr Ala
1145 1150 1155

Ile Thr Val Ser Val Thr Ile Pro Leu Phe Ser Ala Leu Ala Gly
1160 1165 1170

Gly Leu Phe Val Val Ser Gly Ile Met Leu Ala Ile Tyr Leu Pro
1175 1180 1185

Ala Ala Thr His Leu Lys Lys Leu Arg Met Gly Thr Ser Gly Asp
1190 1195 1200

Val Val Thr Leu Ile Ala Glu Ala Leu Asp Gly Leu Gly Val Ile
1205 1210 1215

Gln Ala Tyr Gly Lys Gln Ala Tyr Phe Thr Thr Ile Thr Ser Gln
1220 1225 1230

Tyr Val Asn Asp Ala His Arg Ala Leu Phe Gly Ala Glu Ser Leu
1235 1240 1245

Asn Leu Trp Leu Ala Phe Ile Cys Asp Phe Phe Gly Ala Cys Met
1250 1255 1260

Val Leu Ser Val Ala Cys Phe Gly Ile Gly Gln Trp Ser Thr Leu
1265 1270 1275

Gly Ser Ser Ser Val Gly Leu Ala Phe Ser Gln Ser Ile Gln Met
1280 1285 1290

Leu Val Phe Tyr Thr Trp Ser Ile Arg Leu Val Ala Glu Cys Ile
1295 1300 1305

Gly Leu Phe Gly Ser Ala Glu Lys Ile Ala Trp Leu Ala Asn His
1310 1315 1320

Thr Pro Gln Glu Ala Gly Ser Leu Asp Pro Pro Ser Leu Pro Gly
1325 1330 1335

Ser Gly Glu Thr Lys Ala Ala Pro Lys Lys Arg Gly Thr Ala Gly
1340 1345 1350

Lys Phe Leu Pro Pro Leu Lys Asp Glu Asp Leu Ala Ile Val Pro
1355 1360 1365

Thr Gly Gly Pro Lys Leu Pro Ser Gly Trp Pro Arg Thr Gly Val
1370 1375 1380

Leu Glu Phe Asn Gln Val Val Met Lys Tyr Ala Pro His Leu Pro
1385 1390 1395

Pro Ala Leu Arg Gly Val Ser Phe Lys Val Lys Ser Gly Asp Lys
1400 1405 1410

Val Gly Val Val Gly Arg Thr Gly Ser Gly Lys Ser Thr Leu Leu

US 11,001,853 B2

179**180**

-continued

1415	1420	1425
Leu Ala	Leu Tyr Arg Met Phe Asn Leu Glu Ser Gly Ala Ile Thr	
1430	1435	1440
Leu Asp	Gly Ile Asp Ile Ser Thr Leu Thr Leu Glu Gln Leu Arg	
1445	1450	1455
Arg Gly	Leu Ser Val Ile Pro Gln Glu Pro Thr Val Phe Ser Gly	
1460	1465	1470
Thr Val	Arg Thr Asn Leu Asp Pro Phe Gly Glu Phe Gly Ala Asp	
1475	1480	1485
Ala Ile	Leu Trp Glu Ala Leu Arg Asp Cys Gly Leu Glu Glu Gln	
1490	1495	1500
Val Lys	Ala Cys Gly Gly Leu Asp Ala Lys Leu Asp Gly Thr Gly	
1505	1510	1515
Gly Asn	Ala Trp Ser Ile Gly Gln Gln Gln Leu Met Cys Leu Ala	
1520	1525	1530
Arg Ala	Ala Leu Lys Lys Val Pro Val Leu Cys Leu Asp Glu Ala	
1535	1540	1545
Thr Ala	Ala Met Asp Pro His Thr Glu Ala His Val Leu Glu Ile	
1550	1555	1560
Ile Glu	Arg Ile Phe Ser Asp Arg Thr Met Leu Thr Ile Ala His	
1565	1570	1575
Arg Leu	Asp Asn Val Ile Arg Ser Asp Leu Val Val Val Met Asp	
1580	1585	1590
Ala Gly	Gln Val Cys Glu Met Gly Thr Pro Asp Glu Leu Leu Ala	
1595	1600	1605
Asn Pro	Gln Ser Ala Phe Ser Gln Leu Val Asp Lys Thr Gly Ala	
1610	1615	1620
Ala Ser	Ala Ala Ala Leu Arg Lys Met Ala Ala Asp Phe Leu Asp	
1625	1630	1635
Glu Arg	Ala Arg Gly Gln Lys Leu Gly Phe Lys Pro Arg Pro Ser	
1640	1645	1650
Leu Glu	Glu Ser His Ile Cys Val Ala Pro Ser Pro Ser Leu Ile	
1655	1660	1665
Leu Ser	Thr Leu Leu Phe Pro Pro Ala Phe Met Ala Asn Val Thr	
1670	1675	1680
Ala Leu	Leu Leu Pro Lys Pro Val Leu Ser His Ala Pro Val Ser	
1685	1690	1695
Ser Gln	Thr Val Asn Thr Tyr Ile Arg Leu Asn Ile Ile Gln Leu	
1700	1705	1710
Gln Cys	Asn Val Leu His Pro Ala Thr Lys Glu Ala Thr Trp Ser	
1715	1720	1725
Ser Arg	Arg Ile Thr Phe Thr Ala His Leu Ser Ser Ser Gly Ser	
1730	1735	1740
Lys Pro	Pro Pro Pro Leu Pro Pro Leu Thr Glu Leu Pro Glu Gly	
1745	1750	1755
Arg Gly	Leu Asp Trp Ser Ser Ala Gly Tyr Arg Asp Gly Arg Glu	
1760	1765	1770
Ala Ile	Pro Ser Pro Ser Ala Lys Tyr Ser Ala Ala Asp Tyr Gly	
1775	1780	1785
Ala Ala	Gly Asp Gly Val Thr Asp Asp Thr Gln Ala Leu Gln Val	
1790	1795	1800
Ala Val	Ala Ala Ala His Glu Asp Asp Glu Gly Gly Val Val Tyr	
1805	1810	1815

US 11,001,853 B2

181**182**

-continued

Leu Gly Ala Gly Thr Phe Val Leu Thr Gln Pro Leu Ser Ile Ala
 1820 1825 1830
 Gly Ser Asn Val Val Ile Arg Gly Ala Gly Glu Asp Ala Thr Thr
 1835 1840 1845
 Ile Phe Val Pro Leu Pro Leu Ser Asp Val Phe Pro Gly Thr Trp
 1850 1855 1860
 Ser Met Asp Ala Ser Gly Lys Val Thr Ser Pro Trp Ile Thr Arg
 1865 1870 1875
 Gly Gly Phe Leu Ala Phe Ser Gly Arg Arg Thr Lys Ser Ser Asp
 1880 1885 1890
 Ser Ser Thr Leu Leu Ala Thr Val Ala Gly Ser Val Glu Gln Gly
 1895 1900 1905
 Ala Ser Val Ile Pro Val Asp Ser Thr Ala Glu Phe Arg Leu Gly
 1910 1915 1920
 Gln Trp Val Arg Ile Ile Asn Asp Ala Ser Thr Asp Ala Ser
 1925 1930 1935
 Ala Gly Gly Gly Thr Leu Glu Arg Gly Ser Ser Glu Val Gln Glu
 1940 1945 1950
 Ser Glu Thr Met Ile Ala Glu Gly Ala Thr Gly Gly Ala Gly
 1955 1960 1965
 Val Arg Ala Gln Trp Thr Gly Val Leu His Ala Phe Glu Pro Thr
 1970 1975 1980
 Val Gln Cys Ser Gly Val Glu Gln Leu Thr Ile Arg Phe Asn His
 1985 1990 1995
 Ser Met Met Ala Ala His Leu Ala Glu Arg Gly Tyr Asn Ala Ile
 2000 2005 2010
 Glu Leu Glu Asp Val Val Asp Cys Trp Ile Arg Gln Val Thr Ile
 2015 2020 2025
 Leu Asn Ala Asp Asn Ala Ile Arg Leu Arg Gly Thr Asp His Ser
 2030 2035 2040
 Thr Leu Ser Gly Gln Ala Cys Ser Gly Gly Val Val Ala Val
 2045 2050 2055
 Val Pro Val Trp Cys Arg Arg Gly Leu Pro Ser Pro Ala Asp Val
 2060 2065 2070
 Thr Val Gly Val Thr Glu Leu Arg Trp Glu Pro Asp Thr Arg Glu
 2075 2080 2085
 Val Asn Gly His His Ala Ile Thr Val Ser Lys Gly His Ala Asn
 2090 2095 2100
 Leu Val Thr Arg Phe Arg Ile Thr Ala Pro Phe Tyr His Asp Ile
 2105 2110 2115
 Ser Leu Glu Gly Gly Ala Leu Leu Asn Val Ile Ser Ser Gly Gly
 2120 2125 2130
 Gly Ala Asn Leu Asn Leu Asp Leu His Arg Ser Gly Pro Trp Gly
 2135 2140 2145
 Asn Leu Phe Ser Gln Leu Gly Met Gly Leu Ala Ala Arg Pro Phe
 2150 2155 2160
 Asp Ala Gly Gly Arg Asp Gly Arg Gly Ala His Ala Gly Arg Gln
 2165 2170 2175
 Asn Thr Phe Trp Asn Leu Gln Pro Gly Asp Val Ala Ala Ala Ala
 2180 2185 2190
 Pro Ala Leu Gln Pro Ser Ala Ala Ala Gly Asp Ala Arg Arg Leu
 2195 2200

-continued

Leu Val Asp Gly Asp Ser Leu Leu His Ala Gly Thr Gly Gln Ala
 2210 2215 2220

Arg Leu Leu Arg Gln Leu Glu Ala Asp Asp Ser Ala Glu Pro Leu
 2225 2230 2235

Leu Leu Pro Ser Cys Glu Phe Gly Pro Leu Leu Asn Phe Val Gly
 2240 2245 2250

Gly Phe Ala Gly Glu Leu Cys Lys Ser Ser Gly Trp Leu Val Ala
 2255 2260 2265

Gly Leu Pro Asp Asp Arg Pro Asp Leu His Ala Ser Gln Val Thr
 2270 2275 2280

Ala Arg Leu Gln His Gly Ala Ala Asp Asn Lys Thr His Ala
 2285 2290 2295

<210> SEQ_ID NO 71

<211> LENGTH: 373

<212> TYPE: PRT

<213> ORGANISM: Synechocystis PCC6803

<400> SEQUENCE: 71

Met Asp Phe Leu Ser Asn Phe Leu Met Asp Phe Val Lys Gln Leu Gln
 1 5 10 15

Ser Pro Thr Leu Ser Phe Leu Ile Gly Gly Met Val Ile Ala Ala Cys
 20 25 30

Gly Ser Gln Leu Gln Ile Pro Glu Ser Ile Cys Lys Ile Ile Val Phe
 35 40 45

Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly Met Ala Ile Arg Asn
 50 55 60

Ser Asn Leu Thr Glu Met Val Leu Pro Ala Leu Phe Ser Val Ala Ile
 65 70 75 80

Gly Ile Leu Ile Val Phe Ile Ala Arg Tyr Thr Leu Ala Arg Met Pro
 85 90 95

Lys Val Lys Thr Val Asp Ala Ile Ala Thr Gly Gly Leu Phe Gly Ala
 100 105 110

Val Ser Gly Ser Thr Met Ala Ala Leu Thr Leu Leu Glu Glu Gln
 115 120 125

Lys Ile Pro Tyr Glu Ala Trp Ala Gly Ala Leu Tyr Pro Phe Met Asp
 130 135 140

Ile Pro Ala Leu Val Thr Ala Ile Val Val Ala Asn Ile Tyr Leu Asn
 145 150 155 160

Lys Lys Lys Arg Lys Glu Ala Ala Phe Ala Ser Ala Gln Gly Ala Tyr
 165 170 175

Ser Lys Gln Pro Val Ala Ala Gly Asp Tyr Ser Ser Ser Asp Tyr
 180 185 190

Pro Ser Ser Arg Arg Glu Tyr Ala Gln Gln Glu Ser Gly Asp His Arg
 195 200 205

Val Lys Ile Trp Pro Ile Val Glu Glu Ser Leu Gln Gly Pro Ala Leu
 210 215 220

Ser Ala Met Leu Leu Gly Val Ala Leu Gly Leu Phe Ala Arg Pro Glu
 225 230 235 240

Ser Val Tyr Glu Gly Phe Tyr Asp Pro Leu Phe Arg Gly Leu Leu Ser
 245 250 255

Ile Leu Met Leu Val Met Gly Met Glu Ala Trp Ser Arg Ile Ser Glu
 260 265 270

Leu Arg Lys Val Ala Gln Trp Tyr Val Val Tyr Ser Ile Val Ala Pro
 275 280 285

-continued

Leu Ala His Gly Phe Ile Ala Phe Gly Leu Gly Met Ile Ala His Tyr
 290 295 300
 Ala Thr Gly Phe Ser Met Gly Gly Val Val Val Leu Ala Val Ile Ala
 305 310 315 320
 Ala Ser Ser Ser Asp Ile Ser Gly Pro Pro Thr Leu Arg Ala Gly Ile
 325 330 335
 Pro Ser Ala Asn Pro Ser Ala Tyr Ile Gly Ala Ser Thr Ala Ile Gly
 340 345 350
 Thr Pro Val Ala Ile Gly Ile Ala Ile Pro Leu Phe Leu Gly Leu Ala
 355 360 365
 Gln Thr Ile Gly Gly
 370

<210> SEQ ID NO 72
 <211> LENGTH: 374
 <212> TYPE: PRT
 <213> ORGANISM: Synechocystis PCC 6803
 <400> SEQUENCE: 72

Met	Asp	Phe	Leu	Ser	Asn	Phe	Leu	Thr	Asp	Phe	Val	Gly	Gln	Leu	Gln
1															
		5					10						15		
Ser	Pro	Thr	Leu	Ala	Phe	Leu	Ile	Gly	Gly	Met	Val	Ile	Ala	Ala	Leu
		20					25						30		
Gly	Thr	Gln	Leu	Val	Ile	Pro	Glu	Ala	Ile	Ser	Thr	Ile	Ile	Val	Phe
		35				40						45			
Met	Leu	Leu	Thr	Lys	Ile	Gly	Leu	Thr	Gly	Gly	Met	Ala	Ile	Arg	Asn
		50				55						60			
Ser	Asn	Leu	Thr	Glu	Met	Leu	Leu	Pro	Val	Ala	Phe	Ser	Val	Ile	Leu
		65				70						75			80
Gly	Ile	Leu	Ile	Val	Phe	Ile	Ala	Arg	Phe	Thr	Leu	Ala	Lys	Leu	Pro
		85				90						95			
Asn	Val	Arg	Thr	Val	Asp	Ala	Leu	Ala	Thr	Gly	Gly	Leu	Phe	Gly	Ala
		100					105					110			
Val	Ser	Gly	Ser	Thr	Met	Ala	Ala	Ala	Leu	Thr	Thr	Leu	Glu	Glu	Ser
		115				120						125			
Lys	Ile	Ser	Tyr	Glu	Ala	Trp	Ala	Gly	Ala	Leu	Tyr	Pro	Phe	Met	Asp
		130				135						140			
Ile	Pro	Ala	Leu	Val	Thr	Ala	Ile	Val	Val	Ala	Asn	Ile	Tyr	Leu	Asn
		145				150						155			160
Lys	Arg	Lys	Arg	Lys	Ser	Ala	Ala	Ala	Ser	Ile	Glu	Glu	Ser	Phe	Ser
		165				170						175			
Lys	Gln	Pro	Val	Ala	Ala	Gly	Asp	Tyr	Gly	Asp	Gln	Thr	Asp	Tyr	Pro
		180				185						190			
Arg	Thr	Arg	Gln	Glu	Tyr	Leu	Ser	Gln	Gln	Glu	Pro	Glu	Asp	Asn	Arg
		195				200						205			
Val	Lys	Ile	Trp	Pro	Ile	Glu	Glu	Ser	Leu	Gln	Gly	Pro	Ala	Leu	
		210				215						220			
Ser	Ala	Met	Leu	Leu	Gly	Leu	Ala	Leu	Gly	Ile	Phe	Thr	Lys	Pro	Glu
		225				230						235			240
Ser	Val	Tyr	Glu	Gly	Phe	Tyr	Asp	Pro	Leu	Phe	Arg	Gly	Leu	Leu	Ser
		245				250						255			
Ile	Leu	Met	Leu	Ile	Met	Gly	Met	Glu	Ala	Trp	Ser	Arg	Ile	Gly	Glu
		260				265						270			
Leu	Arg	Lys	Val	Ala	Gln	Trp	Tyr	Val	Val	Tyr	Ser	Leu	Ile	Ala	Pro

US 11,001,853 B2

187

-continued

188

275	280	285
Ile Val His Gly Phe Ile Ala Phe Gly Leu Gly Met	Ile Ala His Tyr	
290	295	300
Ala Thr Gly Phe Ser Leu Gly Gly Val Val Val	Leu Ala Val Ile Ala	
305	310	315
Ala Ser Ser Ser Asp Ile Ser Gly Pro Pro Thr	Leu Arg Ala Gly Ile	
325	330	335
Pro Ser Ala Asn Pro Ser Ala Tyr Ile Gly Ser Ser	Thr Ala Ile Gly	
340	345	350
Thr Pro Ile Ala Ile Gly Val Cys Ile Pro Leu Phe	Ile Gly Leu Ala	
355	360	365
Gln Thr Leu Gly Ala Gly		
370		

<210> SEQ ID NO 73

<211> LENGTH: 370

<212> TYPE: PRT

<213> ORGANISM: Nostoc PCC 7120

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: Anabaena

<400> SEQUENCE: 73

Met Asp Phe Phe Ser Leu Phe Leu Met Asp Phe Val Lys	Gln Leu Gln		
1	5	10	15
Ser Pro Thr Leu Gly Phe Leu Ile Gly Gly Met Val Ile	Ala Ala Leu		
20	25	30	
Gly Ser Glu Leu Ile Ile Pro Glu Ala Ile Cys Gln Ile	Ile Val Phe		
35	40	45	
Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly Ile Ala	Ile Arg Asn		
50	55	60	
Ser Asn Leu Thr Glu Met Val Leu Pro Ala Ala Ser Ala	Val Ala Val		
65	70	75	80
Gly Val Leu Val Val Phe Ile Ala Arg Tyr Thr Leu Ala	Lys Leu Pro		
85	90	95	
Lys Val Asn Thr Val Asp Ala Ile Ala Thr Gly Gly Leu	Phe Gly Ala		
100	105	110	
Val Ser Gly Ser Thr Met Ala Ala Ala Leu Thr Leu Leu	Glu Glu Gln		
115	120	125	
Lys Ile Gln Tyr Glu Ala Trp Ala Ala Ala Leu Tyr Pro	Phe Met Asp		
130	135	140	
Ile Pro Ala Leu Val Thr Ala Ile Val Val Ala Asn Ile	Tyr Leu Asn		
145	150	155	160
Lys Lys Lys Arg Ser Ala Ala Gly Glu Tyr Leu Ser Lys	Gln Ser Val		
165	170	175	
Ala Ala Gly Glu Tyr Pro Asp Gln Gln Asp Tyr Pro Ser	Ser Arg Gln		
180	185	190	
Glu Tyr Leu Arg Lys Gln Gln Ser Ala Asp Asn Arg	Val Lys Ile Trp		
195	200	205	
Pro Ile Val Lys Glu Ser Leu Gln Gly Pro Ala Leu Ser	Ala Met Leu		
210	215	220	
Leu Gly Ile Ala Leu Gly Leu Phe Thr Gln Pro Glu Ser	Val Tyr Lys		
225	230	235	240
Ser Phe Tyr Asp Pro Leu Phe Arg Gly Leu Leu Ser Ile	Leu Met Leu		
245	250	255	

US 11,001,853 B2

189**190**

-continued

Val Met Gly Met Glu Ala Trp Ser Arg Ile Gly Glu Leu Arg Lys Val
260 265 270

Ala Gln Trp Tyr Val Val Tyr Ser Val Val Ala Pro Leu Val His Gly
275 280 285

Phe Ile Ala Phe Gly Leu Gly Met Ile Ala His Tyr Ala Thr Gly Phe
290 295 300

Ser Leu Gly Gly Val Val Ile Leu Ala Val Ile Ala Ala Ser Ser Ser
305 310 315 320

Asp Ile Ser Gly Pro Pro Thr Leu Arg Ala Gly Ile Pro Ser Ala Asn
325 330 335

Pro Ser Ala Tyr Ile Gly Ala Ser Thr Ala Ile Gly Thr Pro Ile Ala
340 345 350

Ile Gly Leu Ala Ile Pro Leu Phe Leu Gly Leu Ala Gln Ala Ile Gly
355 360 365

Gly Arg
370

<210> SEQ ID NO 74

<211> LENGTH: 377

<212> TYPE: PRT

<213> ORGANISM: Cyanothecce sp. PCC 7425

<400> SEQUENCE: 74

Met Asp Phe Trp Ser Tyr Phe Leu Met Asp Phe Val Lys Gln Leu Gln
1 5 10 15

Ser Pro Thr Leu Gly Phe Leu Ile Gly Gly Met Val Ile Ala Ala Leu
20 25 30

Gly Ser Gln Leu Val Ile Pro Glu Ala Ile Cys Gln Ile Ile Val Phe
35 40 45

Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly Met Ala Ile Arg Asn
50 55 60

Ser Asn Leu Thr Glu Met Val Leu Pro Ala Ala Phe Ser Val Ile Ser
65 70 75 80

Gly Ile Leu Ile Val Phe Ile Ala Arg Tyr Thr Leu Ala Lys Leu Pro
85 90 95

Lys Val Arg Thr Val Asp Ala Ile Ala Thr Gly Gly Leu Phe Gly Ala
100 105 110

Val Ser Gly Ser Thr Met Ala Ala Ala Leu Thr Leu Leu Glu Glu Glu
115 120 125

Lys Ile Pro Tyr Glu Ala Trp Ala Gly Ala Leu Tyr Pro Phe Met Asp
130 135 140

Ile Pro Ala Leu Val Thr Ala Ile Val Ile Ala Asn Ile Tyr Leu Asn
145 150 155 160

Lys Lys Lys Arg Arg Ala Glu Ser Glu Ala Leu Ser Lys Gln Glu Tyr
165 170 175

Leu Gly Lys Gln Ser Ile Val Ala Gly Asp Tyr Pro Ala Gln Gln Asp
180 185 190

Tyr Pro Ser Thr Arg Gln Glu Tyr Leu Ser Lys Gln Gln Gly Pro Glu
195 200 205

Asn Asn Arg Val Lys Ile Trp Pro Ile Val Gln Glu Ser Leu Gln Gly
210 215 220

Pro Ala Leu Ser Ala Met Leu Leu Gly Val Ala Leu Gly Ile Leu Thr
225 230 235 240

Lys Pro Glu Ser Val Tyr Glu Ser Phe Tyr Asp Pro Leu Phe Arg Gly
245 250 255

US 11,001,853 B2

191**192**

-continued

Leu Leu Ser Ile Leu Met Leu Val Met Gly Met Glu Ala Trp Ser Arg
 260 265 270
 Ile Gly Glu Leu Arg Lys Val Ala Gln Trp Tyr Val Val Tyr Ser Val
 275 280 285
 Val Ala Pro Phe Val His Gly Leu Ile Ala Phe Gly Leu Gly Met Phe
 290 295 300
 Ala His Tyr Thr Met Gly Phe Ser Met Gly Gly Val Val Val Leu Ala
 305 310 315 320
 Val Ile Ala Ser Ser Ser Asp Ile Ser Gly Pro Pro Thr Leu Arg
 325 330 335
 Ala Gly Ile Pro Ser Ala Asn Pro Ser Ala Tyr Ile Gly Ala Ser Thr
 340 345 350
 Ala Ile Gly Thr Pro Ile Ala Ile Gly Leu Cys Ile Pro Phe Phe Ile
 355 360 365
 Gly Leu Ala Gln Thr Leu Gly Gly Gly
 370 375

<210> SEQ ID NO 75
 <211> LENGTH: 373
 <212> TYPE: PRT
 <213> ORGANISM: *Microcystis aeruginosa*
 <400> SEQUENCE: 75

Met Asp Phe Phe Ser Leu Phe Val Met Asp Phe Ile Gln Gln Leu Gln
 1 5 10 15
 Ser Pro Thr Leu Ala Phe Leu Ile Gly Gly Met Ile Ile Ala Ala Leu
 20 25 30
 Gly Ser Glu Leu Val Ile Pro Glu Ser Ile Cys Thr Ile Ile Val Phe
 35 40 45
 Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly Ile Ala Ile Arg Asn
 50 55 60
 Ser Asn Leu Thr Glu Met Val Leu Pro Met Ile Phe Ala Val Ile Val
 65 70 75 80
 Gly Ile Ile Val Val Phe Val Ala Arg Tyr Thr Leu Ala Asn Leu Pro
 85 90 95
 Lys Val Lys Val Val Asp Ala Ile Ala Thr Gly Gly Leu Phe Gly Ala
 100 105 110
 Val Ser Gly Ser Thr Met Ala Ala Gly Leu Thr Val Leu Glu Glu Gln
 115 120 125
 Lys Ile Pro Tyr Glu Ala Trp Ala Gly Ala Leu Tyr Pro Phe Met Asp
 130 135 140
 Ile Pro Ala Leu Val Thr Ala Ile Val Val Ala Asn Ile Tyr Leu Asn
 145 150 155 160
 Lys Lys Lys Gln Lys Glu Ala Ala Tyr Asp Gln Glu Ser Phe Ser Lys
 165 170 175
 Gln Pro Val Ala Ala Gly Asn Tyr Ser Asp Gln Gln Asp Tyr Pro Ser
 180 185 190
 Ser Arg Gln Glu Tyr Leu Ser Gln Gln Gln Pro Ala Asp Asn Arg Val
 195 200 205
 Lys Ile Trp Pro Ile Ile Glu Glu Ser Leu Arg Gly Pro Ala Leu Ser
 210 215 220
 Ala Met Leu Leu Gly Leu Ala Leu Gly Ile Phe Thr Gln Pro Glu Ser
 225 230 235 240
 Val Tyr Lys Ser Phe Tyr Asp Pro Leu Phe Arg Gly Leu Leu Ser Val

US 11,001,853 B2

193

194

-continued

245 250 255

250

255

Leu Met Leu Val Met Gly Met Glu Ala Trp Ser Arg Val Gly Glu Leu
 260 265 270

Arg Lys Val Ala Gln Trp Tyr Val Val Tyr Ser Val Ile Ala Pro Phe
275 280 285

Thr	Gly	Phe	Ser	Trp	Gly	Gly	Val	Val	Met	Leu	Ala	Val	Ile	Ala	Ser
305				310					315					320	

Ser Ser Ser Asp Ile Ser Gly Pro Pro Thr Leu Arg Ala Gly Ile Pro
325 330 335

Ser Ala Asn Pro Ser Ala Tyr Ile Gly Ala Ser Thr Ala Ile Gly Thr
340 345 350

Pro Val Ala Ile Gly Leu Cys Ile Pro Phe Phe Val Gly Leu Ala Gln
355 360 365

Ala Leu Ser Gly Gly
370

<210> SEQ ID NO: 76
<11> LENGTH: 360

<211> LENGTH: 369
<212> TYPE: PRT

<215> ORGANISM: *Anabacia variabilis*

Met Asp Phe Val Ser Leu Phe Val Lys Asp Phe Ile Ala Gln Leu Gln

Ser Pro Thr Leu Ala Phe Leu Ile Gly Gly Met Ile Ile Ala Ala Leu

Gly Ser Glu Leu Val Ile Pro Glu Ser Ile Cys Thr Ile Ile Val Phe

Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly Ile Ala Ile Arg Asn

65 70 75 80

Gly Ile Thr Ile Val Phe Ile Ser Arg Tyr Ile Leu Ala Lys Leu Pro
85 90 95

Lys Val Lys Val Val Asp Ala Ile Ala Thr Gly Gly Leu Phe Gly Ala
100 105 110

Val Ser Gly Ser Thr Met Ala Ala Gly Leu Thr Val Leu Glu Glu Gln

Lys Met Ala Tyr Glu Ala Trp Ala Gly Ala Leu Tyr Pro Phe Met Asp

Ile Pro Ala Leu Val Thr Ala Ile Val Ile Ala Asn Ile Tyr Leu Asn

Lys Lys Lys Arg Lys Glu Ala Val Tyr Ser Thr Glu Gln Pro Val Ala

Ala-Gly-Asp-Tyr-Pro-Asp-Gln-Lys-Asp-Tyr-Pro-Ser-Ser-Arg-Gln-Glu

180 185 190

195 200 205

210 215 220

US 11,001,853 B2

195**196**

-continued

Ser Phe Tyr Asp Pro Ala Phe Arg Gly Leu Leu Ser Ile Leu Met Leu
245 250 255

Val Met Gly Met Glu Ala Trp Ser Arg Ile Gly Glu Leu Arg Lys Val
260 265 270

Ala Gln Trp Tyr Val Val Tyr Ser Val Val Ala Pro Phe Val His Gly
275 280 285

Leu Ile Ala Phe Gly Leu Gly Met Ile Ala His Tyr Thr Met Asn Phe
290 295 300

Ser Met Gly Gly Val Val Ile Leu Ala Val Ile Ala Ser Ser Ser Ser
305 310 315 320

Asp Ile Ser Gly Pro Pro Thr Leu Arg Ala Gly Ile Pro Ser Ala Asn
325 330 335

Pro Ser Ala Tyr Ile Gly Ala Ser Thr Ala Val Gly Thr Pro Val Ala
340 345 350

Ile Gly Leu Cys Ile Pro Phe Phe Leu Gly Leu Ala Gln Ala Ile Gly
355 360 365

Gly

<210> SEQ_ID NO 77

<211> LENGTH: 1082

<212> TYPE: PRT

<213> ORGANISM: Chlamydomonas reinhardtii

<400> SEQUENCE: 77

Met Leu Pro Gly Leu Gly Val Ile Leu Leu Val Leu Pro Met Gln Tyr
1 5 10 15

Tyr Phe Gly Tyr Lys Ile Val Gln Ile Lys Leu Gln Asn Ala Lys His
20 25 30

Val Ala Leu Arg Ser Ala Ile Met Gln Glu Val Leu Pro Ala Ile Lys
35 40 45

Leu Val Lys Tyr Tyr Ala Trp Glu Gln Phe Phe Glu Asn Gln Ile Ser
50 55 60

Lys Val Arg Arg Glu Glu Ile Arg Leu Asn Phe Trp Asn Cys Val Met
65 70 75 80

Lys Val Ile Asn Val Ala Cys Val Phe Cys Val Pro Pro Met Thr Ala
85 90 95

Phe Val Ile Phe Thr Thr Tyr Glu Phe Gln Arg Ala Arg Leu Val Ser
100 105 110

Ser Val Ala Phe Thr Thr Leu Ser Leu Phe Asn Ile Leu Arg Phe Pro
115 120 125

Leu Val Val Leu Pro Lys Ala Leu Arg Ala Val Ser Glu Ala Asn Ala
130 135 140

Ser Leu Gln Arg Leu Glu Ala Tyr Leu Leu Glu Glu Val Pro Ser Gly
145 150 155 160

Thr Ala Ala Val Lys Thr Pro Lys Asn Ala Pro Pro Gly Ala Val Ile
165 170 175

Glu Asn Gly Val Phe His His Pro Ser Asn Pro Asn Trp His Leu His
180 185 190

Val Pro Lys Phe Glu Val Lys Pro Gly Gln Val Val Ala Val Val Gly
195 200 205

Arg Ile Ala Ala Gly Lys Ser Ser Leu Val Gln Ala Ile Leu Gly Asn
210 215 220

Met Val Lys Glu His Gly Ser Phe Asn Val Gly Gly Arg Ile Ser Tyr
225 230 235 240

US 11,001,853 B2

197**198**

-continued

Val Pro Gln Asn Pro Trp Leu Gln Asn Leu Ser Leu Arg Asp Asn Val
245 250 255

Leu Phe Gly Glu Gln Phe Asp Glu Asn Lys Tyr Thr Asp Val Ile Glu
260 265 270

Ser Cys Ala Leu Thr Leu Asp Leu Gln Ile Leu Ser Asn Gly Asp Gln
275 280 285

Ser Lys Ala Gly Ile Arg Gly Val Asn Phe Ser Gly Gly Gln Arg Gln
290 295 300

Arg Val Asn Leu Ala Arg Cys Ala Tyr Ala Asp Ala Asp Leu Val Leu
305 310 315 320

Leu Asp Asn Ala Leu Ser Ala Val Asp His His Thr Ala His His Ile
325 330 335

Phe Asp Lys Cys Ile Lys Gly Leu Phe Ser Asp Lys Ala Val Val Leu
340 345 350

Val Thr His Gln Ile Glu Phe Met Pro Arg Cys Asp Asn Val Ala Ile
355 360 365

Met Asp Glu Gly Arg Cys Leu Tyr Phe Gly Lys Trp Asn Glu Glu Ala
370 375 380

Gln His Leu Leu Gly Lys Leu Leu Pro Ile Thr His Leu Leu His Ala
385 390 395 400

Ala Gly Ser Gln Glu Ala Pro Pro Ala Pro Lys Lys Lys Ala Glu Asp
405 410 415

Lys Ala Gly Pro Gln Lys Ser Gln Ser Leu Gln Leu Thr Leu Ala Pro
420 425 430

Thr Ser Ile Gly Lys Pro Thr Glu Lys Pro Lys Asp Val Gln Lys Leu
435 440 445

Thr Ala Tyr Gln Ala Ala Leu Ile Tyr Thr Trp Tyr Gly Asn Leu Phe
450 455 460

Leu Val Gly Val Cys Phe Phe Phe Leu Ala Ala Gln Cys Ser Arg
465 470 475 480

Gln Ile Ser Asp Phe Trp Val Arg Trp Trp Val Asn Asp Glu Tyr Lys
485 490 495

Lys Phe Pro Val Lys Gly Glu Gln Asp Ser Ala Ala Thr Thr Phe Tyr
500 505 510

Cys Leu Ile Tyr Leu Leu Leu Val Gly Leu Phe Tyr Ile Phe Met Ile
515 520 525

Phe Arg Gly Ala Thr Phe Leu Trp Trp Val Leu Lys Ser Ser Glu Thr
530 535 540

Ile Arg Arg Lys Ala Leu His Asn Val Leu Asn Ala Pro Met Gly Phe
545 550 555 560

Phe Leu Val Thr Pro Val Gly Asp Leu Leu Asn Phe Thr Lys Asp
565 570 575

Gln Asp Ile Met Asp Glu Asn Leu Pro Asp Ala Val His Phe Met Gly
580 585 590

Ile Tyr Gly Leu Ile Leu Leu Ala Thr Thr Ile Thr Val Ser Val Thr
595 600 605

Ile Asn Phe Phe Ala Ala Phe Thr Gly Ala Leu Ile Ile Met Thr Leu
610 615 620

Ile Met Leu Ser Ile Tyr Leu Pro Ala Ala Thr Ala Leu Lys Lys Ala
625 630 635 640

Arg Ala Val Ser Gly Gly Met Leu Val Gly Leu Val Ala Glu Val Leu
645 650 655

Glu Gly Leu Gly Val Val Gln Ala Phe Asn Lys Gln Glu Tyr Phe Ile

US 11,001,853 B2

199**200**

-continued

660	665	670
Glu Glu Ala Ala Arg Arg Thr Asn Ile Thr Asn Ser Ala Val Phe Asn		
675	680	685
Ala Glu Ala Leu Asn Leu Trp Leu Ala Phe Trp Cys Asp Phe Ile Gly		
690	695	700
Ala Cys Leu Val Gly Val Val Ser Ala Phe Ala Val Gly Met Ala Lys		
705	710	715
Asp Leu Gly Gly Ala Thr Val Gly Leu Ala Phe Ser Asn Ile Ile Gln		
725	730	735
Met Leu Val Phe Tyr Thr Trp Val Val Arg Phe Ile Ser Glu Ser Ile		
740	745	750
Ser Leu Phe Asn Ser Val Glu Gly Met Ala Tyr Leu Ala Asp Tyr Val		
755	760	765
Pro His Asp Gly Val Phe Tyr Asp Gln Arg Gln Lys Asp Gly Val Ala		
770	775	780
Lys Gln Ile Val Leu Pro Asp Gly Asn Ile Val Pro Ala Ala Ser Lys		
785	790	795
Val Gln Val Val Asp Asp Ala Ala Leu Ala Arg Trp Pro Ala Thr		
805	810	815
Gly Asn Ile Arg Phe Glu Asp Val Trp Met Gln Tyr Arg Leu Asp Ala		
820	825	830
Pro Trp Ala Leu Lys Gly Val Thr Phe Lys Ile Asn Asp Gly Glu Lys		
835	840	845
Val Gly Ala Val Gly Arg Thr Gly Ser Gly Lys Ser Thr Thr Leu Leu		
850	855	860
Ala Leu Tyr Arg Met Phe Glu Leu Gly Lys Gly Arg Ile Leu Val Asp		
865	870	875
Gly Val Asp Ile Ala Thr Leu Ser Leu Lys Arg Leu Arg Thr Gly Leu		
885	890	895
Ser Ile Ile Pro Gln Glu Pro Val Met Phe Thr Gly Thr Val Arg Ser		
900	905	910
Asn Leu Asp Pro Phe Gly Glu Phe Lys Asp Asp Ala Ile Leu Trp Glu		
915	920	925
Val Leu Lys Lys Val Gly Leu Glu Asp Gln Ala Gln His Ala Gly Gly		
930	935	940
Leu Asp Gly Gln Val Asp Gly Thr Gly Lys Ala Trp Ser Leu Gly		
945	950	955
Gln Met Gln Leu Val Cys Leu Ala Arg Ala Ala Leu Arg Ala Val Pro		
965	970	975
Ile Leu Cys Leu Asp Glu Ala Thr Ala Ala Met Asp Pro His Thr Glu		
980	985	990
Ala Ile Val Gln Gln Thr Ile Lys Lys Val Phe Asp Asp Arg Thr Thr		
995	1000	1005
Ile Thr Ile Ala His Arg Leu Asp Thr Ile Ile Glu Ser Asp Lys		
1010	1015	1020
Ile Ile Val Met Glu Gln Gly Ser Leu Met Glu Tyr Glu Ser Pro		
1025	1030	1035
Ser Lys Leu Leu Ala Asn Arg Asp Ser Met Phe Ser Lys Leu Val		
1040	1045	1050
Asp Lys Thr Gly Pro Ala Ala Ala Leu Arg Lys Met Ala		
1055	1060	1065
Glu Asp Phe Trp Ser Thr Arg Ser Ala Gln Gly Arg Asn Gln		
1070	1075	1080

-continued

<210> SEQ_ID NO 78
 <211> LENGTH: 366
 <212> TYPE: PRT
 <213> ORGANISM: Cyanothecce
 <400> SEQUENCE: 78

Met	Asp	Phe	Leu	Ser	Leu	Phe	Val	Lys	Asp	Phe	Ile	Ile	Gln	Leu	Gln
1								10						15	
Ser	Pro	Thr	Leu	Ala	Phe	Leu	Ile	Gly	Gly	Met	Val	Ile	Ala	Ala	Leu
			20				25						30		
Gly	Ser	Glu	Leu	Val	Ile	Pro	Glu	Ser	Ile	Cys	Thr	Ile	Ile	Val	Phe
			35				40					45			
Met	Leu	Leu	Thr	Lys	Ile	Gly	Leu	Thr	Gly	Gly	Ile	Ala	Ile	Arg	Asn
			50				55				60				
Ser	Asn	Leu	Thr	Glu	Met	Val	Leu	Pro	Met	Ile	Cys	Ala	Val	Ile	Val
			65				70			75			80		
Gly	Ile	Val	Val	Val	Phe	Ile	Ala	Arg	Tyr	Thr	Leu	Ala	Lys	Leu	Pro
			85				90						95		
Lys	Val	Asn	Val	Val	Asp	Ala	Ile	Ala	Thr	Gly	Gly	Leu	Phe	Gly	Ala
			100				105					110			
Val	Ser	Gly	Ser	Thr	Met	Ala	Ala	Gly	Leu	Thr	Val	Leu	Glu	Gln	
			115				120					125			
Lys	Ile	Pro	Tyr	Glu	Ala	Trp	Ala	Gly	Ala	Leu	Tyr	Pro	Phe	Met	Asp
			130				135				140				
Ile	Pro	Ala	Leu	Val	Thr	Ala	Ile	Val	Val	Ala	Asn	Ile	Tyr	Leu	Asn
			145				150			155			160		
Lys	Lys	Lys	Arg	Lys	Ala	Thr	Val	Met	Gln	Glu	Ser	Leu	Ser	Lys	Gln
			165				170					175			
Pro	Val	Ala	Ala	Gly	Asp	Tyr	Pro	Ser	Ser	Arg	Gln	Glu	Tyr	Val	Ser
			180				185					190			
Gln	Gln	Gln	Pro	Glu	Asp	Asn	Arg	Val	Lys	Ile	Trp	Pro	Ile	Ile	Glu
			195				200				205				
Glu	Ser	Leu	Arg	Gly	Pro	Ala	Leu	Ser	Ala	Met	Leu	Leu	Gly	Leu	Ala
			210				215			220					
Leu	Gly	Ile	Leu	Thr	Gln	Pro	Glu	Ser	Val	Tyr	Lys	Gly	Phe	Tyr	Asp
			225				230			235			240		
Pro	Pro	Phe	Arg	Gly	Leu	Leu	Ser	Ile	Leu	Met	Leu	Val	Met	Gly	Met
			245				250			255					
Glu	Ala	Trp	Ser	Arg	Ile	Gly	Glu	Leu	Arg	Lys	Val	Ala	Gln	Trp	Tyr
			260				265			270					
Val	Val	Tyr	Ser	Val	Ala	Ala	Pro	Phe	Ile	His	Gly	Leu	Leu	Ala	Phe
			275				280					285			
Gly	Leu	Gly	Met	Ile	Ala	His	Tyr	Thr	Met	Gly	Phe	Ser	Met	Gly	Gly
			290				295			300					
Val	Val	Ile	Leu	Ala	Val	Ile	Ala	Ser	Ser	Ser	Ser	Asp	Ile	Ser	Gly
			305				310			315			320		
Pro	Pro	Thr	Leu	Arg	Ala	Gly	Ile	Pro	Ser	Ala	Asn	Pro	Ser	Ala	Tyr
			325				330					335			
Ile	Gly	Ala	Ser	Thr	Ala	Ile	Gly	Thr	Pro	Val	Ala	Ile	Gly	Leu	Cys
			340				345					350			
Ile	Pro	Phe	Phe	Val	Gly	Leu	Ala	Gln	Ala	Ile	Gly	Gly	Phe		
			355				360					365			

-continued

<210> SEQ_ID NO 79
<211> LENGTH: 337
<212> TYPE: PRT
<213> ORGANISM: Volvox carteri
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: f. nagariensis

<400> SEQUENCE: 79

Met	Gln	Thr	Thr	Met	Ser	Val	Thr	Arg	Pro	Cys	Val	Gly	Leu	Arg	Pro
1															
													15		

Leu	Pro	Val	Arg	Asn	Val	Arg	Ser	Leu	Ile	Arg	Ala	Gln	Ala	Ala	Pro
													20	25	30

Gln	Gln	Val	Ser	Thr	Ala	Val	Ser	Thr	Asn	Gly	Asn	Gly	Asn	Gly	Val
													35	40	45

Ala	Ala	Ala	Ser	Leu	Ser	Val	Pro	Ala	Pro	Val	Ala	Ala	Pro	Ala	Gln
													50	55	60

Ala	Val	Ser	Thr	Pro	Val	Arg	Ala	Val	Ser	Val	Leu	Thr	Pro	Pro	Gln	
													65	70	75	80

Val	Tyr	Glu	Asn	Ala	Ala	Asn	Val	Gly	Ala	Tyr	Lys	Ala	Ser	Leu	Gly
													85	90	95

Val	Leu	Ala	Thr	Phe	Val	Gln	Ile	Gln	Ala	Gly	Ala	Tyr	Ile	Ala	
													100	105	110

Phe	Gly	Ala	Phe	Leu	Ala	Cys	Ser	Val	Gly	Gly	Asn	Ile	Pro	Gly	Ile
													115	120	125

Thr	Ala	Ser	Asn	Pro	Gly	Leu	Ala	Lys	Leu	Leu	Phe	Ala	Leu	Val	Phe
													130	135	140

Pro	Val	Gly	Leu	Ser	Met	Val	Thr	Asn	Cys	Gly	Ala	Glu	Leu	Tyr	Thr	
													145	150	155	160

Gly	Asn	Thr	Met	Met	Leu	Thr	Cys	Ala	Ile	Phe	Glu	Lys	Lys	Ala	Thr
													165	170	175

Trp	Ala	Gln	Leu	Val	Lys	Asn	Trp	Val	Val	Ser	Tyr	Ala	Gly	Asn	Phe
													180	185	190

Val	Gly	Ser	Ile	Ala	Met	Val	Ala	Ala	Val	Ala	Thr	Gly	Leu	Met	
													195	200	205

Ala	Ser	Asn	Gln	Leu	Pro	Val	Asn	Met	Ala	Thr	Ala	Lys	Ser	Ser	Leu
													210	215	220

Gly	Phe	Thr	Glu	Val	Leu	Ser	Arg	Ser	Ile	Leu	Cys	Asn	Trp	Leu	Val	
													225	230	235	240

Cys	Cys	Ala	Val	Trp	Ser	Ala	Ser	Ala	Ala	Thr	Ser	Leu	Pro	Gly	Arg
													245	250	255

Ile	Leu	Gly	Leu	Trp	Pro	Pro	Ile	Thr	Ala	Phe	Val	Ala	Ile	Gly	Leu
													260	265	270

Glu	His	Ser	Val	Ala	Asn	Met	Phe	Val	Ile	Pro	Leu	Gly	Met	Met	Leu
													275	280	285

Gly	Ala	Asp	Val	Thr	Trp	Ser	Gln	Phe	Phe	Phe	Asn	Asn	Leu	Val	Pro
													290	295	300

Val	Thr	Leu	Gly	Asn	Thr	Ile	Ala	Gly	Val	Val	Met	Met	Ala	Val	Ala	
													305	310	315	320

Tyr	Ser	Val	Ser	Tyr	Gly	Ser	Leu	Gly	Lys	Thr	Pro	Lys	Pro	Ala	Thr
													325	330	335

Ala

<210> SEQ_ID NO 80
<211> LENGTH: 2297
<212> TYPE: PRT

US 11,001,853 B2

205**206**

-continued

<213> ORGANISM: Chlorella variabilis

<400> SEQUENCE: 80

Met	Val	Pro	Leu	Leu	Ala	Gln	Arg	Gly	Arg	Ile	Arg	Ser	Gln	Ala	Pro
1				5				10						15	

Arg	Thr	Trp	His	Pro	Asp	Pro	Gln	Pro	Leu	His	Ala	Glu	Arg	Ser	Arg
				20				25				30			

Gln	Cys	Pro	Gly	Arg	Gly	Val	Arg	Ala	Ala	Ala	Lys	Arg	Gly	Gly	Gly
						35	40				45				

Ser	Gly	Gly	Ala	Thr	His	Lys	Ser	Lys	Ser	Lys	Glu	Leu	Asp	Glu	
						50	55				60				

Val	Ala	Ala	Phe	Glu	Gln	Leu	Met	Cys	Asp	Trp	Asp	Asp	Ala	Phe	Ala
65						70		75					80		

Ala	Asp	Cys	Tyr	Asp	Asn	Glu	Arg	Ala	Ala	Arg	Met	Ala	Arg	Leu	Ala
						85		90			95				

Glu	Glu	Gly	Tyr	Gln	His	His	Gly	Arg	Gly	Phe	Val	Phe	Val	Arg	Ser
						100		105			110				

Arg	Leu	Asp	Lys	Arg	Ser	Arg	Lys	Ala	Arg	Asn	Asp	Ser	Gly	Ala	Ser
						115		120			125				

Lys	Gly	Phe	Gly	Ala	Ala	Ala	Lys	Ala	Leu	Ser	Val	Glu	Gln	Gly	Thr
						130		135			140				

Pro	Leu	Glu	Asn	Asn	Pro	Gln	Leu	His	Leu	Leu	Ser	Trp	Thr	Ala	Cys
145						150			155			160			

Tyr	Ile	Ala	Ser	Ser	Gln	Leu	Asp	Ser	Leu	Gly	Gly	Leu	Phe	Ser	Thr
						165		170			175				

Gln	Glu	Gly	Val	Leu	Leu	Pro	Asp	Ser	Gly	Ser	Leu	Leu	Thr	Asp	Gly
						180		185			190				

Gly	Ser	Gly	Ala	Ser	Gly	Ser	Asn	Ala	Ala	Asp	Ala	Val	Gly	Glu	Leu
						195		200			205				

Gln	Arg	Val	Leu	Arg	Gly	Gln	Asp	Leu	Ser	Gln	Leu	Arg	Gly	Tyr	Val
						210		215			220				

Gly	Ala	Pro	Pro	Gln	Ala	Arg	Pro	Ala	Ser	Gly	Ser	Asp	Asp	Asp	Gly
225						230			235			240			

Ser	Ser	Thr	Thr	Gly	Ser	Asn	Asn	Gly	Ala	Ala	Gly	Glu	Gly	Ser	Glu
						245		250			255				

Val	Glu	Glu	Gly	Thr	Ala	Met	Gly	Gly	Ile	Arg	Arg	Tyr	Glu	Pro	Glu
						260		265			270				

Ser	Gly	Glu	Leu	Val	Val	Leu	Leu	Ser	Cys	Lys	Ile	Gly	Gly	Lys	Pro
						275		280			285				

Ala	Val	Gly	Ala	Glu	Leu	Leu	Ala	Val	Ala	Gln	Ala	Glu	Asp	Gly	Lys
						290		295			300				

His	Ala	Pro	Gly	Ala	Ser	Pro	Asp	Thr	Arg	Leu	Cys	Lys	Glu	Pro	Ser
305						310			315			320			

Gln	Ser	Ala	Phe	Asp	Leu	Trp	Ser	Phe	Gly	Trp	Met	Asn	Lys	Ile	Val
						325		330			335				

Pro	Ala	Ala	Arg	Arg	Gly	Glu	Val	Glu	Val	Ala	Asp	Leu	Pro	Leu	Pro
						340		345			350				

Glu	Ala	Gln	Gln	Ala	Glu	Pro	Cys	Tyr	Glu	Glu	Leu	Asn	Thr	Asn	Trp
						355		360			365				

Glu	Ala	Ala	Val	Gln	Glu	Ala	Lys	Lys	Ala	Gly	Lys	Glu	Pro	Lys	Leu
						370		375			380				

Met	Lys	Val	Leu	Trp	Lys	Thr	Tyr	Gly	Lys	Asp	Ile	Val	Leu	Ala	Gly
385						390			395			400			

-continued

Ile Phe Lys Leu Met Trp Ser Val Phe Val Ile Leu Gly Ala Tyr Tyr
405 410 415

Phe Thr Arg Ser Ile Leu Met Cys Ile Arg Thr Leu Glu Gly Lys Asp
420 425 430

Asp Ser Ile Tyr Asp Thr Glu Trp Lys Gly Trp Val Leu Thr Gly Phe
435 440 445

Phe Phe Leu Asp Ala Trp Leu Leu Gly Met Met Leu Gln Arg Met Ala
450 455 460

Phe Asn Cys Leu Lys Val Gly Ile Lys Ala Arg Ala Ala Leu Thr Thr
465 470 475 480

Met Ile Ala Arg Lys Cys Tyr Asn Met Ala His Leu Thr Lys Asp Thr
485 490 495

Ala Ala Glu Ala Val Gly Phe Val Ala Ser Asp Ile Asn Lys Val Phe
500 505 510

Glu Gly Ile Gln Glu Val His Tyr Leu Trp Gly Ala Pro Val Glu Ala
515 520 525

Gly Ala Ile Leu Ala Leu Leu Gly Thr Leu Val Gly Val Tyr Cys Ile
530 535 540

Gly Gly Val Ile Ile Val Cys Met Val Val Pro Leu Gln Tyr Tyr Phe
545 550 555 560

Gly Tyr Lys Ile Ile Lys Asn Lys Ile Lys Asn Ala Pro Asn Val Thr
565 570 575

Glu Arg Trp Ser Ile Ile Gln Glu Ile Leu Pro Ala Met Lys Leu Val
580 585 590

Lys Tyr Tyr Ala Trp Glu Arg Phe Phe Glu Lys His Val Ala Asp Met
595 600 605

Arg Thr Arg Glu Arg His Tyr Met Phe Trp Asn Ala Val Val Lys Thr
610 615 620

Val Asn Val Thr Met Val Phe Gly Val Pro Pro Met Val Thr Phe Ala
625 630 635 640

Val Leu Val Pro Tyr Glu Leu Trp His Val Asp Ser Ser Thr Ser Glu
645 650 655

Pro Tyr Ile Lys Pro Gln Thr Ala Phe Thr Met Leu Ser Leu Phe Asn
660 665 670

Val Leu Arg Phe Pro Leu Val Val Leu Pro Lys Ala Met Arg Cys Val
675 680 685

Ser Glu Ala Leu Arg Ser Val Gly Asn Leu Glu Lys Phe Leu Ala Glu
690 695 700

Pro Val Ala Pro Arg Gln Asp Leu Glu Gly Lys Pro Gly Ala Gln Leu
705 710 715 720

Ser Lys Ala Val Leu Arg His Glu Met Asp Thr Ser Gly Phe Thr Leu
725 730 735

Arg Val Pro Glu Phe Ser Val Lys Ala Gly Glu Leu Val Ala Val Val
740 745 750

Gly Arg Val Gly Ala Gly Lys Ser Ser Ile Leu Gln Ala Met Leu Gly
755 760 765

Asn Met Gln Thr Ala Ser Gly Leu Ala Lys Cys Gln His Ser Ala Ser
770 775 780

Ser Cys Leu Pro Phe Leu Val Glu Gly Thr Ala His Ser Gly Gly Arg
785 790 795 800

Ile Ala Tyr Val Pro Gln Thr Ala Trp Cys Gln Asn Leu Ser Leu Arg
805 810 815

Asp Asn Ile Thr Phe Gly Gln Pro Trp Asp Glu Ala Lys Tyr Lys Gln

US 11,001,853 B2

209**210**

-continued

820	825	830
Val Ile His Ala Cys Ala Leu Glu Leu Asp Leu Ala Ile Leu Ala Ala		
835	840	845
Gly Asp Gln Ser Lys Ala Gly Leu Arg Gly Ile Asn Leu Ser Gly Gly		
850	855	860
Gln Arg Gln Arg Leu Asn Leu Ala Arg Cys Ala Tyr Phe Asp Gly Asp		
865	870	875
Leu Val Leu Leu Asp Asn Ala Leu Ser Ala Val Asp His His Thr Ala		
885	890	895
His His Ile Phe Glu His Cys Val Arg Gly Met Phe Arg Asp Lys Ala		
900	905	910
Thr Val Leu Val Thr His Gln Val Glu Phe Leu Pro Gln Cys Asp Lys		
915	920	925
Val Ala Ile Met Asp Asp Gly Thr Cys Val Tyr Phe Gly Pro Trp Asn		
930	935	940
Ala Ala Ala Gln Gln Leu Leu Ser Lys Tyr Leu Pro Ala Ser His Leu		
945	950	955
Leu Ala Ala Gly Gly Asn Ala Glu Gln Pro Arg Asp Thr Lys Lys Lys		
965	970	975
Val Val Lys Lys Glu Glu Thr Lys Lys Thr Glu Asp Ala Gly Lys Ala		
980	985	990
Lys Arg Val His Ser Ala Ser Leu Thr Leu Lys Ser Ala Leu Trp Glu		
995	1000	1005
Tyr Cys Trp Asp Ala Arg Trp Ile Ile Phe Cys Leu Ser Leu Phe		
1010	1015	1020
Phe Phe Leu Thr Ala Gln Ala Ser Arg Gln Leu Ala Asp Tyr Phe		
1025	1030	1035
Ile Arg Trp Trp Thr Arg Asp His Tyr Asn Lys Tyr Gly Val Leu		
1040	1045	1050
Cys Ile Asp Glu Gly Asp Asn Pro Cys Gly Pro Leu Phe Tyr Val		
1055	1060	1065
Gln Tyr Tyr Gly Ile Leu Gly Leu Leu Cys Phe Ile Val Leu Met		
1070	1075	1080
Ala Phe Arg Gly Ala Phe Leu Tyr Thr Trp Ser Leu Gly Ala Ser		
1085	1090	1095
Tyr Arg Gln His Glu Lys Ser Ile His Arg Val Leu Tyr Ala Pro		
1100	1105	1110
Leu Gly Phe Phe Leu Thr Thr Pro Val Gly Asp Leu Leu Val Ser		
1115	1120	1125
Phe Thr Lys Asp Gln Asp Val Met Asp Asp Ala Leu Pro Asp Ala		
1130	1135	1140
Leu Tyr Tyr Ala Gly Ile Tyr Gly Leu Ile Leu Leu Ala Thr Ala		
1145	1150	1155
Ile Thr Val Ser Val Thr Ile Pro Leu Phe Ser Ala Leu Ala Gly		
1160	1165	1170
Gly Leu Phe Val Val Ser Gly Ile Met Leu Ala Ile Tyr Leu Pro		
1175	1180	1185
Ala Ala Thr His Leu Lys Lys Leu Arg Met Gly Thr Ser Gly Asp		
1190	1195	1200
Val Val Thr Leu Ile Ala Glu Ala Leu Asp Gly Leu Gly Val Ile		
1205	1210	1215
Gln Ala Tyr Gly Lys Gln Ala Tyr Phe Thr Thr Ile Thr Ser Gln		
1220	1225	1230

-continued

Tyr Val Asn Asp Ala His Arg Ala Leu Phe Gly Ala Glu Ser Leu
1235 1240 1245

Asn Leu Trp Leu Ala Phe Ile Cys Asp Phe Phe Gly Ala Cys Met
1250 1255 1260

Val Leu Ser Val Ala Cys Phe Gly Ile Gly Gln Trp Ser Thr Leu
1265 1270 1275

Gly Ser Ser Ser Val Gly Leu Ala Phe Ser Gln Ser Ile Gln Met
1280 1285 1290

Leu Val Phe Tyr Thr Trp Ser Ile Arg Leu Val Ala Glu Cys Ile
1295 1300 1305

Gly Leu Phe Gly Ser Ala Glu Lys Ile Ala Trp Leu Ala Asn His
1310 1315 1320

Thr Pro Gln Glu Ala Gly Ser Leu Asp Pro Pro Ser Leu Pro Gly
1325 1330 1335

Ser Gly Glu Thr Lys Ala Ala Pro Lys Lys Arg Gly Thr Ala Gly
1340 1345 1350

Lys Phe Leu Pro Pro Leu Lys Asp Glu Asp Leu Ala Ile Val Pro
1355 1360 1365

Thr Gly Gly Pro Lys Leu Pro Ser Gly Trp Pro Arg Thr Gly Val
1370 1375 1380

Leu Glu Phe Asn Gln Val Val Met Lys Tyr Ala Pro His Leu Pro
1385 1390 1395

Pro Ala Leu Arg Gly Val Ser Phe Lys Val Lys Ser Gly Asp Lys
1400 1405 1410

Val Gly Val Val Gly Arg Thr Gly Ser Gly Lys Ser Thr Leu Leu
1415 1420 1425

Leu Ala Leu Tyr Arg Met Phe Asn Leu Glu Ser Gly Ala Ile Thr
1430 1435 1440

Leu Asp Gly Ile Asp Ile Ser Thr Leu Thr Leu Glu Gln Leu Arg
1445 1450 1455

Arg Gly Leu Ser Val Ile Pro Gln Glu Pro Thr Val Phe Ser Gly
1460 1465 1470

Thr Val Arg Thr Asn Leu Asp Pro Phe Gly Glu Phe Gly Ala Asp
1475 1480 1485

Ala Ile Leu Trp Glu Ala Leu Arg Asp Cys Gly Leu Glu Glu Gln
1490 1495 1500

Val Lys Ala Cys Gly Gly Leu Asp Ala Lys Leu Asp Gly Thr Gly
1505 1510 1515

Gly Asn Ala Trp Ser Ile Gly Gln Gln Gln Leu Met Cys Leu Ala
1520 1525 1530

Arg Ala Ala Leu Lys Lys Val Pro Val Leu Cys Leu Asp Glu Ala
1535 1540 1545

Thr Ala Ala Met Asp Pro His Thr Glu Ala His Val Leu Glu Ile
1550 1555 1560

Ile Glu Arg Ile Phe Ser Asp Arg Thr Met Leu Thr Ile Ala His
1565 1570 1575

Arg Leu Asp Asn Val Ile Arg Ser Asp Leu Val Val Val Met Asp
1580 1585 1590

Ala Gly Gln Val Cys Glu Met Gly Thr Pro Asp Glu Leu Leu Ala
1595 1600 1605

Asn Pro Gln Ser Ala Phe Ser Gln Leu Val Asp Lys Thr Gly Ala
1610 1615 1620

-continued

Ala Ser Ala Ala Ala Leu Arg Lys Met Ala Ala Asp Phe Leu Asp
 1625 1630 1635
 Glu Arg Ala Arg Gly Gln Lys Leu Gly Phe Lys Pro Arg Pro Ser
 1640 1645 1650
 Leu Glu Glu Ser His Ile Cys Val Ala Pro Ser Pro Ser Leu Ile
 1655 1660 1665
 Leu Ser Thr Leu Leu Phe Pro Pro Ala Phe Met Ala Asn Val Thr
 1670 1675 1680
 Ala Leu Leu Leu Pro Lys Pro Val Leu Ser His Ala Pro Val Ser
 1685 1690 1695
 Ser Gln Thr Val Asn Thr Tyr Ile Arg Leu Asn Ile Ile Gln Leu
 1700 1705 1710
 Gln Cys Asn Val Leu His Pro Ala Thr Lys Glu Ala Thr Trp Ser
 1715 1720 1725
 Ser Arg Arg Ile Thr Phe Thr Ala His Leu Ser Ser Ser Gly Ser
 1730 1735 1740
 Lys Pro Pro Pro Leu Pro Pro Leu Thr Glu Leu Pro Glu Gly
 1745 1750 1755
 Arg Gly Leu Asp Trp Ser Ser Ala Gly Tyr Arg Asp Gly Arg Glu
 1760 1765 1770
 Ala Ile Pro Ser Pro Ser Ala Lys Tyr Ser Ala Ala Asp Tyr Gly
 1775 1780 1785
 Ala Ala Gly Asp Gly Val Thr Asp Asp Thr Gln Ala Leu Gln Val
 1790 1795 1800
 Ala Val Ala Ala Ala His Glu Asp Asp Glu Gly Gly Val Val Tyr
 1805 1810 1815
 Leu Gly Ala Gly Thr Phe Val Leu Thr Gln Pro Leu Ser Ile Ala
 1820 1825 1830
 Gly Ser Asn Val Val Ile Arg Gly Ala Gly Glu Asp Ala Thr Thr
 1835 1840 1845
 Ile Phe Val Pro Leu Pro Leu Ser Asp Val Phe Pro Gly Thr Trp
 1850 1855 1860
 Ser Met Asp Ala Ser Gly Lys Val Thr Ser Pro Trp Ile Thr Arg
 1865 1870 1875
 Gly Gly Phe Leu Ala Phe Ser Gly Arg Arg Thr Lys Ser Ser Asp
 1880 1885 1890
 Ser Ser Thr Leu Leu Ala Thr Val Ala Gly Ser Val Glu Gln Gly
 1895 1900 1905
 Ala Ser Val Ile Pro Val Asp Ser Thr Ala Glu Phe Arg Leu Gly
 1910 1915 1920
 Gln Trp Val Arg Ile Ile Ile Asn Asp Ala Ser Thr Asp Ala Ser
 1925 1930 1935
 Ala Gly Gly Gly Thr Leu Glu Arg Gly Ser Ser Glu Val Gln Glu
 1940 1945 1950
 Ser Glu Thr Met Ile Ala Glu Gly Ala Thr Gly Gly Ala Gly
 1955 1960 1965
 Val Arg Ala Gln Trp Thr Gly Val Leu His Ala Phe Glu Pro Thr
 1970 1975 1980
 Val Gln Cys Ser Gly Val Glu Gln Leu Thr Ile Arg Phe Asn His
 1985 1990 1995
 Ser Met Met Ala Ala His Leu Ala Glu Arg Gly Tyr Asn Ala Ile
 2000 2005 2010
 Glu Leu Glu Asp Val Val Asp Cys Trp Ile Arg Gln Val Thr Ile

US 11,001,853 B2

215**216**

-continued

2015	2020	2025
Leu Asn Ala Asp Asn Ala Ile Arg Leu Arg Gly Thr Asp His Ser		
2030	2035	2040
Thr Leu Ser Gly Gln Ala Cys Ser Gly Gly Val Val Ala Val		
2045	2050	2055
Val Pro Val Trp Cys Arg Arg Gly Leu Pro Ser Pro Ala Asp Val		
2060	2065	2070
Thr Val Gly Val Thr Glu Leu Arg Trp Glu Pro Asp Thr Arg Glu		
2075	2080	2085
Val Asn Gly His His Ala Ile Thr Val Ser Lys Gly His Ala Asn		
2090	2095	2100
Leu Val Thr Arg Phe Arg Ile Thr Ala Pro Phe Tyr His Asp Ile		
2105	2110	2115
Ser Leu Glu Gly Gly Ala Leu Leu Asn Val Ile Ser Ser Gly Gly		
2120	2125	2130
Gly Ala Asn Leu Asn Leu Asp Leu His Arg Ser Gly Pro Trp Gly		
2135	2140	2145
Asn Leu Phe Ser Gln Leu Gly Met Gly Leu Ala Ala Arg Pro Phe		
2150	2155	2160
Asp Ala Gly Gly Arg Asp Gly Arg Gly Ala His Ala Gly Arg Gln		
2165	2170	2175
Asn Thr Phe Trp Asn Leu Gln Pro Gly Asp Val Ala Ala Ala Ala		
2180	2185	2190
Pro Ala Leu Gln Pro Ser Ala Ala Ala Gly Asp Ala Arg Arg Leu		
2195	2200	2205
Leu Val Asp Gly Asp Ser Leu Leu His Ala Gly Thr Gly Gln Ala		
2210	2215	2220
Arg Leu Leu Arg Gln Leu Glu Ala Asp Asp Ser Ala Glu Pro Leu		
2225	2230	2235
Leu Leu Pro Ser Cys Glu Phe Gly Pro Leu Leu Asn Phe Val Gly		
2240	2245	2250
Gly Phe Ala Gly Glu Leu Cys Lys Ser Ser Gly Trp Leu Val Ala		
2255	2260	2265
Gly Leu Pro Asp Asp Arg Pro Asp Leu His Ala Ser Gln Val Thr		
2270	2275	2280
Ala Arg Leu Gln His Gly Ala Ala Asp Asn Lys Thr His Ala		
2285	2290	2295

```

<210> SEQ ID NO: 81
<211> LENGTH: 373
<212> TYPE: PRT
<213> ORGANISM: Synechococcus PCC7942
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: PCC 7942

<400> SEQUENCE: 81

Met Asp Phe Leu Ser Asn Phe Leu Met Asp Phe Val Lys Gln Leu Gln
1 5 10 15

Ser Pro Thr Leu Ser Phe Leu Ile Gly Gly Met Val Ile Ala Ala Cys
20 25 30

Gly Ser Gln Leu Gln Ile Pro Glu Ser Ile Cys Lys Ile Ile Val Phe
35 40 45

Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly Met Ala Ile Arg Asn
50 55 60

```

US 11,001,853 B2

217

218

-continued

Ser Asn Leu Thr Glu Met Val Leu Pro Ala Leu Phe Ser Val Ala Ile
 65 70 75 80
 Gly Ile Leu Ile Val Phe Ile Ala Arg Tyr Thr Leu Ala Arg Met Pro
 85 90 95
 Lys Val Lys Thr Val Asp Ala Ile Ala Thr Gly Gly Leu Phe Gly Ala
 100 105 110
 Val Ser Gly Ser Thr Met Ala Ala Ala Leu Thr Leu Leu Glu Gln
 115 120 125
 Lys Ile Pro Tyr Glu Ala Trp Ala Gly Ala Leu Tyr Pro Phe Met Asp
 130 135 140
 Ile Pro Ala Leu Val Thr Ala Ile Val Val Ala Asn Ile Tyr Leu Asn
 145 150 155 160
 Lys Lys Lys Arg Lys Glu Ala Ala Phe Ala Ser Ala Gln Gly Ala Tyr
 165 170 175
 Ser Lys Gln Pro Val Ala Ala Gly Asp Tyr Ser Ser Ser Asp Tyr
 180 185 190
 Pro Ser Ser Arg Arg Glu Tyr Ala Gln Gln Glu Ser Gly Asp His Arg
 195 200 205
 Val Lys Ile Trp Pro Ile Val Glu Glu Ser Leu Gln Gly Pro Ala Leu
 210 215 220
 Ser Ala Met Leu Leu Gly Val Ala Leu Gly Leu Phe Ala Arg Pro Glu
 225 230 235 240
 Ser Val Tyr Glu Gly Phe Tyr Asp Pro Leu Phe Arg Gly Leu Leu Ser
 245 250 255
 Ile Leu Met Leu Val Met Gly Met Glu Ala Trp Ser Arg Ile Ser Glu
 260 265 270
 Leu Arg Lys Val Ala Gln Trp Tyr Val Val Tyr Ser Ile Val Ala Pro
 275 280 285
 Leu Ala His Gly Phe Ile Ala Phe Gly Leu Gly Met Ile Ala His Tyr
 290 295 300
 Ala Thr Gly Phe Ser Met Gly Gly Val Val Val Leu Ala Val Ile Ala
 305 310 315 320
 Ala Ser Ser Ser Asp Ile Ser Gly Pro Pro Thr Leu Arg Ala Gly Ile
 325 330 335
 Pro Ser Ala Asn Pro Ser Ala Tyr Ile Gly Ala Ser Thr Ala Ile Gly
 340 345 350
 Thr Pro Val Ala Ile Gly Ile Ala Ile Pro Leu Phe Leu Gly Leu Ala
 355 360 365
 Gln Thr Ile Gly Gly
 370

<210> SEQ ID NO: 82

<211> LENGTH: 374

<212> TYPE: PRT

<213> ORGANISM: Synechocystis PCC6803

<400> SEQUENCE: 82

Met Asp Phe Leu Ser Asn Phe Leu Thr Asp Phe Val Gly Gln Leu Gln
 1 5 10 15

Ser Pro Thr Leu Ala Phe Leu Ile Gly Gly Met Val Ile Ala Ala Leu
 20 25 30

Gly Thr Gln Leu Val Ile Pro Glu Ala Ile Ser Thr Ile Ile Val Phe
 35 40 45

Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly Met Ala Ile Arg Asn
 50 55 60

US 11,001,853 B2

219**220**

-continued

Ser Asn Leu Thr Glu Met Leu Leu Pro Val Ala Phe Ser Val Ile Leu
65 70 75 80

Gly Ile Leu Ile Val Phe Ile Ala Arg Phe Thr Leu Ala Lys Leu Pro
85 90 95

Asn Val Arg Thr Val Asp Ala Leu Ala Thr Gly Gly Leu Phe Gly Ala
100 105 110

Val Ser Gly Ser Thr Met Ala Ala Leu Thr Thr Leu Glu Glu Ser
115 120 125

Lys Ile Ser Tyr Glu Ala Trp Ala Gly Ala Leu Tyr Pro Phe Met Asp
130 135 140

Ile Pro Ala Leu Val Thr Ala Ile Val Val Ala Asn Ile Tyr Leu Asn
145 150 155 160

Lys Arg Lys Arg Lys Ser Ala Ala Ser Ile Glu Glu Ser Phe Ser
165 170 175

Lys Gln Pro Val Ala Ala Gly Asp Tyr Gly Asp Gln Thr Asp Tyr Pro
180 185 190

Arg Thr Arg Gln Glu Tyr Leu Ser Gln Gln Glu Pro Glu Asp Asn Arg
195 200 205

Val Lys Ile Trp Pro Ile Ile Glu Glu Ser Leu Gln Gly Pro Ala Leu
210 215 220

Ser Ala Met Leu Leu Gly Leu Ala Leu Gly Ile Phe Thr Lys Pro Glu
225 230 235 240

Ser Val Tyr Glu Gly Phe Tyr Asp Pro Leu Phe Arg Gly Leu Leu Ser
245 250 255

Ile Leu Met Leu Ile Met Gly Met Glu Ala Trp Ser Arg Ile Gly Glu
260 265 270

Leu Arg Lys Val Ala Gln Trp Tyr Val Val Tyr Ser Leu Ile Ala Pro
275 280 285

Ile Val His Gly Phe Ile Ala Phe Gly Leu Gly Met Ile Ala His Tyr
290 295 300

Ala Thr Gly Phe Ser Leu Gly Gly Val Val Val Leu Ala Val Ile Ala
305 310 315 320

Ala Ser Ser Ser Asp Ile Ser Gly Pro Pro Thr Leu Arg Ala Gly Ile
325 330 335

Pro Ser Ala Asn Pro Ser Ala Tyr Ile Gly Ser Ser Thr Ala Ile Gly
340 345 350

Thr Pro Ile Ala Ile Gly Val Cys Ile Pro Leu Phe Ile Gly Leu Ala
355 360 365

Gln Thr Leu Gly Ala Gly
370

<210> SEQ_ID NO 83
<211> LENGTH: 370
<212> TYPE: PRT
<213> ORGANISM: Nostoc sp. PCC 7120 (Anabaena sp. PCC 7120)

<400> SEQUENCE: 83

Met Asp Phe Phe Ser Leu Phe Leu Met Asp Phe Val Lys Gln Leu Gln
1 5 10 15

Ser Pro Thr Leu Gly Phe Leu Ile Gly Gly Met Val Ile Ala Ala Leu
20 25 30

Gly Ser Glu Leu Ile Ile Pro Glu Ala Ile Cys Gln Ile Ile Val Phe
35 40 45

Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly Ile Ala Ile Arg Asn

US 11,001,853 B2

221**222**

-continued

50	55	60
Ser Asn Leu Thr Glu Met Val Leu Pro Ala Ala Ser Ala Val Ala Val		
65	70	75
Gly Val Leu Val Val Phe Ile Ala Arg Tyr Thr Leu Ala Lys Leu Pro		
85	90	95
Lys Val Asn Thr Val Asp Ala Ile Ala Thr Gly Gly Leu Phe Gly Ala		
100	105	110
Val Ser Gly Ser Thr Met Ala Ala Ala Leu Thr Leu Leu Glu Gln		
115	120	125
Lys Ile Gln Tyr Glu Ala Trp Ala Ala Ala Leu Tyr Pro Phe Met Asp		
130	135	140
Ile Pro Ala Leu Val Thr Ala Ile Val Val Ala Asn Ile Tyr Leu Asn		
145	150	155
160		
Lys Lys Lys Arg Ser Ala Ala Gly Glu Tyr Leu Ser Lys Gln Ser Val		
165	170	175
Ala Ala Gly Glu Tyr Pro Asp Gln Gln Asp Tyr Pro Ser Ser Arg Gln		
180	185	190
Glu Tyr Leu Arg Lys Gln Gln Ser Ala Asp Asn Arg Val Lys Ile Trp		
195	200	205
Pro Ile Val Lys Glu Ser Leu Gln Gly Pro Ala Leu Ser Ala Met Leu		
210	215	220
Leu Gly Ile Ala Leu Gly Leu Phe Thr Gln Pro Glu Ser Val Tyr Lys		
225	230	235
240		
Ser Phe Tyr Asp Pro Leu Phe Arg Gly Leu Leu Ser Ile Leu Met Leu		
245	250	255
Val Met Gly Met Glu Ala Trp Ser Arg Ile Gly Glu Leu Arg Lys Val		
260	265	270
Ala Gln Trp Tyr Val Val Tyr Ser Val Val Ala Pro Leu Val His Gly		
275	280	285
Phe Ile Ala Phe Gly Leu Gly Met Ile Ala His Tyr Ala Thr Gly Phe		
290	295	300
Ser Leu Gly Gly Val Val Ile Leu Ala Val Ile Ala Ala Ser Ser Ser		
305	310	315
320		
Asp Ile Ser Gly Pro Pro Thr Leu Arg Ala Gly Ile Pro Ser Ala Asn		
325	330	335
Pro Ser Ala Tyr Ile Gly Ala Ser Thr Ala Ile Gly Thr Pro Ile Ala		
340	345	350
Ile Gly Leu Ala Ile Pro Leu Phe Leu Gly Leu Ala Gln Ala Ile Gly		
355	360	365
Gly Arg		
370		

<210> SEQ ID NO 84
<211> LENGTH: 377
<212> TYPE: PRT
<213> ORGANISM: Cyanothecce PCC 7425

<400> SEQUENCE: 84

Met Asp Phe Trp Ser Tyr Phe Leu Met Asp Phe Val Lys Gln Leu Gln		
1	5	10
15		

Ser Pro Thr Leu Gly Phe Leu Ile Gly Gly Met Val Ile Ala Ala Leu		
20	25	30

Gly Ser Gln Leu Val Ile Pro Glu Ala Ile Cys Gln Ile Ile Val Phe		
35	40	45

US 11,001,853 B2

223

224

-continued

Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly Met Ala Ile Arg Asn
 50 55 60

Ser Asn Leu Thr Glu Met Val Leu Pro Ala Ala Phe Ser Val Ile Ser
 65 70 75 80

Gly Ile Leu Ile Val Phe Ile Ala Arg Tyr Thr Leu Ala Lys Leu Pro
 85 90 95

Lys Val Arg Thr Val Asp Ala Ile Ala Thr Gly Gly Leu Phe Gly Ala
 100 105 110

Val Ser Gly Ser Thr Met Ala Ala Leu Thr Leu Leu Glu Glu Glu
 115 120 125

Lys Ile Pro Tyr Glu Ala Trp Ala Gly Ala Leu Tyr Pro Phe Met Asp
 130 135 140

Ile Pro Ala Leu Val Thr Ala Ile Val Ile Ala Asn Ile Tyr Leu Asn
 145 150 155 160

Lys Lys Lys Arg Arg Ala Glu Ser Glu Ala Leu Ser Lys Gln Glu Tyr
 165 170 175

Leu Gly Lys Gln Ser Ile Val Ala Gly Asp Tyr Pro Ala Gln Gln Asp
 180 185 190

Tyr Pro Ser Thr Arg Gln Glu Tyr Leu Ser Lys Gln Gln Gly Pro Glu
 195 200 205

Asn Asn Arg Val Lys Ile Trp Pro Ile Val Gln Glu Ser Leu Gln Gly
 210 215 220

Pro Ala Leu Ser Ala Met Leu Leu Gly Val Ala Leu Gly Ile Leu Thr
 225 230 235 240

Lys Pro Glu Ser Val Tyr Glu Ser Phe Tyr Asp Pro Leu Phe Arg Gly
 245 250 255

Leu Leu Ser Ile Leu Met Leu Val Met Gly Met Glu Ala Trp Ser Arg
 260 265 270

Ile Gly Glu Leu Arg Lys Val Ala Gln Trp Tyr Val Val Tyr Ser Val
 275 280 285

Val Ala Pro Phe Val His Gly Leu Ile Ala Phe Gly Leu Gly Met Phe
 290 295 300

Ala His Tyr Thr Met Gly Phe Ser Met Gly Gly Val Val Val Leu Ala
 305 310 315 320

Val Ile Ala Ser Ser Ser Asp Ile Ser Gly Pro Pro Thr Leu Arg
 325 330 335

Ala Gly Ile Pro Ser Ala Asn Pro Ser Ala Tyr Ile Gly Ala Ser Thr
 340 345 350

Ala Ile Gly Thr Pro Ile Ala Ile Gly Leu Cys Ile Pro Phe Phe Ile
 355 360 365

Gly Leu Ala Gln Thr Leu Gly Gly Gly
 370 375

<210> SEQ ID NO 85

<211> LENGTH: 373

<212> TYPE: PRT

<213> ORGANISM: *Microcystis aeruginosa*

<400> SEQUENCE: 85

Met Asp Phe Phe Ser Leu Phe Val Met Asp Phe Ile Gln Gln Leu Gln
 1 5 10 15

Ser Pro Thr Leu Ala Phe Leu Ile Gly Gly Met Ile Ile Ala Ala Leu
 20 25 30

Gly Ser Glu Leu Val Ile Pro Glu Ser Ile Cys Thr Ile Ile Val Phe
 35 40 45

-continued

Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly Ile Ala Ile Arg Asn
 50 55 60
 Ser Asn Leu Thr Glu Met Val Leu Pro Met Ile Phe Ala Val Ile Val
 65 70 75 80
 Gly Ile Ile Val Val Phe Val Ala Arg Tyr Thr Leu Ala Asn Leu Pro
 85 90 95
 Lys Val Lys Val Val Asp Ala Ile Ala Thr Gly Gly Leu Phe Gly Ala
 100 105 110
 Val Ser Gly Ser Thr Met Ala Ala Gly Leu Thr Val Leu Glu Glu Gln
 115 120 125
 Lys Ile Pro Tyr Glu Ala Trp Ala Gly Ala Leu Tyr Pro Phe Met Asp
 130 135 140
 Ile Pro Ala Leu Val Thr Ala Ile Val Val Ala Asn Ile Tyr Leu Asn
 145 150 155 160
 Lys Lys Lys Gln Lys Glu Ala Ala Tyr Asp Gln Glu Ser Phe Ser Lys
 165 170 175
 Gln Pro Val Ala Ala Gly Asn Tyr Ser Asp Gln Gln Asp Tyr Pro Ser
 180 185 190
 Ser Arg Gln Glu Tyr Leu Ser Gln Gln Pro Ala Asp Asn Arg Val
 195 200 205
 Lys Ile Trp Pro Ile Ile Glu Glu Ser Leu Arg Gly Pro Ala Leu Ser
 210 215 220
 Ala Met Leu Leu Gly Leu Ala Leu Gly Ile Phe Thr Gln Pro Glu Ser
 225 230 235 240
 Val Tyr Lys Ser Phe Tyr Asp Pro Leu Phe Arg Gly Leu Leu Ser Val
 245 250 255
 Leu Met Leu Val Met Gly Met Glu Ala Trp Ser Arg Val Gly Glu Leu
 260 265 270
 Arg Lys Val Ala Gln Trp Tyr Val Val Tyr Ser Val Ile Ala Pro Phe
 275 280 285
 Val His Gly Leu Ile Ala Phe Gly Leu Gly Met Ile Ala His Tyr Ala
 290 295 300
 Thr Gly Phe Ser Trp Gly Gly Val Val Met Leu Ala Val Ile Ala Ser
 305 310 315 320
 Ser Ser Ser Asp Ile Ser Gly Pro Pro Thr Leu Arg Ala Gly Ile Pro
 325 330 335
 Ser Ala Asn Pro Ser Ala Tyr Ile Gly Ala Ser Thr Ala Ile Gly Thr
 340 345 350
 Pro Val Ala Ile Gly Leu Cys Ile Pro Phe Phe Val Gly Leu Ala Gln
 355 360 365
 Ala Leu Ser Gly Gly
 370

<210> SEQ ID NO 86
 <211> LENGTH: 369
 <212> TYPE: PRT
 <213> ORGANISM: Anabaena variabilis

<400> SEQUENCE: 86

Met Asp Phe Val Ser Leu Phe Val Lys Asp Phe Ile Ala Gln Leu Gln
 1 5 10 15
 Ser Pro Thr Leu Ala Phe Leu Ile Gly Gly Met Ile Ile Ala Ala Leu
 20 25 30
 Gly Ser Glu Leu Val Ile Pro Glu Ser Ile Cys Thr Ile Ile Val Phe

US 11,001,853 B2

227

228

-continued

35	40	45
Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly	Ile Ala Ile Arg Asn	
50	55	60
Ser Asn Leu Thr Glu Met Val Leu Pro Met Ile Phe Ala Val Ile Thr		
65	70	75
Gly Ile Thr Ile Val Phe Ile Ser Arg Tyr Thr Leu Ala Lys Leu Pro		
85	90	95
Lys Val Lys Val Val Asp Ala Ile Ala Thr Gly Gly Leu Phe Gly Ala		
100	105	110
Val Ser Gly Ser Thr Met Ala Ala Gly Leu Thr Val Leu Glu Glu Gln		
115	120	125
Lys Met Ala Tyr Glu Ala Trp Ala Gly Ala Leu Tyr Pro Phe Met Asp		
130	135	140
Ile Pro Ala Leu Val Thr Ala Ile Val Ile Asn Ile Tyr Leu Asn		
145	150	155
Lys Lys Lys Arg Lys Glu Ala Val Tyr Ser Thr Glu Gln Pro Val Ala		
165	170	175
Ala Gly Asp Tyr Pro Asp Gln Lys Asp Tyr Pro Ser Ser Arg Gln Glu		
180	185	190
Tyr Leu Ser Gln Gln Lys Gly Asp Glu Asp Asn Arg Val Lys Ile Trp		
195	200	205
Pro Ile Ile Glu Glu Ser Leu Arg Gly Pro Ala Leu Ser Ala Met Leu		
210	215	220
Leu Gly Leu Ala Leu Gly Leu Phe Thr Gln Pro Glu Ser Val Tyr Lys		
225	230	235
Ser Phe Tyr Asp Pro Ala Phe Arg Gly Leu Leu Ser Ile Leu Met Leu		
245	250	255
Val Met Gly Met Glu Ala Trp Ser Arg Ile Gly Glu Leu Arg Lys Val		
260	265	270
Ala Gln Trp Tyr Val Val Tyr Ser Val Val Ala Pro Phe Val His Gly		
275	280	285
Leu Ile Ala Phe Gly Leu Gly Met Ile Ala His Tyr Thr Met Asn Phe		
290	295	300
Ser Met Gly Gly Val Val Ile Leu Ala Val Ile Ala Ser Ser Ser Ser		
305	310	315
Asp Ile Ser Gly Pro Pro Thr Leu Arg Ala Gly Ile Pro Ser Ala Asn		
325	330	335
Pro Ser Ala Tyr Ile Gly Ala Ser Thr Ala Val Gly Thr Pro Val Ala		
340	345	350
Ile Gly Leu Cys Ile Pro Phe Phe Leu Gly Leu Ala Gln Ala Ile Gly		
355	360	365

GLY

<210> SEQ ID NO 87

<211> LENGTH: 366

<212> TYPE: PRT

<213> ORGANISM: Cyanothecce

<400> SEQUENCE: 87

Met Asp Phe Leu Ser Leu Phe Val Lys Asp Phe Ile Ile Gln Leu Gln		
1	5	10
15		

Ser Pro Thr Leu Ala Phe Leu Ile Gly Gly Met Val Ile Ala Ala Leu		
20	25	30

Gly Ser Glu Leu Val Ile Pro Glu Ser Ile Cys Thr Ile Ile Val Phe

US 11,001,853 B2

229**230**

-continued

35	40	45
Met Leu Leu Thr Lys Ile Gly Leu Thr Gly Gly	Ile Ala Ile Arg Asn	
50	55	60
Ser Asn Leu Thr Glu Met Val Leu Pro Met Ile Cys Ala Val Ile Val		
65	70	75
Gly Ile Val Val Val Phe Ile Ala Arg Tyr Thr Leu Ala Lys Leu Pro		
85	90	95
Lys Val Asn Val Val Asp Ala Ile Ala Thr Gly Gly Leu Phe Gly Ala		
100	105	110
Val Ser Gly Ser Thr Met Ala Ala Gly Leu Thr Val Leu Glu Glu Gln		
115	120	125
Lys Ile Pro Tyr Glu Ala Trp Ala Gly Ala Leu Tyr Pro Phe Met Asp		
130	135	140
Ile Pro Ala Leu Val Thr Ala Ile Val Val Ala Asn Ile Tyr Leu Asn		
145	150	155
Lys Lys Lys Arg Lys Ala Thr Val Met Gln Glu Ser Leu Ser Lys Gln		
165	170	175
Pro Val Ala Ala Gly Asp Tyr Pro Ser Ser Arg Gln Glu Tyr Val Ser		
180	185	190
Gln Gln Gln Pro Glu Asp Asn Arg Val Lys Ile Trp Pro Ile Ile Glu		
195	200	205
Glu Ser Leu Arg Gly Pro Ala Leu Ser Ala Met Leu Leu Gly Leu Ala		
210	215	220
Leu Gly Ile Leu Thr Gln Pro Glu Ser Val Tyr Lys Gly Phe Tyr Asp		
225	230	235
Pro Pro Phe Arg Gly Leu Leu Ser Ile Leu Met Leu Val Met Gly Met		
245	250	255
Glu Ala Trp Ser Arg Ile Gly Glu Leu Arg Lys Val Ala Gln Trp Tyr		
260	265	270
Val Val Tyr Ser Val Ala Ala Pro Phe Ile His Gly Leu Leu Ala Phe		
275	280	285
Gly Leu Gly Met Ile Ala His Tyr Thr Met Gly Phe Ser Met Gly Gly		
290	295	300
Val Val Ile Leu Ala Val Ile Ala Ser Ser Ser Ser Asp Ile Ser Gly		
305	310	315
Pro Pro Thr Leu Arg Ala Gly Ile Pro Ser Ala Asn Pro Ser Ala Tyr		
325	330	335
Ile Gly Ala Ser Thr Ala Ile Gly Thr Pro Val Ala Ile Gly Leu Cys		
340	345	350
Ile Pro Phe Phe Val Gly Leu Ala Gln Ala Ile Gly Gly Phe		
355	360	365

<210> SEQ ID NO 88

<211> LENGTH: 378

<212> TYPE: PRT

<213> ORGANISM: Arthrospira platensis str. Paraca

<400> SEQUENCE: 88

Met Asp Phe Leu Ser Gly Phe Leu Thr Arg Phe Leu Ala Gln Leu Gln		
1	5	10
		15

Ser Pro Thr Leu Gly Phe Leu Ile Gly Gly Met Val Ile Ala Ala Val		
20	25	30

Asn Ser Gln Leu Gln Ile Pro Asp Ala Ile Tyr Lys Phe Val Val Phe		
35	40	45

-continued

Met Leu Leu Met Lys Val Gly Leu Ser Gly Gly Ile Ala Ile Arg Gly
 50 55 60
 Ser Asn Leu Thr Glu Met Leu Leu Pro Ala Val Phe Ala Leu Val Thr
 65 70 75 80
 Gly Ile Val Ile Val Phe Ile Gly Arg Tyr Thr Leu Ala Lys Leu Pro
 85 90 95
 Asn Val Lys Thr Val Asp Ala Ile Ala Thr Ala Gly Leu Phe Gly Ala
 100 105 110
 Val Ser Gly Ser Thr Met Ala Ala Ala Leu Thr Leu Leu Glu Glu Gln
 115 120 125
 Gly Met Glu Tyr Glu Ala Trp Ala Ala Ala Leu Tyr Pro Phe Met Asp
 130 135 140
 Ile Pro Ala Leu Val Ser Ala Ile Val Leu Ala Ser Ile Tyr Val Ser
 145 150 155 160
 Lys Gln Lys His Ser Asp Met Ala Asp Glu Ser Leu Ser Lys His Glu
 165 170 175
 Ser Leu Ser Lys Gln Pro Val Ala Ala Gly Asp Tyr Pro Ser Lys Pro
 180 185 190
 Glu Tyr Pro Thr Thr Arg Gln Glu Tyr Leu Ser Gln Gln Arg Gly Ser
 195 200 205
 Ala Asn Gln Gly Val Glu Ile Trp Pro Ile Ile Lys Glu Ser Leu Gln
 210 215 220
 Gly Ser Ala Leu Ser Ala Leu Leu Gly Leu Ala Leu Gly Leu Leu
 225 230 235 240
 Thr Arg Pro Glu Ser Val Phe Gln Ser Phe Tyr Glu Pro Leu Phe Arg
 245 250 255
 Gly Leu Leu Ser Ile Leu Met Leu Val Met Gly Met Glu Ala Thr Ala
 260 265 270
 Arg Leu Gly Glu Leu Arg Lys Val Ala Gln Trp Tyr Ala Val Tyr Ala
 275 280 285
 Phe Ile Ala Pro Leu Leu His Gly Leu Ile Ala Phe Gly Leu Gly Met
 290 295 300
 Ile Ala His Val Val Thr Gly Phe Ser Leu Gly Gly Val Val Ile Leu
 305 310 315 320
 Ala Val Ile Ala Ser Ser Ser Asp Ile Ser Gly Pro Pro Thr Leu
 325 330 335
 Arg Ala Gly Ile Pro Ser Ala Asn Pro Ser Ala Tyr Ile Gly Ser Ser
 340 345 350
 Thr Ala Val Gly Thr Pro Val Ala Ile Ala Leu Gly Ile Pro Leu Tyr
 355 360 365
 Ile Gly Leu Ala Gln Ala Leu Met Gly Gly
 370 375

<210> SEQ ID NO 89

<211> LENGTH: 336

<212> TYPE: PRT

<213> ORGANISM: Chlamydomonas reinhardtii

<400> SEQUENCE: 89

Met Gln Thr Thr Met Thr Arg Pro Cys Leu Ala Gln Pro Val Leu Arg
 1 5 10 15
 Ser Arg Val Leu Arg Ser Pro Met Arg Val Val Ala Ala Ser Ala Pro
 20 25 30
 Thr Ala Val Thr Thr Val Val Thr Ser Asn Gly Asn Gly Asn Gly His
 35 40 45

-continued

Phe Gln Ala Ala Thr Thr Pro Val Pro Pro Thr Pro Ala Pro Val Ala
50 55 60

Val Ser Ala Pro Val Arg Ala Val Ser Val Leu Thr Pro Pro Gln Val
65 70 75 80

Tyr Glu Asn Ala Ile Asn Val Gly Ala Tyr Lys Ala Gly Leu Thr Pro
85 90 95

Leu Ala Thr Phe Val Gln Gly Ile Gln Ala Gly Ala Tyr Ile Ala Phe
100 105 110

Gly Ala Phe Leu Ala Ile Ser Val Gly Gly Asn Ile Pro Gly Val Ala
115 120 125

Ala Ala Asn Pro Gly Leu Ala Lys Leu Leu Phe Ala Leu Val Phe Pro
130 135 140

Val Gly Leu Ser Met Val Thr Asn Cys Gly Ala Glu Leu Phe Thr Gly
145 150 155 160

Asn Thr Met Met Leu Thr Cys Ala Leu Ile Glu Lys Lys Ala Thr Trp
165 170 175

Gly Gln Leu Leu Lys Asn Trp Ser Val Ser Tyr Phe Gly Asn Phe Val
180 185 190

Gly Ser Ile Ala Met Val Ala Ala Val Val Ala Thr Gly Cys Leu Thr
195 200 205

Thr Asn Thr Leu Pro Val Gln Met Ala Thr Leu Lys Ala Asn Leu Gly
210 215 220

Phe Thr Glu Val Leu Ser Arg Ser Ile Leu Cys Asn Trp Leu Val Cys
225 230 235 240

Cys Ala Val Trp Ser Ala Ser Ala Ala Thr Ser Leu Pro Gly Arg Ile
245 250 255

Leu Ala Leu Trp Pro Cys Ile Thr Ala Phe Val Ala Ile Gly Leu Glu
260 265 270

His Ser Val Ala Asn Met Phe Val Ile Pro Leu Gly Met Met Leu Gly
275 280 285

Ala Glu Val Thr Trp Ser Gln Phe Phe Asn Asn Leu Ile Pro Val
290 295 300

Thr Leu Gly Asn Thr Ile Ala Gly Val Leu Met Met Ala Ile Ala Tyr
305 310 315 320

Ser Ile Ser Phe Gly Ser Leu Gly Lys Ser Ala Lys Pro Ala Thr Ala
325 330 335

<210> SEQ_ID NO 90

<211> LENGTH: 337

<212> TYPE: PRT

<213> ORGANISM: Volvox carteri

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: f. nagariensis

<400> SEQUENCE: 90

Met Gln Thr Thr Met Ser Val Thr Arg Pro Cys Val Gly Leu Arg Pro
1 5 10 15

Leu Pro Val Arg Asn Val Arg Ser Leu Ile Arg Ala Gln Ala Ala Pro
20 25 30

Gln Gln Val Ser Thr Ala Val Ser Thr Asn Gly Asn Gly Asn Gly Val
35 40 45

Ala Ala Ala Ser Leu Ser Val Pro Ala Pro Val Ala Ala Pro Ala Gln
50 55 60

Ala Val Ser Thr Pro Val Arg Ala Val Ser Val Leu Thr Pro Pro Gln

US 11,001,853 B2

235**236**

-continued

65	70	75	80
Val	Tyr	Glu	Asn
Ala	Ala	Ala	Asn
Val	Gly	Ala	Tyr
85	90	95	
Leu	Ala	Thr	Phe
100	105	110	
Leu	Ala	Gln	Gly
		Ile	Gln
		Ala	Gly
		Tyr	Ile
		Ala	
Phe	Gly	Ala	Phe
115	120	125	
Leu	Leu	Ala	Cys
		Ser	Val
		Gly	Gly
		Asn	Ile
		Pro	Gly
		Ile	
Thr	Ala	Ser	Asn
130	135	140	
Pro	Val	Gly	Leu
		Ser	Ser
		Met	Val
		Thr	Asn
		Cys	Gly
		Ala	Glu
		Leu	Tyr
145	150	155	160
Gly	Asn	Thr	Met
165	170	175	
Met	Leu	Thr	Cys
		Ala	Ile
		Phe	Glu
		Lys	Lys
		Ala	Thr
Trp	Ala	Gln	Leu
180	185	190	
Leu	Val	Lys	Asn
		Trp	Val
		Val	Ser
		Tyr	Ala
		Gly	Asn
		Phe	
Val	Gly	Ser	Ile
195	200	205	
Ala	Ser	Asn	Gln
210	215	220	
Leu	Pro	Val	Asn
			Met
			Ala
			Thr
			Gly
			Leu
			Met
Ala	Ser	Asn	Gln
225	230	235	240
Phe	Thr	Glu	Val
			Leu
			Ser
			Arg
Cys	Cys	Ala	Val
245	250	255	
Trp	Ser	Ala	Ser
			Ala
			Thr
			Ser
			Leu
			Pro
			Gly
			Arg
Ile	Leu	Gly	Leu
260	265	270	
Trp	Pro	Pro	Ile
			Thr
			Ala
			Phe
			Val
			Ala
			Ile
			Gly
			Leu
Glu	His	Ser	Val
275	280	285	
Ala	Asn	Met	Phe
			Val
			Ile
			Pro
			Leu
			Gly
			Met
			Met
			Leu
Gly	Ala	Asp	Val
290	295	300	
Thr	Trp	Trp	Ser
			Gln
			Phe
			Phe
			Asn
			Asn
			Leu
			Val
			Pro
Val	Thr	Leu	Gly
305	310	315	320
Asn	Thr	Ile	Ala
			Gly
			Val
			Met
			Met
			Ala
			Val
			Ala
Tyr	Ser	Val	Ser
325	330	335	
Tyr	Gly	Ser	Leu
			Gly
			Lys
			Thr
			Pro
			Lys
			Pro
			Ala
			Thr

Ala

<210> SEQ ID NO 91
<211> LENGTH: 3249
<212> TYPE: DNA
<213> ORGANISM: Engineered construct (codon optimized gene)

<400> SEQUENCE: 91

```

atgcgtcccg gcctggcggt ctcctgtcg gtgctgcccc tgcgtacta cttcggtac 60
aagatcgtgc agatcaagct gcagaacgcc aagcacgtcg ccctgcgcctc cgccatcatg 120
caggaggtgc tgccgcctat caagctggtc aagtactacg cctggggagca gttcttgag 180
aaccagatca gcaagggtccg ccggggaggag atccgcctca acttctggaa ctgcgtatg 240
aagggtcatca acgtggcctg cgtgttctgc gtgcgcctca tgaccgcctt cgtcatctc 300
accacccatcg agttccagcg cgccgcctg gtgtccagcg tcgccttac caccctgtcg 360
ctgttcaaca ttctgcgttt cccctggtc gtgctgcccc agggccctgcg tgccgtgtcc 420
gaggccaacg cgtctctcca gcgcgcggag gcctacgtgc tggaggaggt gcccctgggc 480
actgcccggc tcaagacccc caagaacgct ccccccggcg ccgtcatcga gaacgggttg 540
ttccaccacc cttccaaaccc caactggcac ctgcacgtgc ccaagttcga ggtcaagccc 600

```

-continued

ggccaggctcg ttgctgtggt	ggccgcatac gcccggca agtcgtccct	660
atcctcggca acatggtaa	ggagcacggc agcttcaacg tggcgcccg	720
gtgccgcaga acccctggct	gcagaacctg tccctgcgtg acaacgtgt	780
cagttcgatg agaacaagta	caccgacgtc atcgagtcc	840
cagatcctgt ccaacggtaa	ccagtcuaag gccggcatcc	900
ggccagcgc agcgegtgaa	cctggccgc tgcgctacg	960
ctcgacaacg ccctgtccgc	cgtggaccac cacaccgccc	1020
atcaaggccc tggctccga	caaggccgtg gtgctggta	1080
ccccgtgcg acaacgtggc	catcatggac gagggccgt	1140
aacgaggagg occagcacct	gctggcaag ctgctgccc	1200
gccccgtccc aggaggtccc	tcacccaccc	1260
cagaagtcgc agtcgtgca	gctgacccctg	1320
aagcccaagg acgtccagaa	gctgactgac	1380
ggcaacctgt tccctgggt	cgtgtgttc	1440
cagatctccg atttctgggt	gctgggtgg	1500
aaggggagc aggactcgcc	cgccaccacc	1560
ggcctgttct acatcttcat	gatcttccgc	1620
tcctcgaga ccateccgcag	gaaggccctg	1680
ttcctggta cgccggcgtgg	cgacctgtcg	1740
gtgagaacc tgcccgatgc	cgttacttc	1800
accaccatca ccgtgtccgt	caccatcaac	1860
atcatgaccc tcatacatgt	ctccatctac	1920
cgccgcgtgt ctggggcat	gctggtcggc	1980
gtgggtttagg cttcaacaa	gcaggagtag	2040
atcaccaact ccgcgtctt	caacgcccgg	2100
gacttcatcg gcgcgtgcct	ggtgccgtgt	2160
gacctggcgccg	cgccgtggcc	2220
tacacctggg tggccgttt	catctccgag	2280
atggccctacc tcgcccacta	cgtgccccac	2340
gacggcgtcg ccaagcaaat	cgtccgtccc	2400
gtccaggctcg tgggtgacga	cgccgccttc	2460
ttcgaggacg tgtggatgca	gtaccggctg	2520
ttcaagatca acgacggcga	gaagggtcgcc	2580
accacgtgc tggcgctgt	ccgcatgttc	2640
ggcgtggaca tggccaccc	gtcgctcaag	2700
caggagcccg tcatgttac	cgccacccgt	2760
aaggacgtat	ccattctgtg	2820
cacgcggcgcc	gcctggacgg	2880
cagatgcagc tgggtgtcc	ggctcgccgc	2940

US 11,001,853 B2

239

240

-continued

gacgaggcta ccgcggccat ggaccgcac actgaggcca tcgtgcagca gaccatcaa	3000
aagggtttcg acgaccgcac caccatcacc attgcccacc gcctggacac catcatcgag	3060
tccgacaaga tcategtatgg ggagcaggc tcgctgtatgg agtacgagtc gcccctgaa	3120
ctgctcgcca accgegactc catgttctcc aagctggtcg acaagaccgg cccccccgccc	3180
gccgctgcgc tgcgcaagat ggccgaggac ttctggtcca ctgcgtccgc gcagggccgc	3240
aaccagtaa	3249

<210> SEQ ID NO 92
<211> LENGTH: 1008
<212> TYPE: DNA
<213> ORGANISM: Engineered construct (codon optimized gene)

<400> SEQUENCE: 92

atgcagacca ctatgactcg cccttgcattt gcccagcccg tgctgcgatc tcgtgtgtc	60
cggtcgccta tgcgggttgt tgcagcgacg gctccttaccc cggtgacgac agtctgtgacc	120
tcaaatggaa atggcaacgg tcatttccaa gctgctacta cgcgggtgcc ccctactccc	180
gtctccgtcg ctgtttccgc gcctgtgcgc gctgtgtcg tgctgactcc tcctcaagt	240
tatgagaacg ccattaatgt tggcgcctac aaggccgggc taacgcctct ggcaacgttt	300
gtccaggggca tccaagccgg tgcctacattt gcgttcggc ctttcctcgc catctccgt	360
ggaggcaaca tccccggcgt cgccggccgc aacccggcc tggcaagct gctatttgct	420
ctgggtttcc cctgtgggtct gtccatggtg accaactgcg gcccggagct gttcacggc	480
aacaccatga tgctcacatg cgcgcatac gagaagaagg ccacttgggg gcagcttctg	540
aagaactgga gcgtgtccta ctccggcaac ttctgggtctt ccattgcctt ggtcgccgc	600
gtgggtggcca cccggcgcctt gaccaccaac accctgcctt tgcaatggc caccctcaag	660
gccaacctgg gtttacccga ggtgtgtcg cgctccatcc tggcaactg gctgggtgtc	720
tgcggccgtgt ggtccgcctc cgccggccacc tcgctggccg gcccgcattt ggcgtgtgg	780
ccctgcatac cccgccttcgtt ggccatggc ctggagactt ccgtcgccaa catgttcgt	840
attccctctgg gcatgatgtt gggcgcttagt gtcacgtggaa gccagttttt tttcaacaac	900
ctgatccccg tcaccctggg caacaccattt gctggcgatc tcatgatggc catgcctac	960
tccatctcg tccggccctt cggcaagttcc gccaaggcccg ccaccgcg	1008

<210> SEQ ID NO 93
<211> LENGTH: 148
<212> TYPE: PRT
<213> ORGANISM: Arabidopsis thaliana
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: Ferredoxin1

<400> SEQUENCE: 93

Met Ala Ser Thr Ala Leu Ser Ser Ala Ile Val Ser Thr Ser Phe Leu			
1	5	10	15

Arg Arg Gln Gln Thr Pro Ile Ser Leu Arg Ser Leu Pro Phe Ala Asn		
20	25	30

Thr Gln Ser Leu Phe Gly Leu Lys Ser Ser Thr Ala Arg Gly Gly Arg		
35	40	45

Val Thr Ala Met Ala Thr Tyr Lys Val Lys Phe Ile Thr Pro Glu Gly		
50	55	60

Glu Gln Glu Val Glu Cys Glu Glu Asp Val Tyr Val Leu Asp Ala Ala			
65	70	75	80

-continued

Glu Glu Ala Gly Leu Asp Leu Pro Tyr Ser Cys Arg Ala Gly Ser Cys
 85 90 95
 Ser Ser Cys Ala Gly Lys Val Val Ser Gly Ser Ile Asp Gln Ser Asp
 100 105 110
 Gln Ser Phe Leu Asp Asp Glu Gln Met Ser Glu Gly Tyr Val Leu Thr
 115 120 125
 Cys Val Ala Tyr Pro Thr Ser Asp Val Val Ile Glu Thr His Lys Glu
 130 135 140
 Glu Ala Ile Met
 145

<210> SEQ ID NO 94
 <211> LENGTH: 783
 <212> TYPE: DNA
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 94

atgtcccatc	actgggggta	cgccaaacac	aacggacctg	agcaactggca	taaggacttc	60
cccattgc	ccggagagcg	ccagtccc	gttgacatcg	acactcatac	agccaagtat	120
gacccttccc	tgaagccc	ctgtgtttcc	tatgatcaag	caactccct	gaggatcc	180
aacaatggtc	atgc	tttcaa	cgtggaggtt	gatgactctc	aggacaaagc	240
ggaggacccc	ttgatggcac	tttacagattt	attcagttt	actttcactg	gggttca	300
gtatggacaag	gttca	gagca	tactgtggat	aaaaagaaat	atgctgcaga	360
gttca	actgg	acaccaata	tgggat	tttggaaagctg	tgcagcaacc	420
gccgttctag	gtat	ttttt	gaagg	tttggc	agcgcta	480
gtatgtgtgg	tttcc	attttaaa	aacaagg	gc	tttca	540
cgtggcctcc	ttc	cttgaatc	tttggattt	tttggac	tttacc	600
cctcttctgg	aat	gtgtgac	tttggattt	ctcaagg	aaacccgt	660
cagggttgta	aattccgtaa	acttaactt	aatggggagg	gtgaacccga	agaactgtat	720
gttggacaact	ggcgcccc	tttggact	tttggact	aaatcaa	aggc	780
taa						783

<210> SEQ ID NO 95
 <211> LENGTH: 148
 <212> TYPE: PRT
 <213> ORGANISM: Arabidopsis thaliana
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: Ferredoxin2(thale cress)

<400> SEQUENCE: 95

Met Ala Ser Thr	Ala Leu Ser Ser	Ala Ile Val	Gly Thr Ser	Phe Ile		
1	5	10	15			
Arg Arg Ser Pro	Ala Pro Ile Ser	Leu Arg Ser	Leu Pro	Ser Ala Asn		
20	25	30				
Thr Gln Ser	Leu Phe Gly	Leu Lys Ser	Gly Thr	Ala Arg Gly	Gly Arg	
35	40	45				
Val Thr Ala Met	Ala Thr Tyr	Lys Val Lys	Phe Ile	Thr Pro	Glu Gly	
50	55	60				
Glu Leu Glu Val	Glu Cys Asp Asp	Asp Val	Tyr Val	Leu Asp	Ala Ala	
65	70	75	80			
Glu Glu Ala Gly	Ile Asp Leu Pro	Tyr Ser	Cys Arg	Ala Gly	Ser Cys	

-continued

85	90	95
----	----	----

Ser Ser Cys Ala Gly Lys Val Val Ser Gly Ser Val Asp Gln Ser Asp
100 105 110

Gln Ser Phe Leu Asp Asp Glu Gln Ile Gly Glu Gly Phe Val Leu Thr
115 120 125

Cys Ala Ala Tyr Pro Thr Ser Asp Val Thr Ile Glu Thr His Lys Glu
130 135 140

Glu Asp Ile Val
145

<210> SEQ_ID NO 96

<211> LENGTH: 253

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana (thale cress)

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: ferredoxin-NADP(+)oxidoreductase(FNR1)

<400> SEQUENCE: 96

Phe Thr Thr Glu Gly Glu Val Pro Tyr Arg Glu Gly Gln Ser Ile Gly
1 5 10 15

Val Ile Pro Glu Gly Ile Asp Lys Asn Gly Lys Pro His Lys Leu Arg
20 25 30

Leu Tyr Ser Ile Ala Ser Ser Ala Ile Gly Asp Phe Gly Asp Ser Lys
35 40 45

Thr Val Ser Leu Cys Val Lys Arg Leu Val Tyr Thr Asn Asp Gly Gly
50 55 60

Glu Ile Val Lys Gly Val Cys Ser Asn Phe Leu Cys Asp Leu Lys Pro
65 70 75 80

Gly Asp Glu Ala Lys Ile Thr Gly Pro Val Gly Lys Glu Met Leu Met
85 90 95

Pro Lys Asp Pro Asn Ala Thr Ile Ile Met Leu Gly Thr Gly Thr Gly
100 105 110

Ile Ala Pro Phe Arg Ser Phe Leu Trp Lys Met Phe Phe Glu Glu His
115 120 125

Glu Asp Tyr Lys Phe Asn Gly Leu Ala Trp Leu Phe Leu Gly Val Pro
130 135 140

Thr Ser Ser Leu Leu Tyr Lys Glu Glu Phe Glu Lys Met Lys Glu
145 150 155 160

Lys Asn Pro Asp Asn Phe Arg Leu Asp Phe Ala Val Ser Arg Glu Gln
165 170 175

Thr Asn Glu Lys Gly Glu Lys Met Tyr Ile Gln Thr Arg Met Ala Glu
180 185 190

Tyr Ala Glu Leu Trp Glu Leu Leu Lys Lys Asp Asn Thr Phe Val
195 200 205

Tyr Met Cys Gly Leu Lys Gly Met Glu Lys Gly Ile Asp Asp Ile Met
210 215 220

Val Ser Leu Ala Ala Lys Asp Gly Ile Asp Trp Leu Glu Tyr Lys Lys
225 230 235 240

Gln Leu Lys Arg Ser Glu Gln Trp Asn Val Glu Val Tyr
245 250

<210> SEQ_ID NO 97

<211> LENGTH: 294

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana (thale cress)

<220> FEATURE:

US 11,001,853 B2

245**246**

-continued

<221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: ferredoxin-NADP(+)oxidoreductase(FNR2)

<400> SEQUENCE: 97

```

Met Ala Thr Thr Met Asn Ala Ala Val Ser Leu Thr Ser Ser Asn Ser
1           5          10          15

Ser Ser Phe Pro Ala Thr Ser Cys Ala Ile Ala Pro Glu Arg Ile Arg
20          25          30

Phe Thr Lys Gly Ala Phe Tyr Tyr Lys Ser Asn Asn Val Val Thr Gly
35          40          45

Lys Arg Val Phe Ser Ile Lys Ala Gln Ile Thr Thr Glu Thr Asp Thr
50          55          60

Pro Thr Pro Ala Lys Lys Val Glu Lys Val Ser Lys Lys Asn Glu Glu
65          70          75          80

Gly Val Ile Val Asn Arg Tyr Arg Pro Lys Glu Pro Tyr Thr Gly Lys
85          90          95

Cys Leu Leu Asn Thr Lys Ile Thr Ala Asp Asp Ala Pro Gly Glu Thr
100         105         110

Trp His Met Val Phe Ser His Gln Gly Glu Ile Pro Tyr Arg Glu Gly
115         120         125

Gln Ser Val Gly Val Ile Ala Asp Gly Ile Asp Lys Asn Gly Lys Pro
130         135         140

His Lys Val Arg Leu Tyr Ser Ile Ala Ser Ser Ala Leu Gly Asp Leu
145         150         155         160

Gly Asn Ser Glu Thr Val Ser Leu Cys Val Lys Arg Leu Val Tyr Thr
165         170         175

Asn Asp Gln Gly Glu Thr Val Lys Gly Val Cys Ser Asn Phe Leu Cys
180         185         190

Asp Leu Ala Pro Gly Ser Asp Val Lys Leu Thr Gly Pro Val Gly Lys
195         200         205

Glu Met Leu Met Pro Lys Asp Pro Asn Ala Thr Val Ile Met Leu Ala
210         215         220

Thr Gly Thr Gly Ile Ala Pro Phe Arg Ser Phe Leu Trp Lys Met Phe
225         230         235         240

Phe Glu Lys His Asp Asp Tyr Lys Phe Asn Gly Leu Ala Trp Leu Phe
245         250         255

Leu Gly Val Pro Thr Thr Ser Ser Leu Leu Tyr Gln Glu Glu Phe Asp
260         265         270

Lys Met Lys Ala Lys Ala Pro Glu Asn Phe Arg Val Asp Tyr Ala Ile
275         280         285

Ser Arg Glu Gln Ala Asn
290

```

<210> SEQ ID NO 98
 <211> LENGTH: 249
 <212> TYPE: PRT
 <213> ORGANISM: Proteobacteria

<400> SEQUENCE: 98

```

Met Lys Leu Leu Ile Leu Gly Ser Val Ile Ala Leu Pro Thr Phe
1           5          10          15

Ala Ala Gly Gly Gly Asp Leu Asp Ala Ser Asp Tyr Thr Gly Val Ser
20          25          30

Phe Trp Leu Val Thr Ala Ala Leu Ala Ser Thr Val Phe Phe Phe
35          40          45

```

US 11,001,853 B2

247

248

-continued

Val	Glu	Arg	Asp	Arg	Val	Ser	Ala	Lys	Trp	Lys	Thr	Ser	Leu	Thr	Val
50					55				60						
Ser Gly Leu Val Thr Gly Ile Ala Phe Trp His Tyr Met Tyr Met Arg															
65					70				75				80		
Gly Val Trp Ile Glu Thr Gly Asp Ser Pro Thr Val Phe Arg Tyr Ile															
85					90				95						
Asp Trp Leu Leu Thr Val Pro Leu Ile Cys Glu Phe Tyr Leu Ile															
100					105				110						
Leu Ala Ala Ala Thr Asn Val Ala Gly Ser Leu Phe Lys Lys Leu Leu															
115					120				125						
Val Gly Ser Leu Val Met Leu Val Phe Gly Tyr Met Gly Glu Ala Gly															
130					135				140						
Ile Met Ala Ala Trp Pro Ala Phe Ile Ile Gly Cys Leu Ala Trp Val															
145					150				155				160		
Tyr Met Ile Tyr Glu Leu Trp Ala Gly Glu Gly Lys Ser Ala Cys Asn															
165					170				175						
Thr Ala Ser Pro Ala Val Gln Ser Ala Tyr Asn Thr Met Met Tyr Ile															
180					185				190						
Ile Ile Phe Gly Trp Ala Ile Tyr Pro Val Gly Tyr Phe Thr Gly Tyr															
195					200				205						
Leu Met Gly Asp Gly Gly Ser Ala Leu Asn Leu Asn Leu Ile Tyr Asn															
210					215				220						
Leu Ala Asp Phe Val Asn Lys Ile Leu Phe Gly Leu Ile Ile Trp Asn															
225					230				235				240		
Val Ala Val Lys Glu Ser Ser Asn Ala															
245															

<210> SEQ ID NO 99
<211> LENGTH: 446
<212> TYPE: DNA
<213> ORGANISM: Arabidopsis thaliana (thale cress)

<400> SEQUENCE: 99

atttcgaaag	agaatctcag	aaagatcaat	ctagagagac	ccgttcgtct	ccttcctta	60
agccattacc	tctgaaacca	tccaaggctt	tggttgcAAC	tggaggcaga	gcacagaggc	120
ttcaagttaa	ggccctcaag	atggacaagg	cttgcACGG	tatctccgcg	gctgtctta	180
ctgcttcgat	ggtgattccg	gagatAGCTG	aagctgtgg	ttctggaatc	tctcTTCCC	240
tcaagaATTt	cttgctcAGC	attgcttctg	gtggcCTCGT	cctcaCTGTc	atcattggTG	300
tcgtcgTCGG	cgtctccAAC	tttgaccCTG	tcaAGAGAAC	ctaAGACCTA	tataCTTTC	360
ttacatcatt	attgtaatct	gttctccTTc	tgtgtattcg	tttcaatgtt	gcagcaatga	420
actttttggat	aaaaaaaaaa	aaaaaaa				446

<210> SEQ ID NO 100
<211> LENGTH: 642
<212> TYPE: DNA
<213> ORGANISM: Mus musculus

<400> SEQUENCE: 100

aaggcagaag	caccggTCAG	ctggggGAAG	ggacacAGAG	gaAGAGACGG	agtgtacagg	60
gaccaaggTT	gtatgtcAG	gagcaAGAG	caggaAGACA	ggaggCTTG	agcacacACG	120
gctttgtcta	ttccagtaac	aaccccTTG	ctgccgCTCA	ccggTTCCAT	ggagataATA	180
tttggccaga	ataagaaAGA	acagctggAG	ccagttcAGG	ccaaAGTGAC	aggcAGCATT	240

US 11,001,853 B2

249

-continued

250

```

ccagcatggc tgcaggggac cctgctccga aacgggcccggatgcacac agtgggagag 300
agcaagtaca accatgggtt tgatggctcgcccttc acagtttctc catcagagat 360
ggggaggtct tctacaggag caaatacctg cagagtgaca cctacatcgc caacatttag 420
gccaacagaa tcgtgggtc tgagttcgga accatggcct acccggaccc ctgaaaaaac 480
atctttcca aagctttctc ctatgtct cacaccatcc ccgacttcac agacaactgt 540
ctgatcaaca tcatgaaatg tggagaagac ttctatgcaa ccacggagac caactacatc 600
aggaaaatcg acccccagac cctagagacc ttggagaagg tg 642

```

<210> SEQ ID NO 101

<211> LENGTH: 217

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: (thale cress)

<400> SEQUENCE: 101

```

Met Ala Ser Leu Ser Thr Ile Thr Gln Pro Ser Leu Val His Ile Pro
1           5          10          15

```

```

Gly Glu Ser Val Leu His His Val Pro Ser Thr Cys Ser Phe Pro Trp
20          25          30

```

```

Lys Pro Thr Ile Asn Thr Lys Arg Ile Ile Cys Ser Pro Ala Arg Asn
35          40          45

```

```

Ser Ser Glu Val Ser Ala Glu Ala Glu Thr Glu Gly Gly Ser Ser Thr
50          55          60

```

```

Ala Val Asp Glu Ala Pro Lys Glu Ser Pro Ser Leu Ile Ser Ala Leu
65          70          75          80

```

```

Asn Val Glu Arg Ala Leu Arg Gly Leu Pro Ile Thr Asp Val Asp His
85          90          95

```

```

Tyr Gly Arg Leu Gly Ile Phe Arg Asn Cys Ser Tyr Asp Gln Val Thr
100         105         110

```

```

Ile Gly Tyr Lys Glu Arg Val Lys Glu Leu Lys Glu Gln Gly Leu Asp
115         120         125

```

```

Glu Glu Gln Leu Lys Thr Lys Met Asp Leu Ile Lys Ser Tyr Thr Ile
130         135         140

```

```

Leu Ser Thr Val Glu Glu Arg Arg Met Tyr Asp Trp Ser Leu Ala Arg
145         150         155         160

```

```

Ser Glu Lys Ala Glu Arg Tyr Val Trp Pro Phe Glu Val Asp Ile Met
165         170         175

```

```

Glu Pro Ser Arg Glu Glu Pro Pro Gln Glu Pro Glu Asp Val Gly
180         185         190

```

```

Pro Thr Arg Ile Leu Gly Tyr Phe Ile Gly Ala Trp Leu Val Leu Gly
195         200         205

```

```

Val Ala Leu Ser Val Ala Phe Asn Arg
210         215

```

<210> SEQ ID NO 102

<211> LENGTH: 53

<212> TYPE: PRT

<213> ORGANISM: Cyanophora paradoxa

<400> SEQUENCE: 102

```

Met Asn Ala Phe Val Ala Ser Val Ala Pro Ile Ala Val Ala Gly Ser
1           5          10          15

```

```

Ala Thr Leu Ser Ser Ala Val Cys Ala Gln Lys Lys Ala Phe Phe Gly

```

US 11,001,853 B2

251**252**

-continued

20

25

30

Ala Gln Val Ala Ala Lys Lys Thr Thr Phe Glu Ala Ala Pro Ala Arg
 35 40 45

Phe Ile Val Arg Ala
 50

<210> SEQ ID NO 103
<211> LENGTH: 45
<212> TYPE: PRT
<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 103

Met Ala Thr Gln Ala Ala Gly Ile Phe Asn Ser Ala Ile Thr Thr Ala
 1 5 10 15

Ala Thr Ser Gly Val Lys Lys Leu His Phe Phe Ser Thr Thr His Arg
 20 25 30

Pro Lys Ser Leu Ser Phe Thr Lys Thr Ala Ile Arg Ala
 35 40 45

<210> SEQ ID NO 104
<211> LENGTH: 30
<212> TYPE: PRT
<213> ORGANISM: Arabidopsis thaliana
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: (thale cress)
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: CAB transit peptide(thale cress)

<400> SEQUENCE: 104

Met Gln Ser Ser Ala Val Phe Ser Leu Ser Pro Ser Leu Pro Leu Leu
 1 5 10 15

Lys Pro Arg Arg Leu Ser Leu Arg His His Pro Ile Thr Thr
 20 25 30

<210> SEQ ID NO 105
<211> LENGTH: 44
<212> TYPE: PRT
<213> ORGANISM: Arabidopsis thaliana
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: (thale cress)
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: PGR5 transit peptide(thale cress)

<400> SEQUENCE: 105

Met Ala Ala Ala Ser Ile Ser Ala Ile Gly Cys Asn Gln Thr Leu Ile
 1 5 10 15

Gly Thr Ser Phe Tyr Gly Gly Trp Gly Ser Ser Ile Ser Gly Glu Asp
 20 25 30

Tyr Gln Thr Met Leu Ser Lys Thr Val Ala Pro Pro
 35 40

<210> SEQ ID NO 106
<211> LENGTH: 2955
<212> TYPE: DNA
<213> ORGANISM: Arabidopsis thaliana
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: PCRL1 gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: PCRL1 gene(thale cress)

-continued

<400> SEQUENCE: 106

catatttgat	tttcacatgg	attaacgaaa	cstatattatg	gaacacatc	aaaattataa	60
caacaaaaaa	aatacaagta	ttatattcaa	actacacaag	gttgtgctt	tttcttgaat	120
tatTTTactt	tcctaatgag	agcaaagt	tttcaagaag	taatcatatg	atgttttct	180
ttgaatgtgc	ctcacactta	cttacaaaca	caacacaagc	caatgagagc	tacatgaaaa	240
gatctgaaga	ttatacaaaa	cagcatacaa	actttggttt	ttctcccttct	tcttcaattt	300
ctccaccc	ttcatttgtt	agtattaatt	ttacatacac	ttctacataaa	ccctgagaaa	360
aagaaaaacc	taaaattttg	aattttccat	tgaatcaaga	aagatttcat	cagaaatcaa	420
agttgagata	agaattaaac	cttggctt	agatttaagc	tttccctcct	tctggtaatg	480
tgatcaaacg	agaacctgag	tcatalogca	tctccgttcc	acagctaaaa	accagaagaa	540
tcataagact	tcaagaaacg	ttttagacaa	tttgggtgat	cgattcgagt	ctacagctga	600
gaagcttacc	otgagcattt	gacatttttg	gtgttactat	cattggggat	ggatagtatc	660
gttccaaaga	aagatacatt	ctctgttcca	cagtttgggc	aagggcctg	tgaaagatata	720
gttccacgaa	aattaaaagc	atttcataat	aatcgatata	aactcgtagg	atttggcact	780
ataccaatcc	aaattttag	cgttttagc	aaaatagatt	attatctaa	gtctaatttc	840
ttgtttaagc	atttttgata	ctgagaaaac	aagattttagt	tctataactt	tttattttcc	900
acttcatgaa	ctgatcttgg	aagatgatta	atgttttac	cttcaagatc	aagaagtctt	960
tgaggatcg	tttggtgagt	gataacgcca	gatatacaat	tgcaggcacc	gcagegaacc	1020
atgtgaaat	gaaactgaat	ggttccggaa	gctgcagatt	tttgggggg	tttttaatc	1080
agttgcata	atactggAAC	aattactacg	agtatataatt	ctccaaacca	tagtagaata	1140
gtcgaaagag	gttttacctc	gagcaggtac	gtgatttcaa	aaccagtaat	gtcatcaaga	1200
aagaaaaatc	tggagaacaa	ttgagaaaa	caacaattt	aaactaacta	acataagcta	1260
agatcatgt	atttggaaat	tgagagagga	acagaaccgg	aggtactcac	agtccgagag	1320
caacaacagt	tgctggtaca	ttcaacaaga	acattttgaa	gtaatcaatg	gcaagatcac	1380
tataaacctg	cataatcaag	gaggccaca	agtctataac	atctctagag	tttgcctcaa	1440
acaatgaatc	taatgttatg	ttctgtatg	tccaaagaat	atatgagcta	tccgaacaat	1500
taagagttt	tacctttta	ctacgaagac	tgcatttgg	accctcacac	acaatctcac	1560
tgccgtccat	ctacagaaac	caaagaaaca	atcataacgt	ttgtccaaat	tacacatgt	1620
acaagatgga	tgaaaactaag	aaatagtatt	tgtaagtata	aatagattaa	gaacctttag	1680
tttcattttg	agcttatcat	actcttcatc	actcaagatt	ggatttccag	agacataagc	1740
cattgaagct	tcaaggaaatc	tttggttcatc	agaacctaca	atacatagat	aaaatttagat	1800
caagaatcaa	gaaccttaggc	gaatggatta	ttgacaaaac	tataaatcat	aagtgttcat	1860
tacttagcat	gacaacactg	cttccttccc	acatcaactc	ttctttaagg	ttatcaaact	1920
cttcatttga	cataatcgct	ttgccttcgt	aataaaacga	ctgcaaaaga	aaagaaacag	1980
aaacaatcc	cgatttata	gagataaaacc	catactaattg	ataaaaaacac	tttatttgat	2040
gtgttacttg	catcgcttgg	aggaactctt	gttccatttc	accgatagtt	ctcttctcat	2100
tcttggat	gctacaataa	ggtaaaatct	tgctatcaac	ttcttcccc	cccacctgac	2160
ctgaagacaa	gtcataaaaa	tgatTTTaa	aagtaaggaa	actctcaagg	agcaatctt	2220
tagtggatta	gagtataaaa	actaaaaatc	cacagaggaa	aaaagttcca	tataacaact	2280

US 11,001,853 B2

255**256**

-continued

tttcttaact agaattaaag cttgagtat tttattctat gattgaataa aatcaaact	2340
ttctcaaaag ccactgtgtt cccaaacaat gatcagagac aaaatcaaag ctacaataca	2400
acagctttc tcaactaaat ttgaagattg agtgctttt tgttcgatc acataacgt	2460
gagttataaa cttagaacc ttaagctaca cacaatttt aatcctaaaa aggctacaaa	2520
ttggaaatca tttatcta atatcttat gatcataaaa atctcaactt ttcacacca	2580
ttcgttccc aaagaaagat cagaggcaa aacaataaa aaaatcgaaa cttaaagag	2640
gcaaataaaa atcgagacct gattgatcg tagaagctt aaggggcaat aaggtaaatc	2700
ttcgtcttag agaaatcgat cgatccatggg taaagggagc aggacactgt gtcctcgaa	2760
aagaacaagt gatgggtttg cgagaaattt cagaaaatctt agggattgtt agagtaaaag	2820
ccatcgctt tatccctcac gccatgatt gagttagatc gttgttttcttgcggg	2880
acgaagaaca aaaaaaaaaa tttagaagctt tggatttgtt tggttgagaa ttgagatgg	2940
gatgtttttt actgt	2955

<210> SEQ_ID NO 107

<211> LENGTH: 55

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana (thale cress)

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: Rubisco

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: Rubisco(thale cress)

<400> SEQUENCE: 107

Met Ala Ser Ser Met Leu Ser Ser Ala Thr Met Val Ala Ser Pro Ala			
1	5	10	15

Gln Ala Thr Met Val Ala Pro Phe Asn Gly Leu Lys Ser Ser Ala Ala		
20	25	30

Phe Pro Ala Thr Arg Lys Ala Asn Asn Asp Ile Thr Ser Ile Thr Ser		
35	40	45

Asn Gly Gly Arg Val Asn Cys	
50	55

<210> SEQ_ID NO 108

<211> LENGTH: 795

<212> TYPE: DNA

<213> ORGANISM: homo sapien

<400> SEQUENCE: 108

gaattcatgt ctcatcattt gggttatggt aaacacaatg gtcctgaaca ctggcataaa	60
gactttccaa ttgcaaaagg tgaacgtcaa tcacctgttg atattgacac tcatacagct	120
aaatatgacc cttctttaaa accattatct gtttcatatg atcaagcaac ttctttacgt	180
atttaaaca atggcatgc tttaatgtt gaatttgcgtt actctcaaga taaaggatgt	240
ttaaaaagggtt gtccattttaga tggactttt cgttttattt aatttcattt tcactgggt	300
tcatttagat gtcaagggtt agaacataact gtagataaaa aaaaatatgc tgcagaatta	360
cacttagttt actggAACAC AAAATATGGT GATTTGGTAAAGCTGTACA ACAACCTGAT	420
ggtttagctt ttttaggtat ttttttttttt gttggtagtg ctaaaccagg tcttcaaaaa	480
gttggtagtttattttttttttt aattttttttttt aatggatggaa gtgtgtactt tactaatttc	540
gttcctcggtt gtttacttttcc tgaatcttta gattactgg catatccagg ttcatttaca	600
acaccccttc ttttagaatg tgtaacatgg attgtattaa aagaaccaat tagtgtaaat	660

-continued

agtgaacaag tattaaaatt ccgtaaactt aatttcaatg gtgaaggta accagaagaa	720
ttaatggttg ataaactggcg tccagctaa ccataaaaaa atcgtcaa ataaagttca	780
ttcaaataag catgc	795

What is claimed is:

1. A construct comprising:
 - i) a first heterologous nucleic acid sequence comprising a first heterologous polynucleotide sequence encoding a cyclic electron modulator gene wherein the cyclic electron modulator is operatively linked to at least one regulatory element wherein the first heterologous nucleic acid sequence encodes a protein having a sequence selected from PRG5 of SEQ ID NO: 1 or PGRL1 of SEQ ID NO: 3 or a sequence with at least 80% sequence homology thereto; and
 - ii) a second heterologous nucleic acid sequence comprising a second heterologous polynucleotide sequence encoding an ATP dependent bicarbonate anion transporter localized to the plasma membrane wherein the ATP dependent bicarbonate anion transporter localized to the plasma membrane is operatively coupled to the at least one regulatory element wherein the second heterologous nucleic acid sequence encodes a protein having a sequence selected from HLA3 (SEQ ID NO:77) or a sequence with at least 80% sequence homology thereto.
 2. The construct of claim 1 wherein the HLA3 is codon optimized for plant expression.
 3. The construct of claim 1 wherein the at least one regulatory element includes a promoter.
 4. The construct of claim 1 wherein the at least one regulatory element is a tissue specific promoter.
 5. The construct of claim 1 wherein the at least one regulatory element includes a promoter that is a green tissue/leaf-specific promoter.
 6. The construct of claim 1 wherein the promoter is selected from among CAB and rbcS.
 7. The construct of claim 1 wherein the first nucleic acid sequence and the second nucleic acid sequence encode:
 - i) the PGR5 protein, and the HLA3 protein; or
 - ii) the PGRL1 protein, and the HLA3 protein.
 8. The construct of claim 1 further comprising iii) a third heterologous nucleic acid sequence comprising a third heterologous polynucleotide sequence encoding a bicarbonate anion transporter protein localized to the chloroplast envelope wherein the bicarbonate anion transporter protein localized to the chloroplast envelope is operatively coupled to the regulatory element wherein the third heterologous nucleic acid sequence i-encodes a sequence selected from LCIA of SEQ ID NO: 18 or a sequence with at least 80% sequence homology thereto.
 9. The construct of claim 8 wherein the at least one regulatory element is a green tissue/leaf-specific promoter.
- 10 10. The construct of claim 8 wherein the at least one regulatory element includes a promoter selected from among CAB and rbcS.
- 15 11. The construct of claim 1 further comprising iii) a third heterologous nucleic acid sequence comprising a third heterologous polynucleotide sequence encoding a carbonic anhydrase wherein the carbonic anhydrase is operatively coupled to the at least one regulatory element wherein the third heterologous nucleic acid sequence encodes a protein selected from a human carbonic anhydrase-2 (HCA2) of SEQ ID NO: 18, bacterial *Neisseria gonorrhoeae* carbonic anhydrase (BCA) of SEQ ID NO: 5 or a sequence with at least 80% sequence homology thereto.
- 20 12. The construct of claim 11 wherein the at least one regulatory element includes a green tissue/leaf-specific promoter.
- 25 13. The construct of claim 11 wherein the at least one regulatory element includes a promoter selected from among CAB and rbcS.
- 30 14. The construct of claim 11 wherein the third heterologous nucleic acid sequence encodes the BCA protein of SEQ ID NO: 5 protein or a sequence with at least 80% sequence homology thereto.
- 35 15. The construct of claim 8 wherein the heterologous nucleotide sequences encode the PGR5 protein, the HLA3 protein, and the LCIA protein or sequences with at least 80% homology thereto.
- 40 16. The construct of claim 11 wherein the heterologous nucleotide sequences encode the PGR5 protein, the HLA3 protein, and the BCA protein or sequences with at least 80% homology thereto.
- 45 17. The construct of claim 11, wherein
 - a) the PGR5 protein has an amino acid sequence at least 80% identical to SEQ ID NO:1;
 - b) the HLA 3 protein has an amino acid sequence at least 80% identical to SEQ ID NO:77; and
 - c) the BCA protein has an amino acid sequence at least 80% identical to SEQ ID NO:21.
- 50 18. The construct of claim 1 wherein the at least one regulatory element of the first heterologous nucleic acid sequence and the at least one regulatory element of the second heterologous nucleic acid sequence includes a promoter which can be the same or different for the first heterologous nucleic acid and the second heterologous nucleic acid.
- 55 19. A seed comprising the construct of claim 1.
20. A vector comprising the construct of claim 1.

* * * * *